

Viral Ateşlerin Tanısında Sağlık Bakanlığı Perspektifi ve Ülkemizde Son Beş Yılın Epidemiyolojisi

Uzm. Dr. Dilek Yağcı Çağlayık
Türkiye Halk Sağlığı Kurumu
Ulusal Arbovirus ve Viral Zoonotik Hastalıklar Ünitesi

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Kurum Başkanı

Hukuk Müşavirliği

Denetim Hizmetleri DB.

Strateji Geliştirme DB.

Birinci Basamak Sağlık Hizmetleri Başkan Yardımcılığı

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Bulaşıcı Olmayan Hastalıklar Programları ve Kansere Başkan Yardımcılığı

Tüketici ve Çalışan Güvenliği Başkan Yardımcılığı

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İstatistik ve Bilgi İşlem DB.

Mikrobiyoloji Referans Laboratuvarları DB.

Kadın ve Üreme Sağlığı DB.

Çocuk ve Ergen Sağlığı DB.



1-Viral Ateş Tanımı ve Etiyolojisi

Viral ateşler;

- Arbovirus (artropod kaynaklı), robovirus (rodent kaynaklı) ve diğer zoonotik virusların neden olduğu (yarasa, maymun)
- Ani başlangıçlı, multisistemik ve ölümcül seyirli olabilen hastalıklardır.
- Etkenler, RNA virusudur.
- Farklı coğrafi bölgelerde; farklı vektörlerin taşınması nedeniyle farklı etkenler fakat benzer klinik özellikler izlenmektedir.

Sunum Planı

- 1-Viral ateş tanımı ve etiyolojisi
- 2-Tanı yöntemleri ve algoritmalar
- 3-Son beş yılın vaka verileri
- 4-Tanısal kapasiteyi ve kaliteyi arttırmaya yönelik yapılanlar ve hedefler

1-Viral Ateş Tanımı ve Etiyolojisi

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AİLE

CİNS

TÜR

COĞRAFI DAĞILIM

VEKTÖR

*Bunyaviridae**Ortobunyavirus*

- Kaliforniya ensefalit virusu
- Oropouche ateşi

Amerika

Aedes triseriatus*Nairovirus*

KKKAV

Asya, Afrika, Doğu Avrupa

*Phlebovirus*

- Rift Vadisi Ateşi Virusu
- Tatarcık humması virusu

Afrika, Asya

Akdeniz havzası

Phlebotomus (tatarcık)

*Hantavirus*

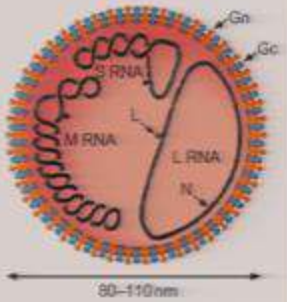
- Puumala
- Hantaan
- Dobrova, Seoul
- Sin Nombre, Andes, Tula

Avrupa, Amerika

Myodes glareolus, (NE)



Apodemus flavicollis (BSKA)

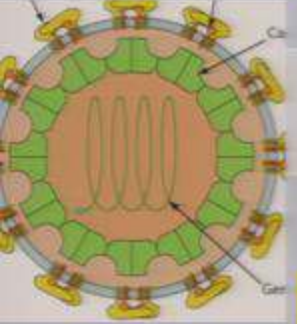


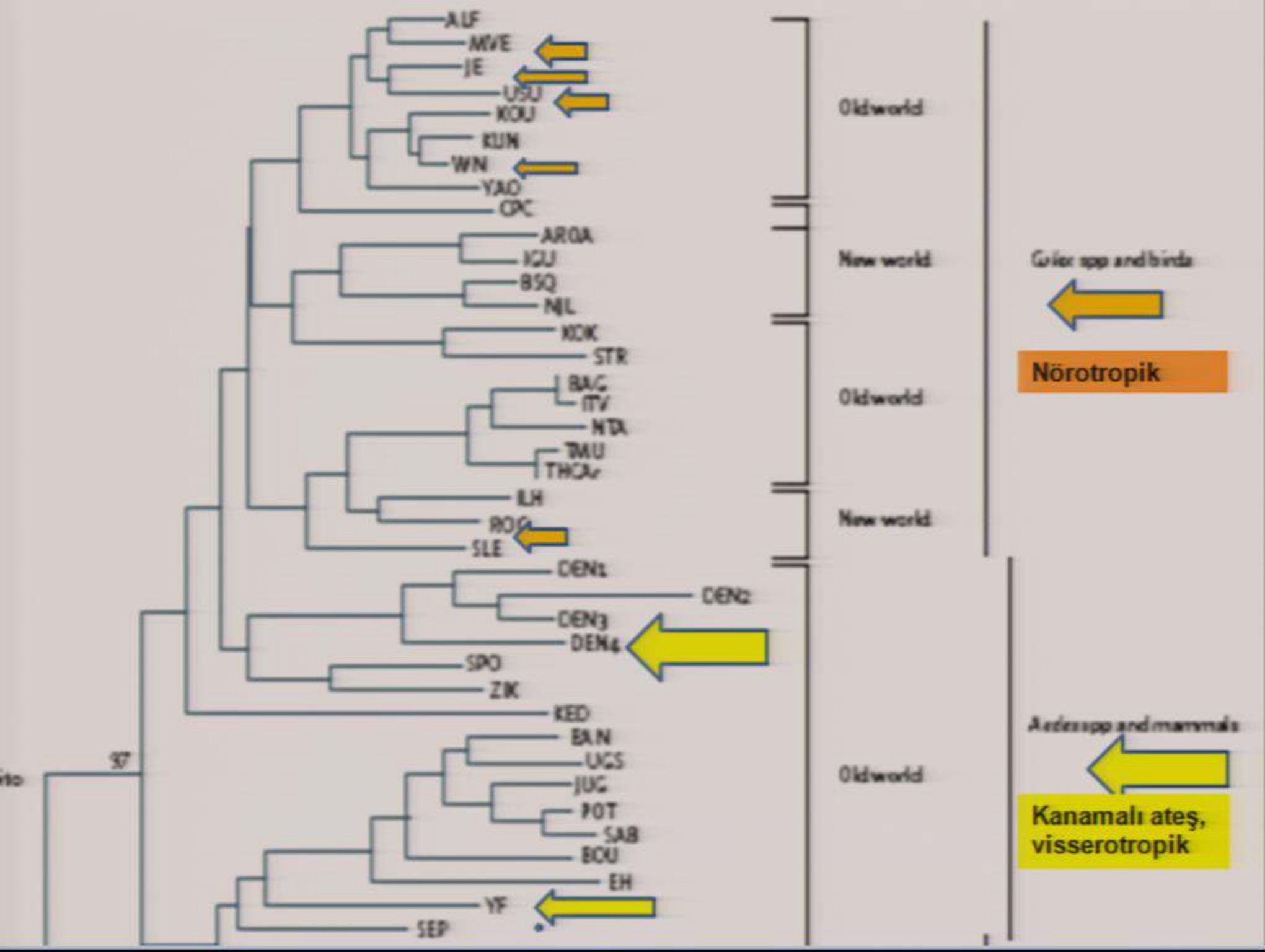
AİLE	CİNS	TÜR	COĞRAFI DAĞILIM	VEKTÖR
Flaviviridae	Flavivirus (Sivrisinek kaynaklı)	Japon ensefaliti virus grubu (Batı Nil virusu, Murray vadisi ensefalit virusu, Usutu virus, JEV)	Asya-Afrika-Avrupa, Avustralya, Afrika-Avrupa, Asya	Culex spp. Ochleratatus caspius
		Spondweni virus grubu (Zika virus)	Afrika, Asya	Aedes spp.
		Sarı humma virus grubu (Sarı humma virusu, Sepik virus)	Sahra altı Afrika ve Güney Amerika, Yeni Gine	Aedes spp.
		Deng virusu	Tropik ve subtropik yerler	Aedes aegypti Aedes albopictus
		Flavivirus (Kene kaynaklı)	Kene Kaynaklı Ensefalit virusu (TBE)	Avrupa, Asya
		Omsk kanamalı ateşi virusu	Batı Sibirya	Dermacentor
		Kyasanur ormanı hastalığı virusu, Alkhurma virusu	Hindistan, Suudi Arabistan	Haemaphysalis




Nörotropik, rezervuar kuşlar

Visserotropik, kanamalı ateş, rezervuar maymun ve insan

Rezervuar kemiriciler





AİLE	CİNS	TÜR	COĞRAFI DAĞILIM	VEKTÖR
<i>Togaviridae</i>	<i>Alphavirus</i>	<i>Chikungunya virus</i>	Güneydoğu Asya, Afrika	Aedes spp. 
		<i>Sindbis virus</i>	Kuzey Avrupa, Avustralya, Çin, Güney Afrika	Culex spp. 
<i>Arenaviridae</i>	<i>Arenavirus</i>	Lenfositik koryomenenjit virus	Avrupa, Amerika, Avustralya, Japonya	Ev faresi 
		Lassa virus	Batı Afrika	Sıçan
<i>Filoviridae</i>		Ebolavirus	Batı Afrika (Liberya, Gine, S. Leone)	Yarasa
		Marburgvirus	Kenya, Uganda, Angola	Maymun
				8

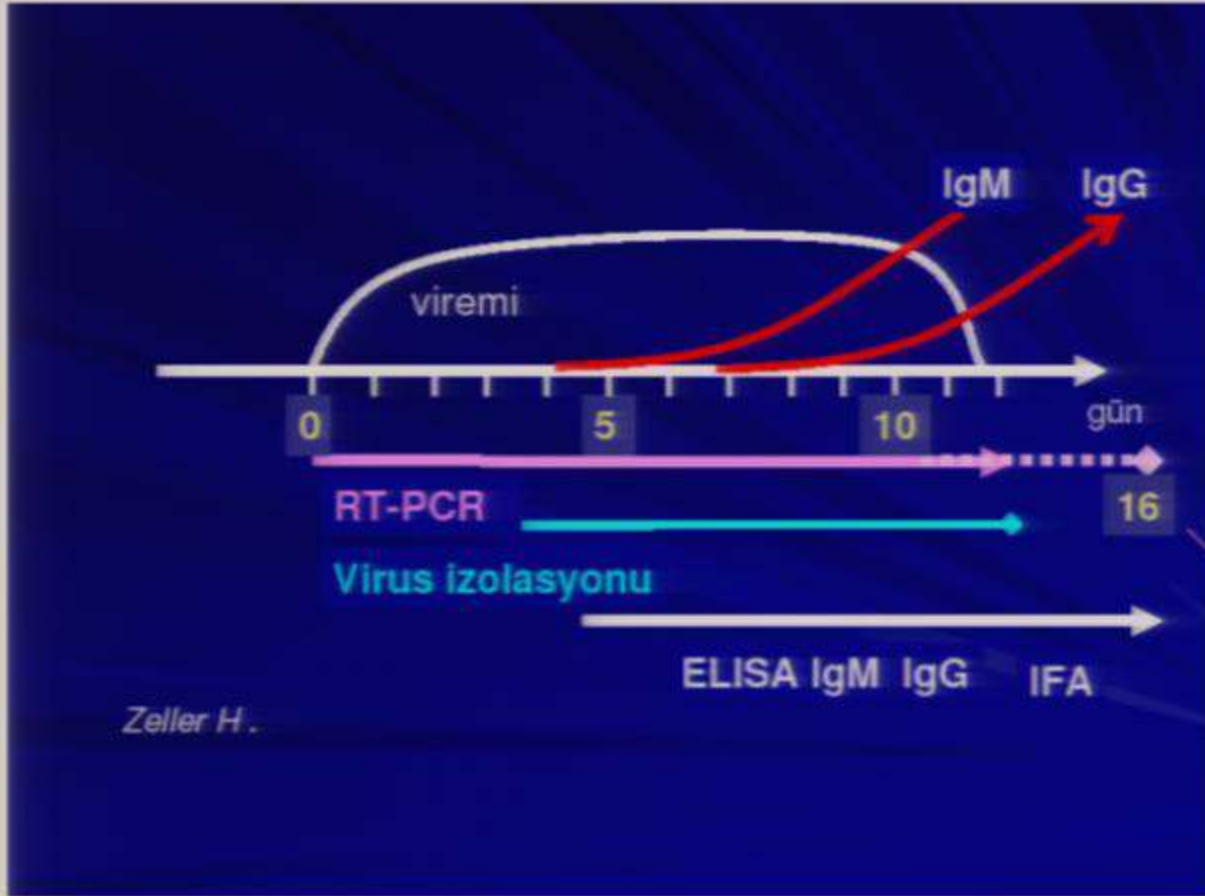
2-Tanı yöntemleri

Tanısı konabilen arboviral etkenler;

1. KKKA
2. Hantavirus
3. Tatarcık humması
4. Batı Nil virusu
5. Kene kaynaklı ensefalit virusu
6. Japon Ensefaliti virusu
7. Sarı humma virusu
8. Deng ateşi virusu
9. Chikungunya virusu



Kırım Kongo Kanamalı Ateşi Virüsü Tanı Algoritması



İlk örnekte rRT-PCR

rRT-PCR
negatif

rRT-PCR
pozitif (306 vaka)



IgM

İkinci örnekte IgM (104 vaka)

İstenildiğinde (adli
vaka vb. gibi) IgG



Kırım Kongo Kanamalı Ateşi Virüsü Tanı Algoritması-2

rRT-PCR (gerçek zamanlı ters transkriptaz polimeraz zincir reaksiyonu)

- Ticari Kit(Kantitatif)
- *in-house* (Yapar et-al, HPA Protokolleri-Kalitatif)

Serolojik Testler (IgM,IgG)

- ELISA (CDC protokolü)
- IFA (Ticari)



2- Tanı yöntemleri

CDC ELISA protokolü;

- Tüm dünyada referans yöntem olarak kabul edilmektedir.
- Capture ELISA (sandviç/tuzak ELISA)..özgüllüğü yüksek

-antihuman IgM(kaplama-1 gece)

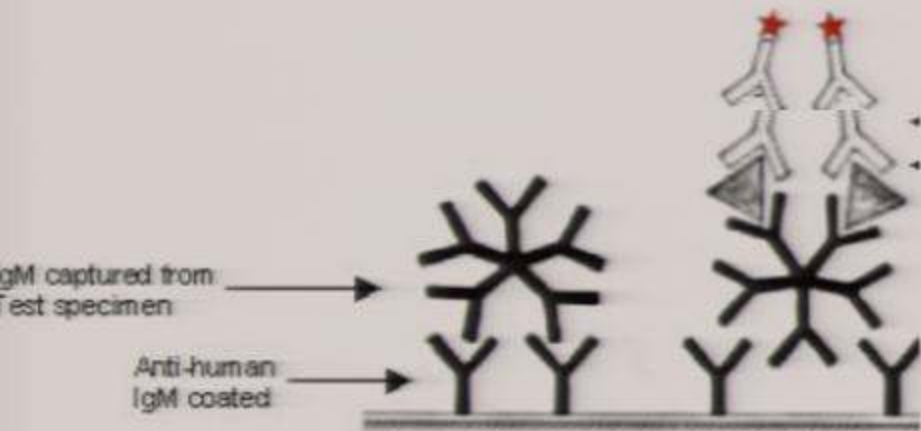
-hasta örneği (IgM)

-KKKA Antijeni (Pozitif ve negatif)

-HMAF (Hiperimmün mouse ascitic fluid)

-Konjugat (Antimouse IgM-HRP)

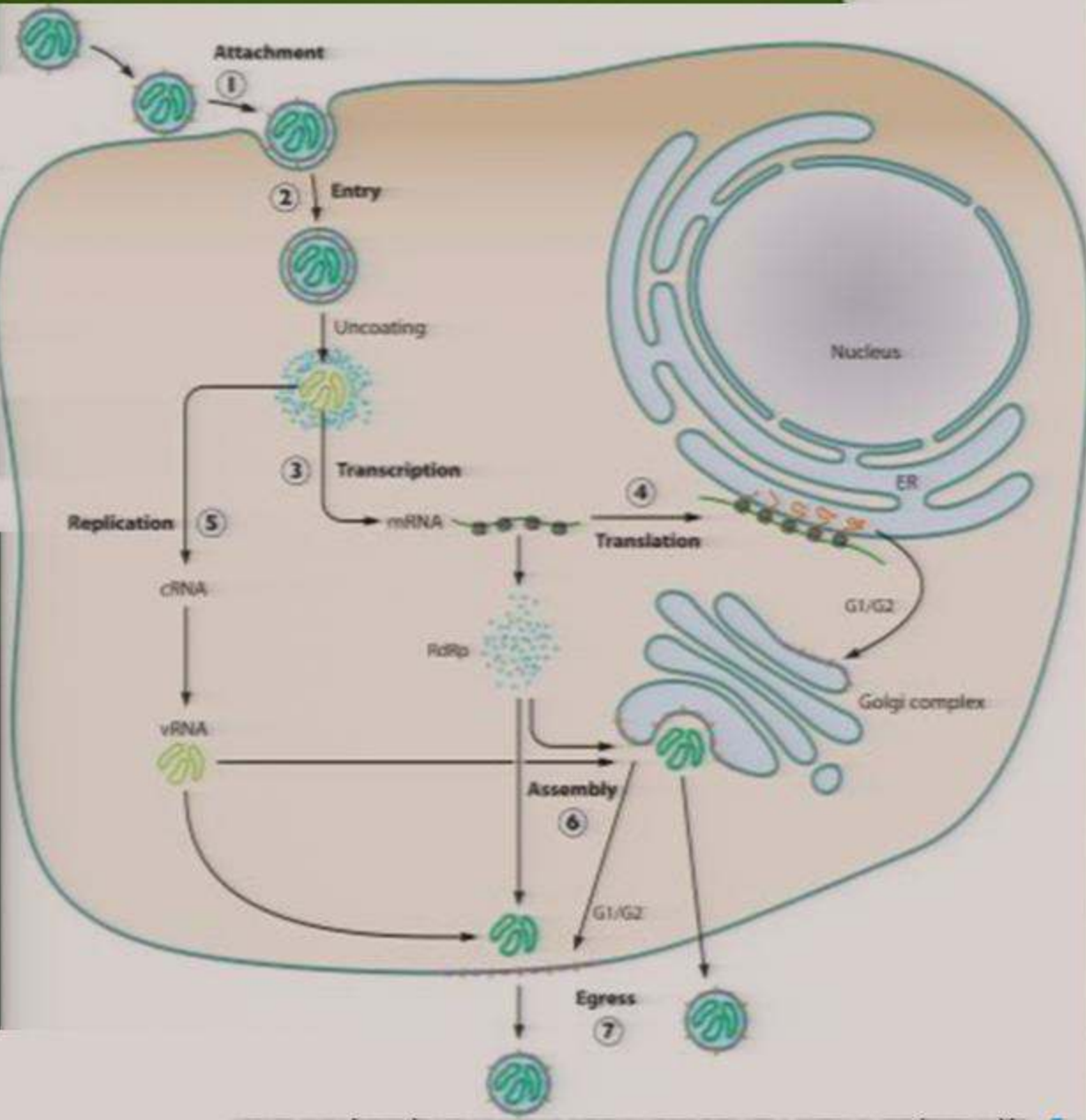
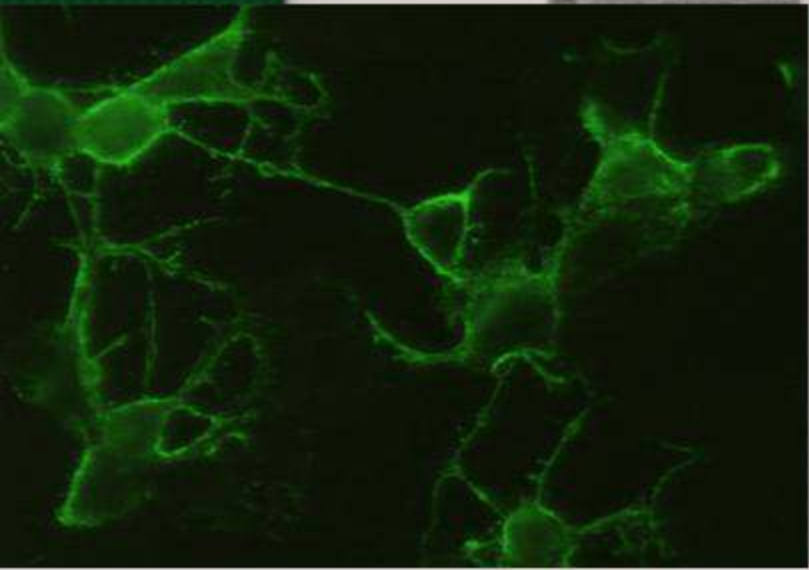
-Substrat



	01	02	03	04	05	06	07	08	09	10
A	0.112	0.084	0.074	0.074	0.071	0.074	0.074	0.074	0.074	0.074
B	0.084	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074
C	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074
D	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074
E	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074
F	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074
G	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074
H	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074	0.074



2- Tanı yöntemleri



2- Tanı yöntemleri

Ticari IFA sisteminin referans yöntem ELISA'ya göre duyarlılık ve özgüllüğü

Table 4. Overall performance of assays compared in study of CCHF diagnostic tools

Parameter	IgM serology		IgG serology		Genome detection	
	ELISA	IFA	ELISA	IFA	qRT-PCR	LCD array
No. samples tested	138	90	137	92	71	70
No. true positive	43	31	41	31	39	40
No. false negative	6	2	10	5	10	8
No. true negative	88	57	86	56	21	21
No. false positive	1	0	0	0	1	1
Sensitivity, % (95% CI)	87.8 (75.2–95.3)	93.9 (79.8–99.3)	80.4 (66.9–90.2)	86.1 (70.5–95.3)	79.6 (65.7–89.8)	83.3 (69.8–92.5)
Specificity, % (95% CI)	98.9 (93.9–100.0)	100.0 (93.7–100.0)†	100.0 (95.8–100.0)	100.0 (93.6–100.0)	95.5 (77.2–99.9)	95.5 (77.2–99.9)

*CCHF, Crimean-Congo hemorrhagic fever; IFA, immunofluorescent assay; qRT-PCR, quantitative reverse transcription PCR; LCD, low-cost, low-density.

†One-sided 95% CI.

to 100.0%, with an overall sensitivity of 80.4% (95% CI 69.5%–91.3%). For the IgG IFA, sensitivity ranged from 40.0% to 100.0%, with an overall sensitivity of 86.1% (95% CI 74.8%–97.4%). Specificity was estimated to be 100% for both assays (Table 4).

the requirement for specific technical training (0.3/1). The observed scores for molecular tests were within the same range (6.0–6.3/10). Both molecular assays demonstrated low scores for technical complexity (1.3–1.5/2) and training requirements for equipment and technique (0.3–0.5/1).



2- Tanı yöntemleri

- **ISO 15189 Akreditasyon çalışmaları**

(Uygunluğun ve geçerliliğin uluslar arası bilimsel kriterlere göre kanıtlanması çalışması)

KKKA rRT-PCR Metod Validasyon

(Yöntem geçerlilik) testleri gerçekleştirildi.

1. Doğruluk çalışması
2. Tekrarlanabilirlik çalışması
3. Özgüllük çalışması
4. Analitik duyarlılık çalışması

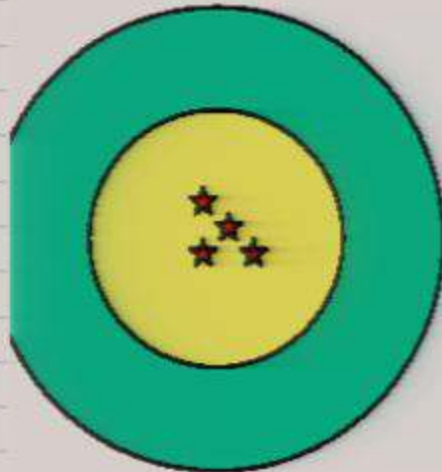


1-Doğruluk çalışması

- Referans materyal oluşturuldu.
- Serum havuzu (13 örnekten 5 paket)
- Ankara(AVZHÜ)
 - ↙
 - ↘ ↙ ↘
- Erzurum Samsun
- Dört (4) ayrı çalışma ile 13 örneğin sonucuyla ilgili olarak konsensusa varıldı.
- Doğruluk %100

2-Tekrarlanabilirlik çalışması (%CV) intra ve interassay (çalışma içi ve çalışmalar arası)

A	B	C	D	E	F	G	H	I	J	K	L	M	N
Referans Materyal (LAK Yeterlilik örnekleri): ct=19-20-21 olan yaklaşık 20 Pozitif hasta serumu+Dilusyonlar için KKKA PCR neg gönüllü sağlıklı serumu (17 Haziran 2013)													
Analist: Nilgün Gökalp							Analist: İhsan Durmaz						
Ref. Mat. No: Bulunan değer							Doğruluk %100						
	17.6.2013 (1)	18.6.2013 (1)	18.6.2013 (2)	YP; %6			19.6.2013 (2)	19.6.2013 (3)	20.6.2013 (1)				
1	7,78E+06	8,51E+06 8,22E+06 9,62E+06	8,93E+06				1	4,24E+06 6,96E+06 7,52E+06 6,34E+06	9,33E+06				
2	6,28E+05							2	6,55E+05				
3	1,18E+05							3	6,14E+04				
4	1,24E+04	1,90E+04 2,04E+04 1,31E+04	5,99E+03	DP; %20			4	1,92E+03 2,91E+03 1,18E+04 5,43E+03	5,28E+03				
(3X1/2)5	4,52E+04							5	3,03E+04				
(3X1/4)6	4,03E+04							6	1,34E+04				
(3X1/8)7	1,73E+04							7	MULTİ CT				
(1X1/2)8	2,95E+06							8	2,90E+06				
(1X1/4)9	2,10E+06							9	1,51E+06				
(1X1/8)10	9,50E+05							10	8,12E+05				
11	Negatif							11	Negatif				
12	Negatif							12	Negatif				
13	Negatif							13	Negatif				



3-Özgüllük çalışması

No	Yılı	Lab Protokol	Hasta adı	Pozitiflik	matrix	KKKA-PCR
1	2013	864	H.B.	HBV	plazma	negatif
2	2013	865	A.Ç.	HBV	plazma	negatif
3	2013	866	H.E.	HBV	plazma	negatif
4	2013	867	M.S.	HBV	plazma	negatif
5	2013	868	N.E.	HBV	plazma	negatif
6	2013	191	N.K.	HCV	plazma	negatif
7	2013	192	S.I.	HCV	plazma	negatif
8	2013	195	S.E.	HCV	plazma	negatif
9	2013	201	M.G.	HCV	plazma	negatif
10	2013	227	M.A.B.	HCV	plazma	negatif
11	2013	115	H.K.	HIV	plazma	negatif
12	2013	169	K.İ.	HIV	plazma	negatif
13	2013	173	PAALYU	HIV	plazma	negatif
14	2013	216	N.D.	HIV	plazma	negatif
15	2013	229	Z.K.	HIV	plazma	negatif
16	2013		Ü.	SFV	serum	negatif
17	2013		U.	SFV	serum	negatif
18	2013	3179	İtir Beril Y	Kızamık	Tam kan	negatif
19	2013		Rega	Coxiella	Tam kan	negatif
20	2013			Coxiella	Tam kan	negatif
21	2013			Brucella	Tam kan	negatif

Özgüllük %100

4-Analitik Duyarlılık çalışması

Referans Mat: 4 numaralı ref. Mat.serumun 1/2'lik 8 dilusyonu

predicted viral copy/mcl	Örn	Çalışma içi-18.06.2013(1)	Çalışmalar arası18.06.2013(2)-19.06.2013(1,2,3)-20.06.2013(1,2,3)-21.06.2013(1,2,3*100mcl*)											
11.800	4-0	1,66E+04	1,11E+04	1,57E+04	4,38E+03	1,18E+04	2,00E+03	3,44E+03	1,41E+04	4,92E+03	2,41E+03	2,25E+03	9,57E+02	7,03E+02
5900	4-1	2,16E+03	5,18E+03	5,53E+03	3,91E+03	4,11E+02	2,41E+03	5,06E+02	3,61E+03	1,93E+03	7,30E+02	1,08E+03	1,11E+03	9,78E+02
2950	4-2	1,39E+03	9,61E+02	9,71E+02	1,31E+03	8,84E+02	8,63E+02	6,64E+02	2,47E+03	6,48E+02	7,16E+00	5,32E+02	3,73E+02	4,51E+01
1475	4-3	1,46E+03	4,28E+01	6,31E+02	Negatif	Negatif	1,67E+02	Negatif	1,80E+03	5,55E+02	1,76E+02	Negatif	3,74E+02	Negatif
737	4-4	2,79E+02	1,15E+03	5,59E+02	3,37E+01	Negatif	Negatif	Negatif	5,28E+01	Negatif	8,04E+00	Negatif	2,50E+00	Negatif
368	4-5	Negatif	Negatif	1,20E+01	Negatif	Negatif	Negatif	Negatif	3,53E+02	Negatif	Negatif	Negatif	Negatif	3,36E+01
184	4-6	Negatif	Negatif	1,34E+01	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif
92	4-7	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif
46	4-8	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif

,550	1131,479	887,261	1565,707
,600	1214,836	958,202	1695,259
,650	1300,992	1029,333	1831,354
,700	1391,787	1102,424	1976,648
,750	1489,769	1179,641	2135,102
,800	1598,877	1264,077	2313,098
,850	1726,056	1360,945	2522,124
,900	1886,076	1481,093	2786,863
,910	1924,725	1509,886	2851,032
,920	1966,713	1541,081	2920,827
,930	2012,880	1575,290	2997,663
,940	2064,442	1613,390	3083,581
,950	2123,249	1656,722	3181,693
,960	2192,339	1707,483	3297,111
,970	2277,277	1769,694	3439,197
,980	2390,187	1852,107	3628,359
,990	2568,147	1981,463	3927,040

Probit Transformed Response



LOD(SS)mızı bulmak için dokuz (9) konsantrasyon için 13 çalışma yaptık.

%95 GA'da %95 olasılıkla ile saptama alt sınırimızın 1656 kopya/ml olduğunu gördük.

PCR ile saptama alt sınırı yüksek tespit edildiğinden, PCR negatif hastalarda tanısal algoritmamıza göre mutlaka IgM testi yapıyoruz.

Title:	Method validation of a Crimean-Congo haemorrhagic fever real-time RT-PCR assay
Programme No:	R314
Speaker:	Dilek Yagci Caglayik
Author(s):	Yagci Caglayik D.; Menemenoglu D.; Uyar Y.; Konukdoglu G.; Ozkutuk A.
Affiliation(s):	National Arbovirus and Viral Zoonoses Reference and Research Laboratory, Public Health Institute of Turkey (Ankara, Turkey); Medical Microbiology, Istanbul University Cerrahpasa Medical Faculty (Istanbul, Turkey); Virology, Public Health Institute of Turkey (Ankara, Turkey); Microbiology and Clinical Microbiology, Dokuz Eylul University Faculty of Medicine (Izmir, Turkey)
Subtopic:	Molecular biology, including diagnostics: Molecular virology
Topic:	Publication Only

24th

ECCMIDBarcelona, Spain
10 – 13 May 2014ESCMID EUROPEAN SOCIETY OF CLINICAL
MICROBIOLOGY AND INFECTIOUS DISEASES

Objectives

The aim of this study was to reveal the analytical performance features of a commercially available Altona marked Research Use Only qualitative Crimean Congo hemorrhagic fever virus (CCHFV) real time Reverse Transcription Polymerase Chain reaction (rt RT-PCR) test (Altona Diagnostics RealStar® CCHFV RT-PCR Kit 1.2, Germany).

Methods

In order to have a consensus reference material as no other means were achievable at that moment, 10 positive (low and high positive) and 3 negative serum pool samples were sent to two regional laboratories that are officially responsible from CCHFV diagnosis in Turkey and evaluated and confirmed for the results with PCR before the validation tests. Positive sera pools were prepared from 14 PCR positive patient sera and negative sera. Sera for dilutions were prepared from PCR negative CCHFV sera. All of the sera samples tested by three laboratories with PCR were found to be the same in this interlaboratory comparison hence the samples were accepted as reference material.

Validation experiments were performed to validate accuracy, precision, specificity and analytical sensitivity. Accuracy was determined with 3 positive, 3 low positive and 3 negative sera. For intra and interassay precision detections 1 positive and 1 low positive sera were used for each. Specificity was tested with 21 CCHFV-PCR negative but 5 HepatitisB, 5 HepatitisC, 5 HIV, 2 Sandfly fever virus, 2 *Coxiella burnetii*, 1 *Brucella* spp., 1 Measles PCR positive blood samples. Analytical sensitivity, also called limit of detection (LOD) test was done with serial 2 fold dilutions of known low positive reference material.

Probit model analysis (SPSS version 20) was used to detect the lowest viral copy that is positive in the 95% of the tests performed.

Results

Accuracy was detected as 100% with 13 reference sera samples. As CV% (coefficient of variance) values; interassay precision was detected as 6.9% for high positive samples and 23% for low positive samples. Intraassay precision was 8.41% and 22.1% for high and low positive samples, respectively. Specificity was found to be 100%, neither bacterial nor viral pathogens showed positivity for CCHFV. According to the probit model the limit of detection for this assay was 2123 copies/ml.

ISO 15189 belgesinin Analitik süreç ile ilgili gereklilikleri uyarınca; kullandığımız moleküler yöntem için uygunluk ve geçerliliği uluslar arası bilimsel kriterlere göre kanıtlamış olduk.



4-Analitik Duyarlılık çalışması

Referans Mat: 4 numaralı ref. Mat.serumun 1/2'lik 8 dilusyonu

predicted viral copy/mcl	Örn	Çalışma içi-18.06.2013(1)	Çalışmalar arası18.06.2013(2)-19.06.2013(1,2,3)-20.06.2013(1,2,3)-21.06.2013(1,2,3*100mcl*)												
11.800	4-0	1,66E+04	1,11E+04	1,57E+04	4,38E+03	1,18E+04	2,00E+03	3,44E+03	1,41E+04	4,92E+03	2,41E+03	2,25E+03	9,57E+02	7,03E+02	
5900	4-1	2,16E+03	5,18E+03	5,53E+03	3,91E+03	4,11E+02	2,41E+03	5,06E+02	3,61E+03	1,93E+03	7,30E+02	1,08E+03	1,11E+03	9,78E+02	
2950	4-2	1,39E+03	9,61E+02	9,71E+02	1,31E+03	8,84E+02	8,63E+02	6,64E+02	2,47E+03	6,48E+02	7,16E+00	5,32E+02	3,73E+02	4,51E+01	
1475	4-3	1,46E+03	4,28E+01	6,31E+02	Negatif	Negatif	1,67E+02	Negatif	1,80E+03	5,55E+02	1,76E+02	Negatif	3,74E+02	Negatif	
737	4-4	2,79E+02	1,15E+03	5,59E+02	3,37E+01	Negatif	Negatif	Negatif	5,28E+01	Negatif	8,04E+00	Negatif	2,50E+00	Negatif	
368	4-5	Negatif	Negatif	1,20E+01	Negatif	Negatif	Negatif	Negatif	3,53E+02	Negatif	Negatif	Negatif	Negatif	3,36E+01	
184	4-6	Negatif	Negatif	1,34E+01	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	
92	4-7	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	
46	4-8	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	Negatif	

,550	1131,479	887,261	1565,707
,600	1214,836	958,202	1695,259
,650	1300,992	1029,333	1831,354
,700	1391,787	1102,424	1976,648
,750	1489,769	1179,641	2135,102
,800	1598,877	1264,077	2313,098
,850	1726,056	1360,945	2522,124
,900	1886,076	1481,093	2786,863
,910	1924,725	1509,886	2851,032
,920	1966,713	1541,081	2920,827
,930	2012,880	1575,290	2997,663
,940	2064,442	1613,390	3083,581
,950	2123,249	1656,722	3181,693
,960	2192,339	1707,483	3297,111
,970	2277,277	1769,694	3439,197
,980	2390,187	1852,107	3628,359
,990	2568,147	1981,463	3927,040

Probit Transformed Response



LOD(SS)mızı bulmak için dokuz (9) konsantrasyon için 13 çalışma yaptık.

%95 GA'da %95 olasılıkla ile saptama alt sınırimızın 1656 kopya/ml olduğunu gördük.

PCR ile saptama alt sınırı yüksek tespit edildiğinden, PCR negatif hastalarda tanısal algoritmamıza göre mutlaka IgM testi yapıyoruz.

Title:	Method validation of a Crimean-Congo haemorrhagic fever real-time RT-PCR assay
Programme No:	R314
Speaker:	Dilek Yagci Caglayik
Author(s):	Yagci Caglayik D.; Menemenoglu D.; Uyar Y.; Konukdogru G.; Ozkutuk A.
Affiliation(s):	National Arbovirus and Viral Zoonoses Reference and Research Laboratory, Public Health Institute of Turkey (Ankara, Turkey); Medical Microbiology, Istanbul University Cerrahpasa Medical Faculty (Istanbul, Turkey); Virology, Public Health Institute of Turkey (Ankara, Turkey); Microbiology and Clinical Microbiology, Dokuz Eylul University Faculty of Medicine (Izmir, Turkey)
Subtopic:	Molecular biology, including diagnostics: Molecular virology
Topic:	Publication Only

24th

ECCMIDBarcelona, Spain
10 – 13 May 2014ESCMID EUROPEAN SOCIETY OF CLINICAL
MICROBIOLOGY AND INFECTIOUS DISEASES

Objectives

The aim of this study was to reveal the analytical performance features of a commercially available Altona marked Research Use Only qualitative Crimean Congo hemorrhagic fever virus (CCHFV) real time Reverse Transcription Polymerase Chain reaction (rt RT-PCR) test (Altona Diagnostics RealStar® CCHFV RT-PCR Kit 1.2, Germany).

Methods

In order to have a consensus reference material as no other means were achievable at that moment, 10 positive (low and high positive) and 3 negative serum pool samples were sent to two regional laboratories that are officially responsible from CCHFV diagnosis in Turkey and evaluated and confirmed for the results with PCR before the validation tests. Positive sera pools were prepared from 14 PCR positive patient sera and negative sera. Sera for dilutions were prepared from PCR negative CCHFV sera. All of the sera samples tested by three laboratories with PCR were found to be the same in this interlaboratory comparison hence the samples were accepted as reference material.

Validation experiments were performed to validate accuracy, precision, specificity and analytical sensitivity. Accuracy was determined with 3 positive, 3 low positive and 3 negative sera. For intra and interassay precision detections 1 positive and 1 low positive sera were used for each. Specificity was tested with 21 CCHFV-PCR negative but 5 Hepatitis B, 5 Hepatitis C, 5 HIV, 2 Sandfly fever virus, 2 *Coxiella burnetii*, 1 *Brucella* spp., 1 Measles PCR positive blood samples. Analytical sensitivity, also called limit of detection (LOD) test was done with serial 2 fold dilutions of known low positive reference material.

Probit model analysis (SPSS version 20) was used to detect the lowest viral copy that is positive in the 95% of the tests performed.

Results

Accuracy was detected as 100% with 13 reference sera samples. As CV% (coefficient of variance) values; interassay precision was detected as 6.9% for high positive samples and 23% for low positive samples. Intraassay precision was 8.41% and 22.1% for high and low positive samples, respectively. Specificity was found to be 100%, neither bacterial nor viral pathogens showed positivity for CCHFV. According to the probit model the limit of detection for this assay was 2123 copies/ml.

ISO 15189 belgesinin Analitik süreç ile ilgili gereklilikleri uyarınca; kullandığımız moleküler yöntem için uygunluk ve geçerliliği uluslar arası bilimsel kriterlere göre kanıtlamış olduk.



Hantavirus Tanısı

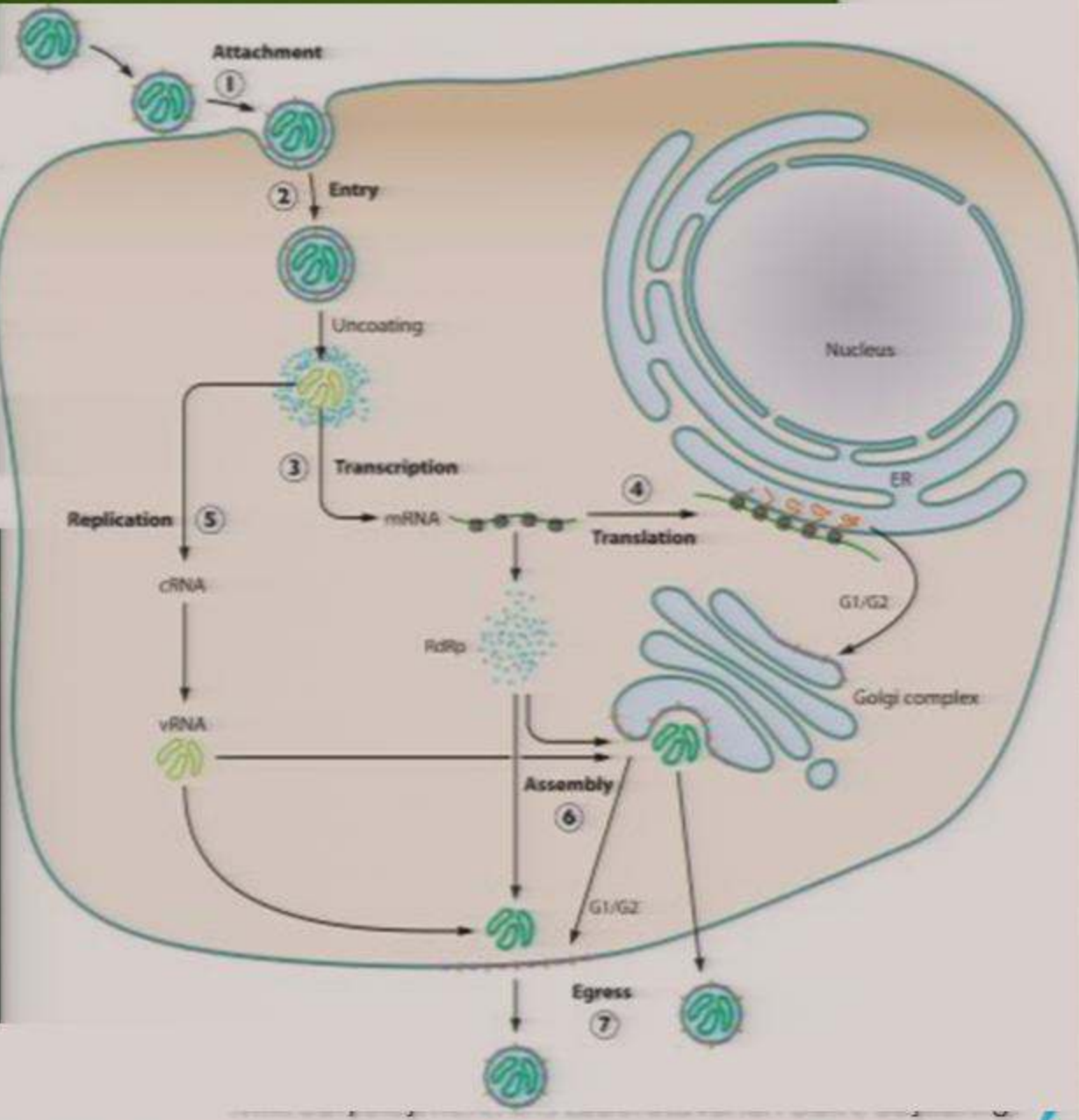
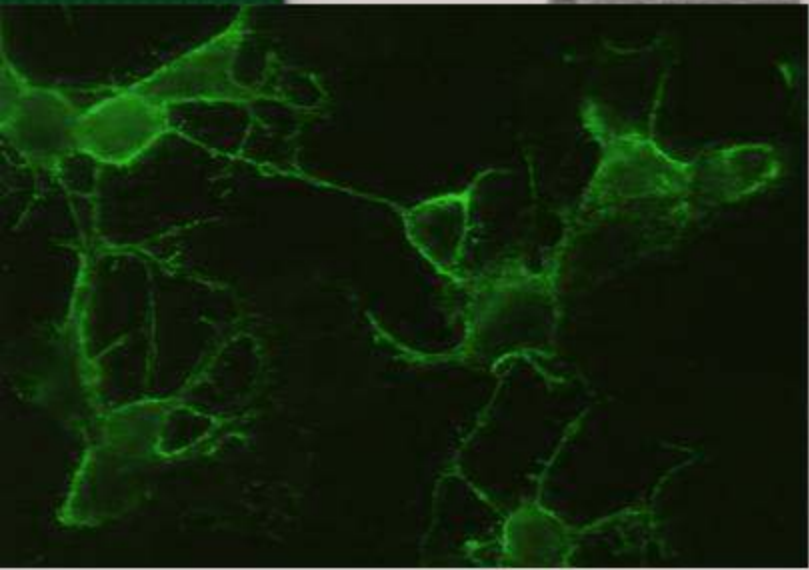
RT-PCR (Standart ters transkriptaz polimeraz zincir reaksiyonu)

- *in-house* (Laboratuvar yapımı)-idrar!

Serolojik Testler (IgM,IgG)

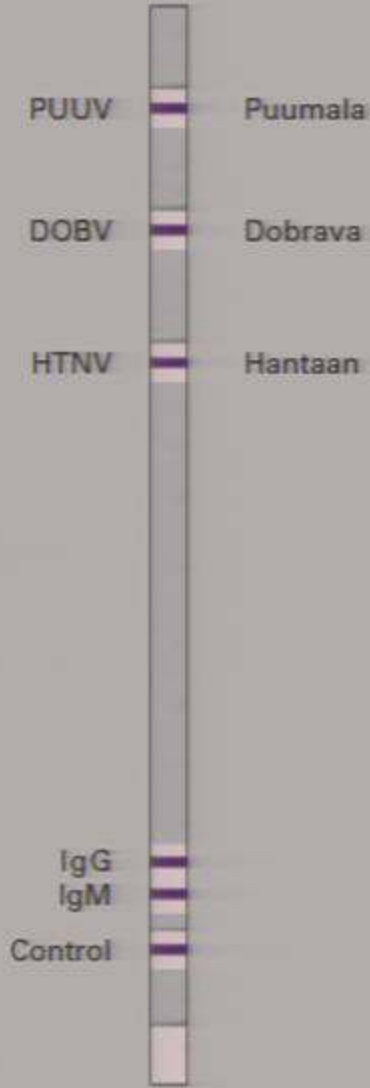
- ELISA (Ticari)—yalancı pozitiflik, negatiflik
- IFA (Ticari)
- Immunblot (Ticari)

2- Tanı yöntemleri



Tanıma yöntemleri

Antigen combination



- Nükleokapsid antijenlerine karşı oluşan antikolar yakalanıyor.



Tatarcık Humması Tanısı

rRT-PCR

- *in-house* (Laboratuvar yapımı)
- SFTurkeyV, SFSV, SFToscV, SFNV

Serolojik Testler (IgM, IgG)

- IFA (Ticari)

Batı Nil Virüsü Tanı Algoritması

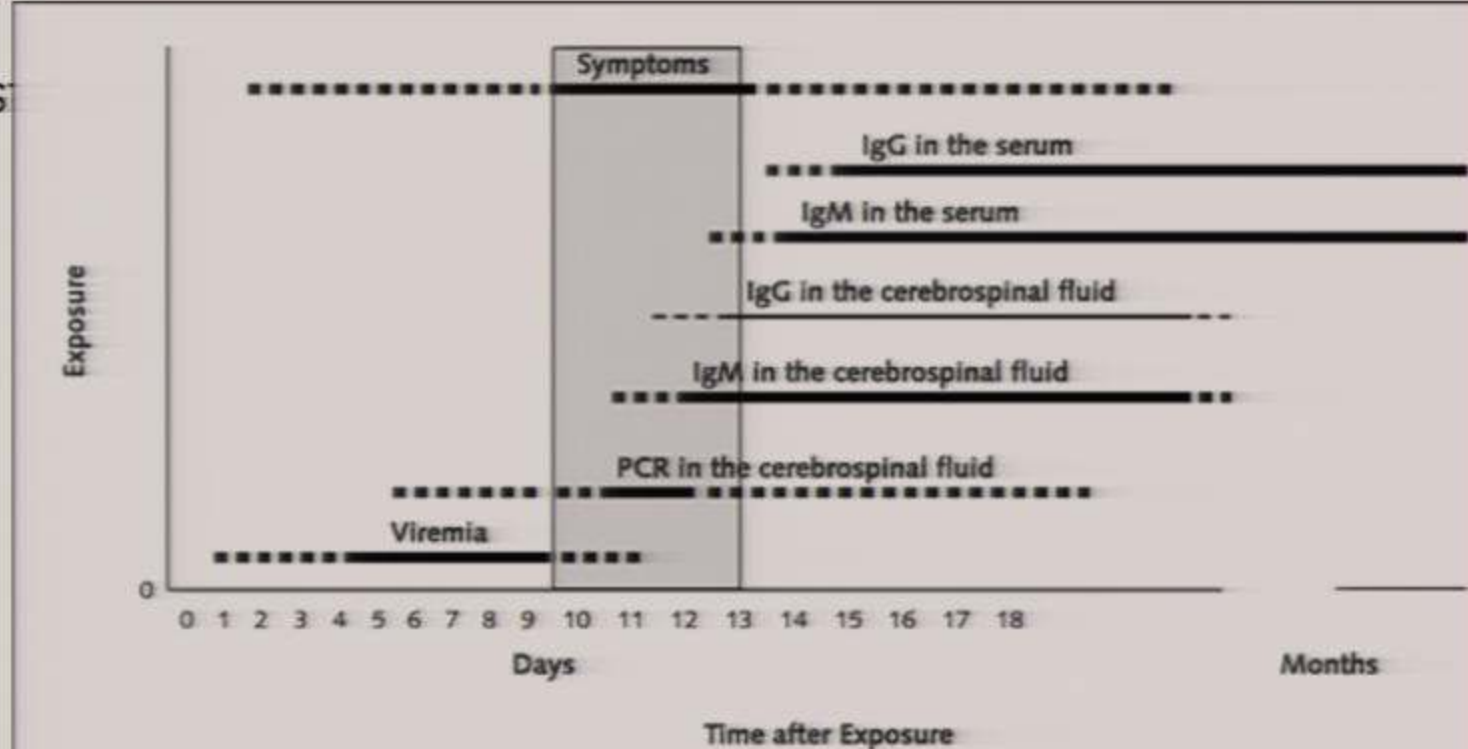
rRT-PCR (BOS, Serum ve İDRAR!)

- Ticari Kit(Kantitatif)
- *in-house* (Ozkul A et-al)

Serolojik Testler (IgM,IgG)

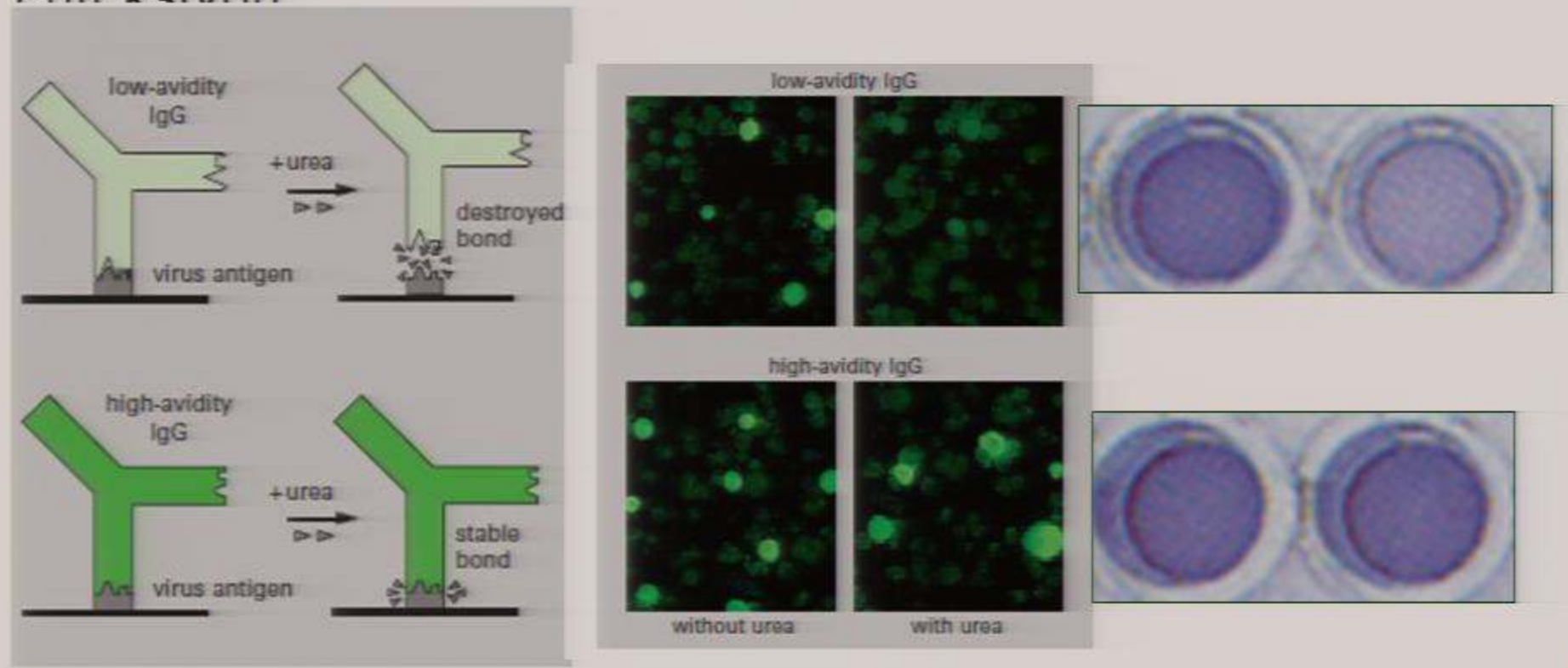
- ELISA (Ticari)
- IFA (Ticari)
- Avidite testi
- Nötralizasyon tes

Ann Intern Med. 2004;140(7):545-55



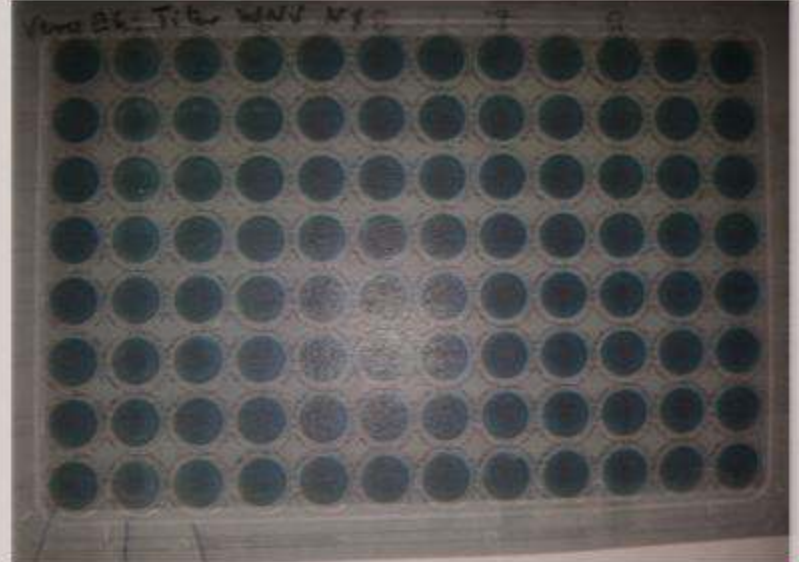
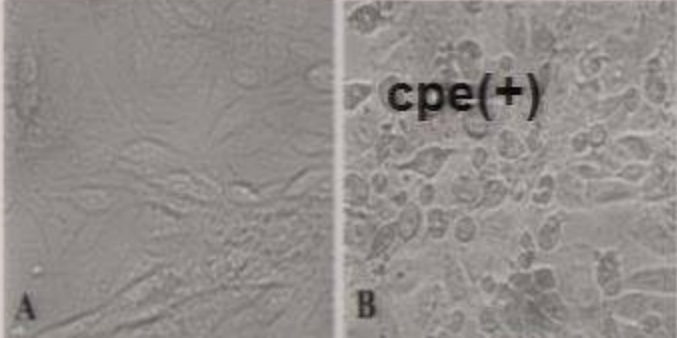
BNV Avidite testi

- Düşük avidite (ürelü/üresiz OD <%40); <20 gün geçirilmiş enfeksiyon
- Yüksek avidite (>%60); >40 gün önce geçirilmiş enfeksiyon



BNV Mikronötralizasyon testi Virus Titrasyonu (1. hafta)

- RKI ile eşleşme projesi kapsamında canlı viruslar (Lineage 1-NY99 ve Lineage 2-hEJa) 2013 yılı sonunda temin edildi.
- 2014 yılı başında BSL-3 laboratuvarında çalışmalara başlandı.
- Virus titrasyonu (Lineage 1-NY99 ve Lineage 2-hEJA) (1/10....1/10⁸)..>cpe
- 1. gün; Vero E6 hücresi hazırlanıyor.
- 2. gün; virus ekleniyor.
- 5. gün; intakt hücre sayısına göre kuyucukların yarısında cpe'ye neden olan virus titrasyonu (TCID₅₀) hesaplanır.

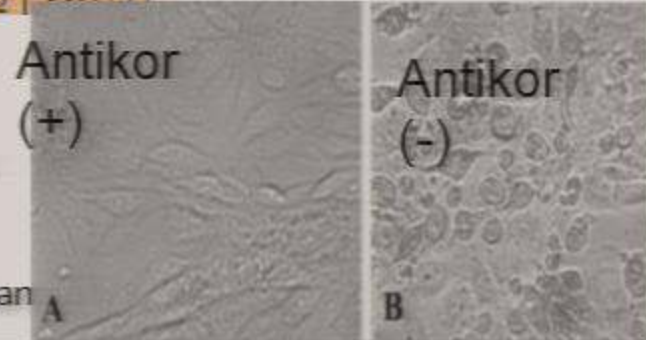


BNV Mikronötralizasyon testi (2. hafta)

- 1. gün Vero E6 hücresi ekiliyor.
- 2. gün hasta serumu ekleniyor. Ardından titre edilmiş virus ekleniyor.
- 5. gün invert mikroskop altında değerlendiriliyor.

	1	2	3	4	5	6	7	8	9	10	11	12
A	Serum 1 1:5	Serum 1 1:5	Serum 2 1:5	Serum 2 1:5							cell control	virus control
B	Serum 1 1:10	Serum 1 1:10	Serum 2 1:10	Serum 2 1:10							cell control	virus control
C	Serum 1 1:20	Serum 1 1:20	Serum 2 1:20	Serum 2 1:20							cell control	virus control
D	Serum 1 1:40	Serum 1 1:40	Serum 2 1:40	Serum 2 1:40							control titer-2	control titer-2
E	Serum 1 1:80	Serum 1 1:80	Serum 2 1:80	Serum 2 1:80							control titer-1	control titer-1
F	Serum 1 1:160	Serum 1 1:160	Serum 2 1:160	Serum 2 1:160							control titer	control titer
G	Serum 1 1:320	Serum 1 1:320	Serum 2 1:320	Serum 2 1:320							control titer+1	control titer+1
H	Serum 1 control	Serum 1 control	Serum 2 control	Serum 2 control							control titer+2	control titer+2

Hasta serumunda nötralizan antikor varsa hücrelerde cpe izlenmiyor.



2- Tanı yöntemleri

- Kene kaynaklı ensefalit
ELISA ve IFA IgM, IgG
- Sarı humma
IFA IgM, IgG
- Dengue 1,2,3,4
Multiplex PCR ve IFA IgM, IgG
- Chikungunya
IFA IgM, IgG

3- Pozitif vaka verileri KKKA



- Yaklaşık %5 mortalite
- En yüksek vaka sayısı Tokat (1800 vaka), Yozgat, Çorum, Sivas, Erzurum
- Vakanın henüz görülmediği iller; Düzce, Karaman ve Kilis.

KKKA İNSİDANSININ İLLERE GÖRE DAĞILIMI (1/100.000), 2011



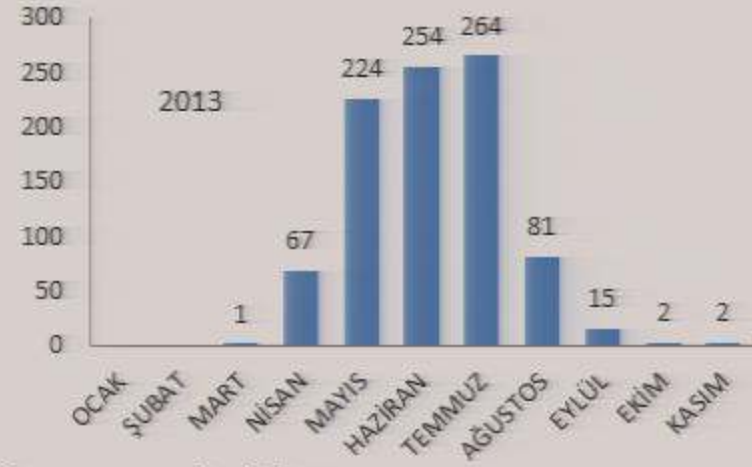
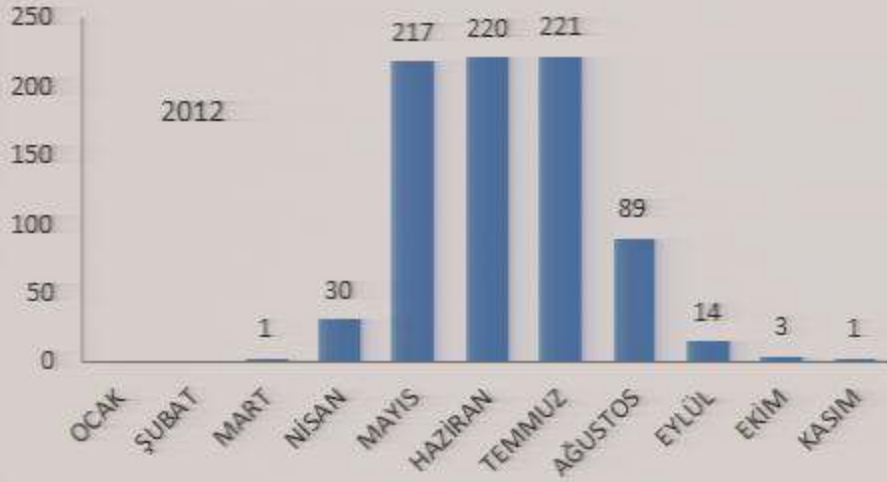
KKKA İNSİDANSININ İLLERE GÖRE DAĞILIMI (1/100.000), 2012



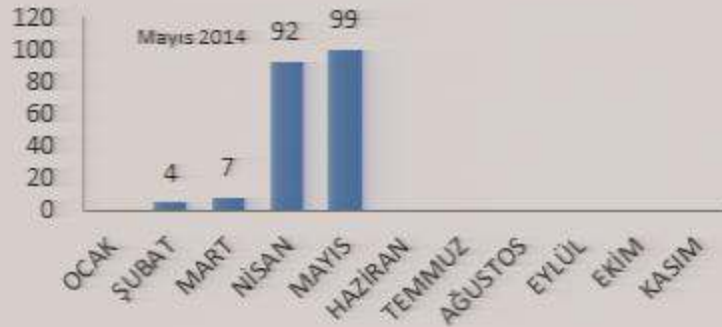
KKKA İNSİDANSININ İLLERE GÖRE DAĞILIMI (1/100.000), 2013



Aylara göre vaka dağılımı



Vakalar en çok Mayıs,
Haziran ve Temmuz
aylarında görülüyor.



*Zoonotik ve Vektörel Hastalıklar Daire Başkanlığı





Sidira P et al, CMI
2012 %27,5!

Vaka sayısı ile seropozitiflik oranı korele değil.
Sebebi;
1-Dolaşan suşların farklı olması
2-İmmun yanıtta farklılık olması

Güneş T ve ark, EID 2009	%12,8 (Tokat)	~1800 vaka/184.000kişi
Gargılı A ve ark, VBZ 2011	%10,9 (Kırklareli kırsalı)	~ 12 vaka/89.000 kişi
Bodur H ve ark, EID 2012	%10 (Endemik 12 il kırsalı)	
Ertuğrul B ve ark, Scand J Infect Dis 2012	%19,6 (Aydın kırsalı)	~ 60 vaka/1.000.000 kişi
Yagci-Caglayik D ve ark, JMV 2013	%16,7 (Yozgat kırsalı)	~ 840 vaka/97.000 kişi





Sebep;

- 1-Dolaşan suşların farklı olması
- 2-İmmun yanıtta farklılık olması



The relationship between the human leukocyte antigen (HLA) polymorphisms and the prevalence of Crimean-Congo hemorrhagic fever in the Turkish

Esragül Akıncı^{a,*}, Hürrem Bodur^a, Uğur Muşabak^b, Raşan I. S.

^aAnkara Numune Education And Research Hospital, Infectious Diseases and Clinical Microbiology Department, Ankara, Turkey
^bGülhan Military Medical Academy, Immunology and Allergy Division, Ankara, Turkey

Kontrol grubuna göre hasta grubunda;
HLA A23; şiddetli seyir
 HLA B27 ; koruyucu (Chikungunya'da ise; yatkinlık)
HLA A2; duyarlılıktan sorumlu bulunmuş.

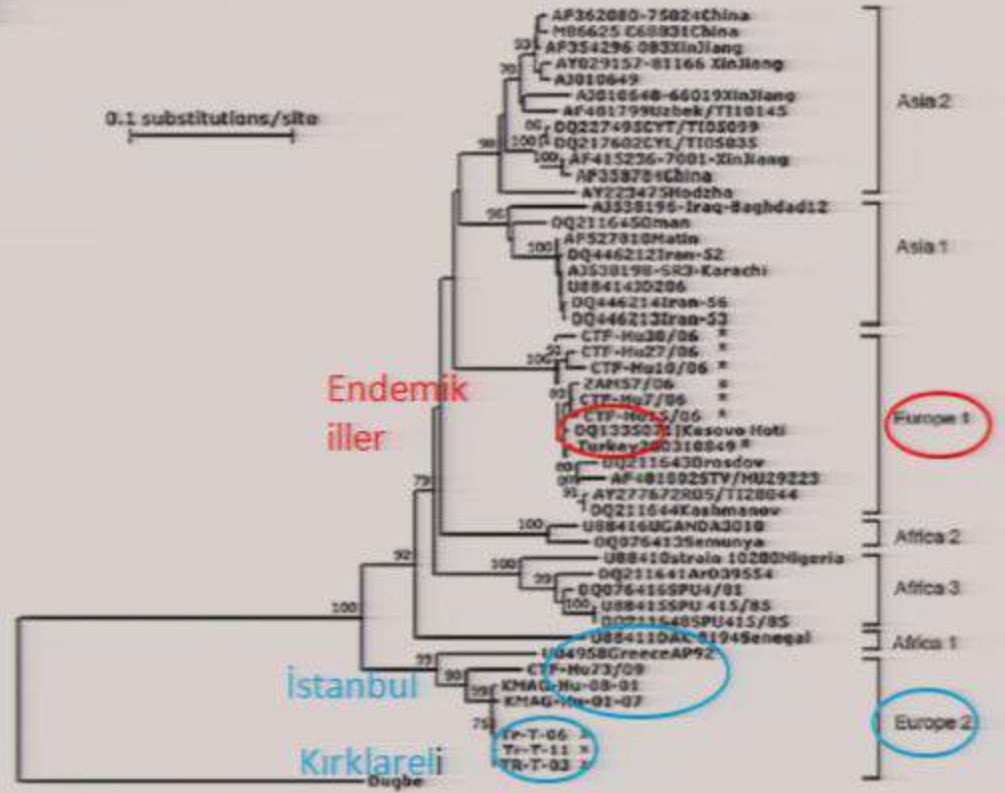


FIG. 3. The phylogenetic relationships of the CCHFV strains. Strains marked with * have been previously recorded humans, those with # have been recorded from ticks in Turkey. Strains marked with x are recorded for the first time study.

Midilli K ve ark, BMC Infect Dis 2009
 Gargılı A ve ark, VBZ 2011



3- Son beş yılın pozitif vaka verileri HANTAVİRUS



*Zoonotik ve Vektörel Hastalıklar Daire Başkanlığı



Hantavirus vakalarının dağılımı



- Batı karadenizde Puumala serotipine, Trabzon ve Giresun başta olmak üzere Dobrava serotipi görülüyor. İç batı Anadolu'da da DOBV görülüyor.
- Şimdiye kadar PCR pozitif 2 vaka oldu, 2si de İstanbul'dan ve DOBV saptandı.
- Akdeniz kıyısında vaka görülüyor.



İzmir ve Trabzon'da 2004'te rodentlerde
PUUV varlığı serolojik olarak
gösterilmiştir. Laakkonen J et al, Journal
of Wildlife Diseases 2006

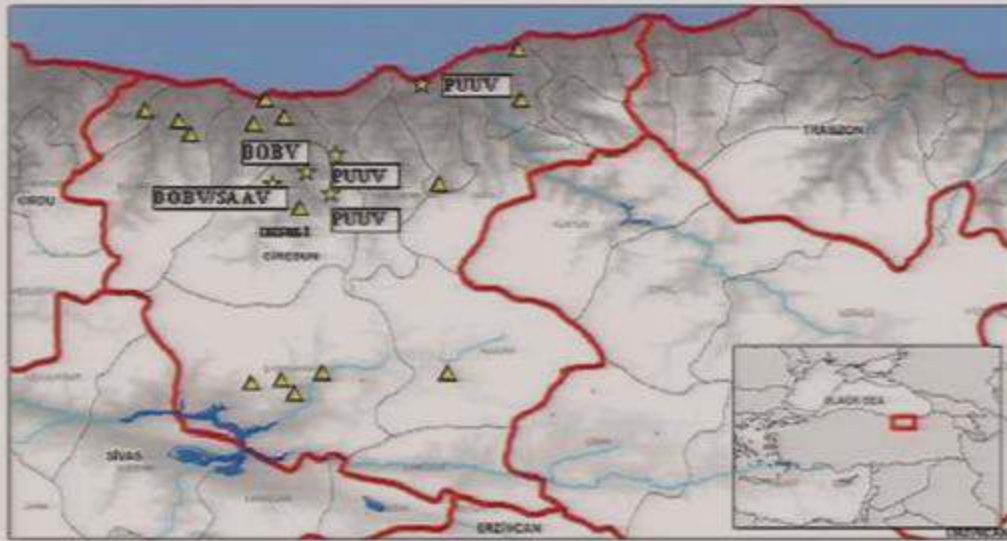
Avrupa'da PUUV ve DOBV taşıyan
rodentler Türkiye'de de bulunuyor.

Bartın'da rodentlerde DOBV varlığı
araştırıldı.

M. glareolus Ø

A. flavicollis %20'sinde DOBV RNA (+)
Öktem İMA ve ark, EID 2014





★ seropositivity by FRNT

▲ seropositivity by immunoblotting assay

0 5 10 20 30 40
Kilometers



Gözalan A ve ark, VBZ 2013



Fig. 1. Map of Giresun, results of the seropositivity by using immunoblotting assay and focus reduction neutralization test (FRNT). (Color image available online at www.liebertpub.com/vbz).

- Seropozitiflik %3 (DOBV+PUUV)
- 2000 yılından itibaren fındık yerine kivi üretilmesi ve orman alanlarının azalması; rodentleri yiyecek bulmak için insanların yerleşim bölgelerine gitmeye zorladı.

HLA-B8, -DR3, -DQ2 alellerine sahip bireylerde hemodiyaliz ihtiyacının daha fazla ve prognozun daha kötü olduğu; HLA-B27 aleline sahip olanlarda ise hastalığın daha hafif seyrettiği bildirilmektedir. Kaya S, Mikrobiyol bul 2014



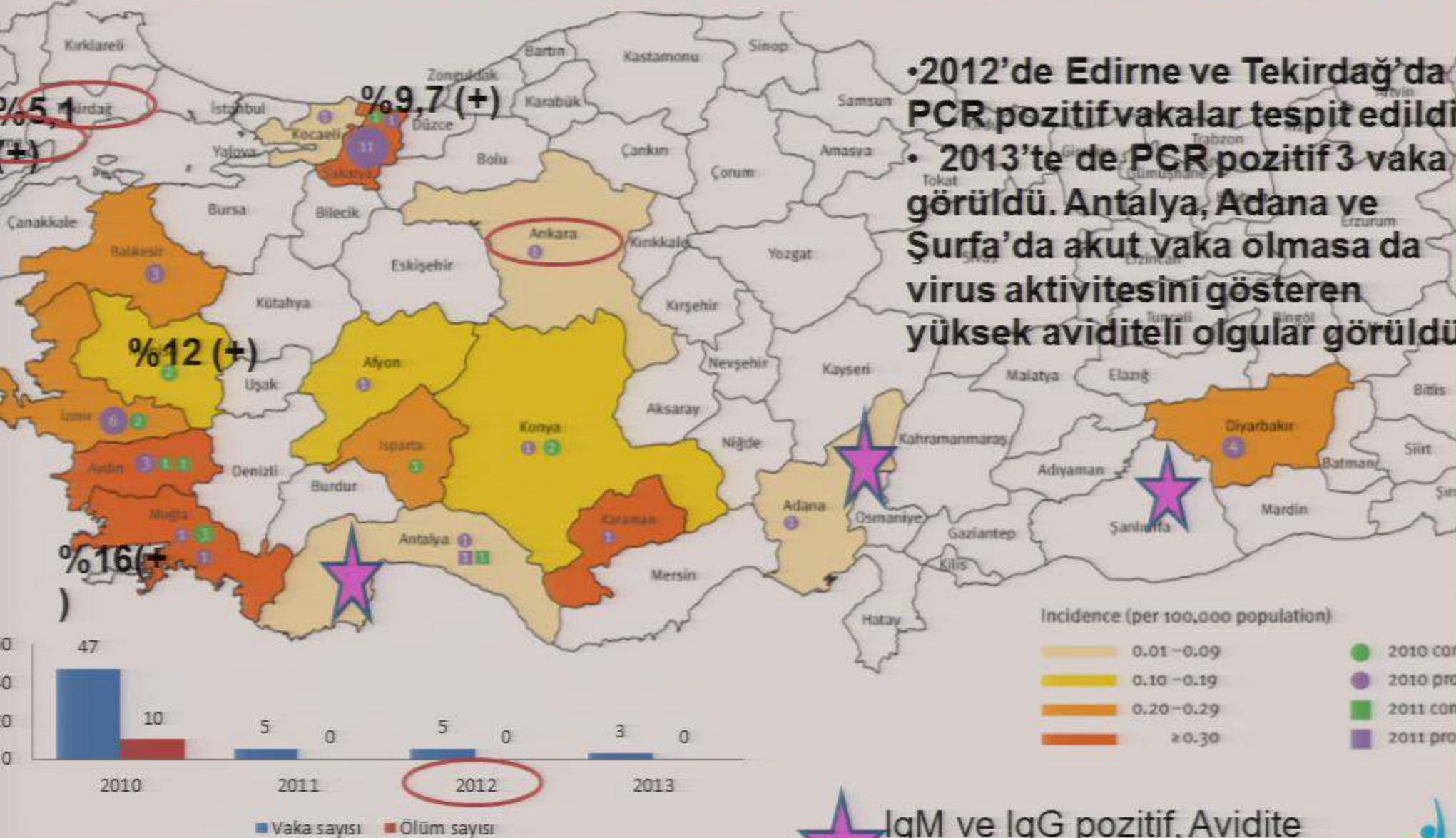
3- Son beş yılın pozitif vaka verileri BATI NİL VİRUSU

Emergence of West Nile virus infections in humans in Turkey, 2010 to 2011

FIGURE 2

Number of West Nile virus cases in 2010 (n=47) and

H Kalaycıoğlu (h.kalaycioglu@hotmail.com)¹, G Korukluoğlu¹, A Özkul², O Oncul³, S Tosun³, O Karabay⁴, A Gozalan⁵, Y Uyar⁶, D Y Çağlayık⁷, G Atasoylu⁸, A B Altas⁹, S Yolbakan⁹, T N Özden⁹, F Bayraktar⁹, N Sezak⁹, T S Pelitli⁹, Z Ö Kurtcebe⁹, E Aydın⁹, M Ertek⁹



	23.7.2013	31.7.2013
	POZİTİF (BOS)	POZİTİF (idrar)
PCR	POZİTİF (serum)	negatif (serum)
ELISA IgM	Negatif	POZİTİF
IFA IgM	POZİTİF	POZİTİF

2013 yılında, idrarda PCR pozitifliği yakalandı.

- Çocuk nörolojisinde optik nörit öntanısıyla izlenen hastada sonrasında şiddetli seyirli MS tanısı kondu.
- BNV enfeksiyonunun MS tetikleyicisi olabileceğini düşündürdü.
- BOS pozitifliği yakalanamayan hastalarda mutlaka idrar PCR bakmak gerekiyor.



3- Son beş yılın pozitif vaka verileri

- 2010 vakalarından sonra RSHMB tarafından Temmuz-Aralık 2011 arasında vaka çıkan 4 ilde (Sakarya, Manisa, Muğla, Edirne) vaka surveyansı başlatıldı.
- Vaka tanımına uyan 46 hastaya ait 51 örnek analiz edildi.
- Tam kan, plazma, lökosit, BOS, serumdan PCR çalışıldı. PCR pozitifliğine rastlanamadı.
- Serumda ELISA, IFA, PRNT (A.Ü.V.F.) çalışıldı.



VS-54/018 (9.10.2011)	ELISA IgM/IgG	IFA IgM/IgG	PRNT
VS-54/018 (10.10.2011 (5. gün))	Aradeğer/Negatif	Pozitif/Negatif	Pozitif
VS-48/002 (15.8.2011)	ELISA IgM/IgG	IFA IgM/IgG	PRNT
VS-48/002 (18.8.2011 (3. gün))	Aradeğer/Negatif	Pozitif/Negatif	Pozitif
VS-48/002 (12.2011 (3. ay))	Negatif/Negatif		Negatif
VS-45/002 (15.8.2011)	ELISA IgM/IgG	IFA IgM/IgG	PRNT
VS-45/002 (18.8.2011 (6. gün))	Pozitif/Negatif	Aradeğer /Negatif	Pozitif
VS-45/002 (10.2011 (2. ay))	Negatif/Negatif		Negatif
VS-48/102 (16.7.2011)	ELISA IgM/IgG	IFA IgM/IgG	PRNT
VS-48/102 (13.2011 (9.gün))	Negatif/ Pozitif		

Tekrar kanlarında nötralizasyon testi negatifleşti, IgG pozitifleşmedi.

Tekrar örnek istenen vakaların laboratuvar sonuçları-Sakarya

VS-54-005 (30.7.2011)	ELISA IgM/IgG	IFA IgM/IgG	PRNT
VS-54-005 (5.8.2011 (6. gün))	Pozitif/Negatif	Aradeğer/Negatif	Negatif
VS-54-005 (7.9.2011 (1. ay))	Pozitif/Negatif	Aradeğer/Negatif	Negatif
VS-54-121 (15.9.2011)	ELISA IgM/IgG	IFA IgM/IgG	PRNT
VS-54-121 (19.9.2011 (4. gün))	Aradeğer/Negatif	Aradeğer/Negatif	Negatif
VS-54-121 (21.10.2011 (1. ay))	Aradeğer/Negatif	Pozitif/Negatif	Pozitif
VS-54-121 (21.11.2011(2. ay))	Negatif/Negatif		Negatif

•PRNT cut off değeri yükseltildi.

•Mikronötralizasyon yöntemi tanısal kapasiteye eklendi.

•Ülkemizde, çapraz rxn'a neden olan başka bir flavivirusun dolaşıyor olabileceği düşünüldü.





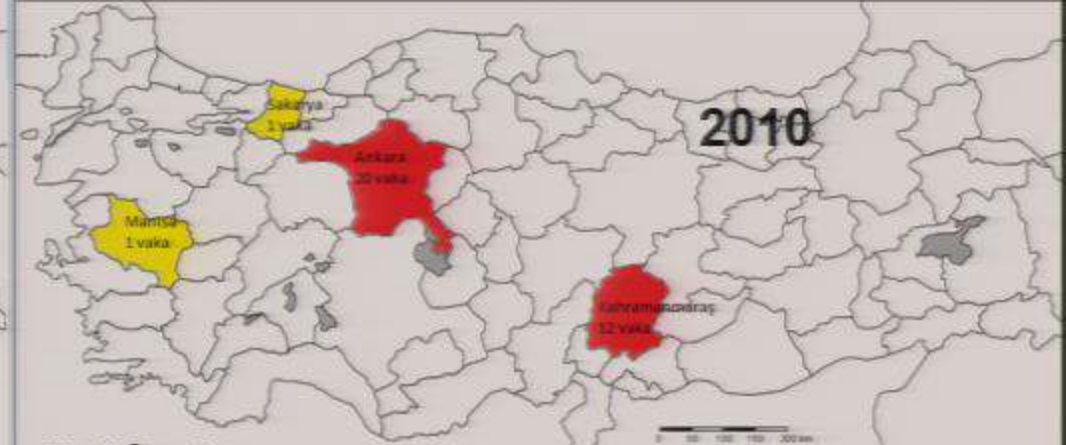
- Erdem H ve ark, Clin Microbiol Infect 2013
- Ergunay K ve ark, VBZ 2013
- Karakoç ZÇ ve ark, VBZ 2013
- Albayrak H ve ark, J Arthropod-Borne Dis 2013
- Öcal M ve ark, Mikrobiyol Bul 2013



KAN BANKASI -BNV

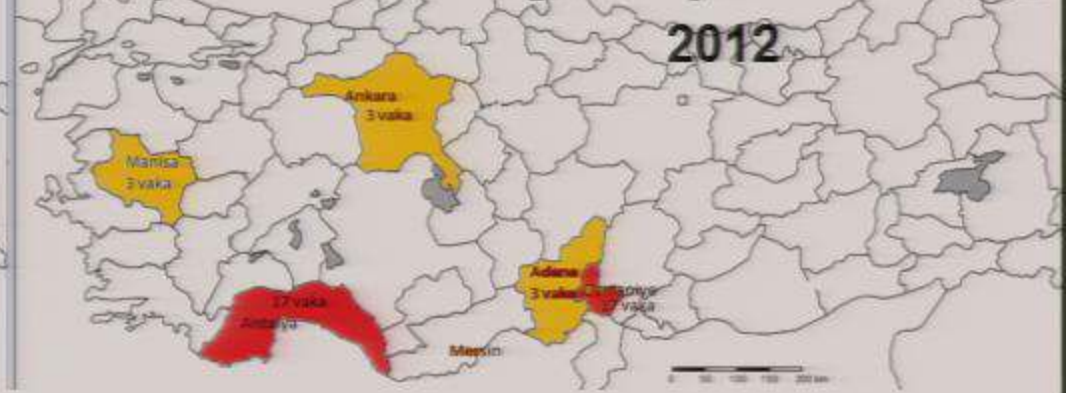
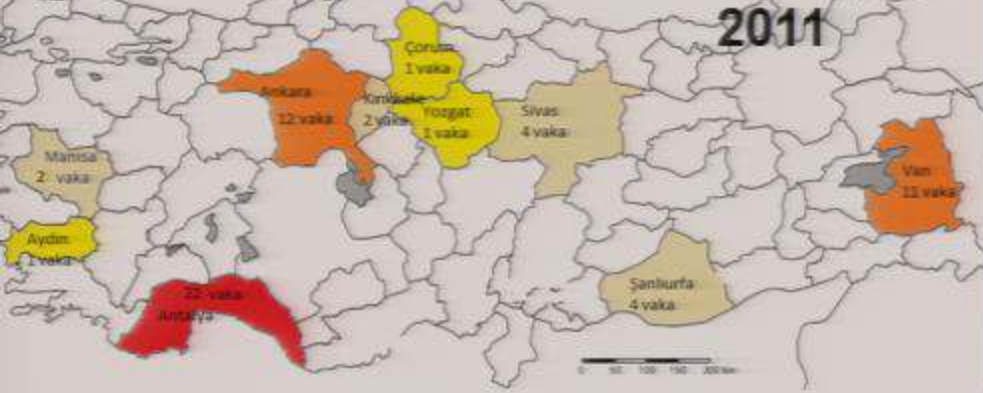
Kan bankası çalışmalarının hiçbirinde PCR pozitifliğe rastlanmadı.

- -Muğla, Sakarya, Edirne, Manisa
- -Ankara(2) (Ergunay K ve ark., Hızal K ve ark, Mikrobiyol Bul 2010)
- -Tekirdağ (Erdem H ve ark, Clin Microbiol Infect 2013)



Hantavirusun aksine Karadenizden vaka görülüyor.

Ağırlıklı Sicilya alt tipi olmak üzere, tüm serotipler ülkemizde görülüyor.



Ortalama 45 vaka/yıl



3- Son beş yılın pozitif vaka verileri Sandfly fever virus

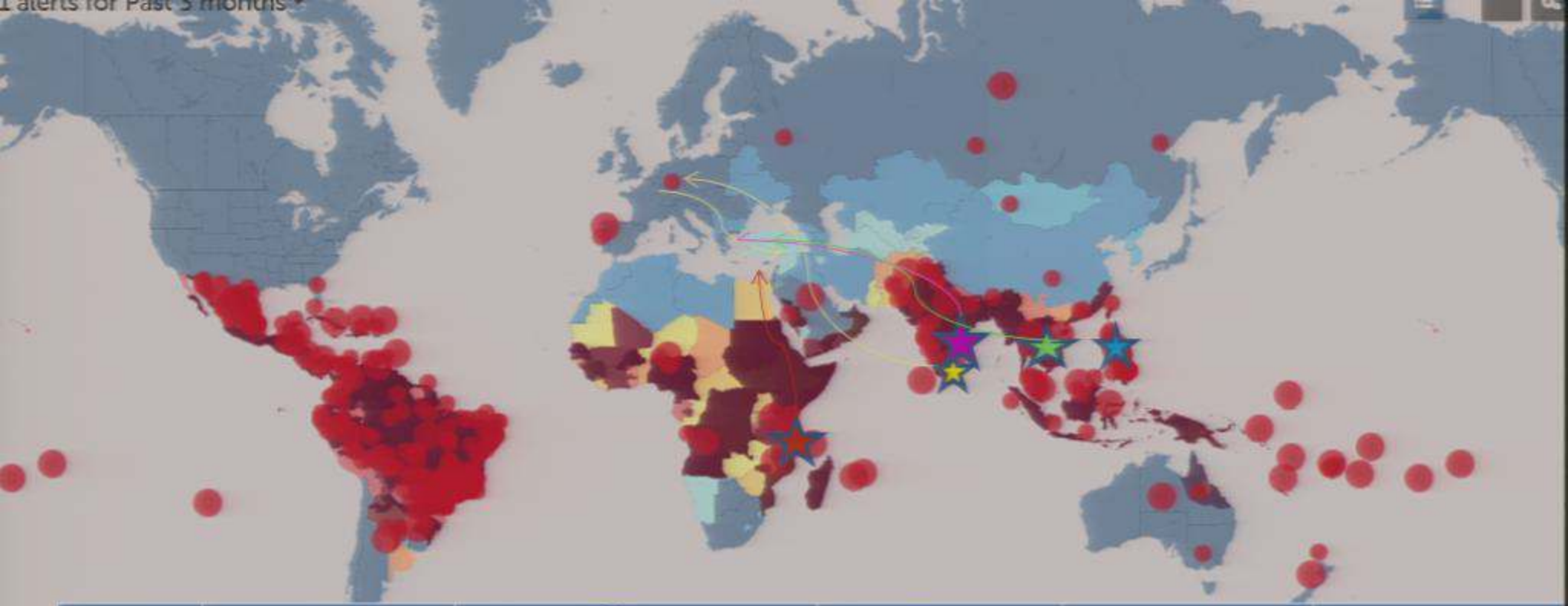
- Aynı virus ailesine mensup olmaları nedeniyle Sandfly IgM testinde, KKKA IgM testiyle çapraz reaksiyonlar izlenmektedir.
- Bunun için,
- -KKKA PCR ve Sandfly nötralizasyon testleri önemlidir.
- -SFV nötralizasyon testi de hedeflerimiz arasındadır.

3- Son beş yılın pozitif vaka verileri

- Temmuz 2011 Ankara'da tatarcık dağılımı
- P. major s.l. (38.9%), ..SFTV ile enfekte.
- P. sergenti (20.4%),
- P. halepensis (17.7%), and P. papatasi (10.2%),

Ergunay ve ark, 2013

3- Son beş yılın pozitif vaka verileri DANG ATEŞİ VE CHIKUNGUNYA



	Tarih	Geldiği Ülke	Durumu	Uyruk	
1	Eylül 2011 (İzmir)	Hindistan 	Sağ	İsviçre	DENV tip 3
2	Mart 2013 (Köln- Ankara**)	Sri Lanka (KİT sonrası) 	Ex	Türkiye	DENV tip 1
3	Nisan 2014 (Ankara)	Tayland 	Ex (otopsi numunesi)	Tayland	DENV tip 3
4	Nisan 2014 (İzmir)	Filipinler 	Sağ	Filipinler	DENV tip 3
5	Mayıs 2014 (Antalya)	Tanzanya 	Sağ	Tanzanya	DENV tip 3

Dengue virus transmission due to allogeneic blood stem cell transplantation

Journal:	<i>Emerging Infectious Diseases</i>
Manuscript ID:	EID-14-0508.R1
Manuscript Type:	Dispatch
Date Submitted by the Author:	04-May-2014
Complete List of Authors:	Punzel, Michael: Cellex Apheresis and Collection Center, MediaPark Klinik, Konukluoglu, Gulay: Refik Saydam National Public Health Agency, Virology Dep. Yago, Caclavik, Dilek: Refik Saydam National Public Health Agency (RSNPHA), Virology Reference and Research Laboratory Menemenliolu, Dilek: Refik Saydam National Public Health Agency, Virology Dep. Bozdog, Sinem: Abdurrahman Yurtarslan Ankara Oncology Hospital, Tekgunduz, Emre: Abdurrahman Yurtarslan Ankara Oncology Hospital, Altuntas, Fevzi: Abdurrahman Yurtarslan Ankara Oncology Hospital, Burde, Bernd: MVZ Labor Dr. Quade & Kollegen. Gunther, Stephan: Bernhard Nocht Institute for Tropical Medicine, Department of Virology Tappe, Dennis: Bernhard Nocht Institute for Tropical Medicine, Department of Virology Cadar, Daniel: Bernhard Nocht Institute for Tropical Medicine, Department of Virology Schmidt-Chanasit, Jonas: Bernhard Nocht Institute for Tropical Medicine, Department of Virology
Keywords:	Dengue, Transplantation, PCR, Germany, Turkey, Fatal



Countries and territories where chikungunya cases have been reported* (as of May 12, 2014)



Olgu Sunumu/Case Report

Mikrobiyol Bul 2012; 46(1): 122-

Ankara'ya Hindistan Yeni Delhi'den Gelen Bir Chikungunya Ateşi Olgusu: Türkiye'nin İlk Yurt Dışı Kaynaklı Olgusu ve Literatürün Gözden Geçirilmesi

An Imported Chikungunya Fever Case from New Delhi, India to Ankara, Turkey: The First Imported Case of Turkey and Review of the Literature

Dilek YAĞCI CAĞLAYIK¹, Yavuz UYAR¹, Gülay KORUKLUOĞLU¹, Mustafa ERTEK², Serhat ÜNAL³

24th **ECCMID** Barcelona, Spain
10 – 13 May 2014

ESCMID EUROPEAN SOCIETY OF CLINICAL MICROBIOLOGY AND INFECTIOUS DISEASES

	Tarih	Geldiği Ülke	Durumu
1	Aralık 2010 (Ankara)	Hindistan	Sağ (eşinde IgG+)
2	Temmuz 2013 (Ankara)	Tayland	Sağ
3	Temmuz 2013 (Ankara)	Tayland	Sağ

Abstract
Chikungunya fever is a disease caused by Chikungunya virus which is transmitted to humans by Aedes mosquitoes and presents with fever, headache, rash, and severe arthralgia. Chikungunya is endemic in Africa, Indian Ocean, West Pacific Ocean islands, and in South-East Asia. Only one case of Chikungunya fever was detected two years ago in Turkey. In this report we presented the imported Chikungunya fever case from Turkey who visited Thailand.

Case: A married couple admitted to our clinic with the complaints of fever, muscle and arthralgia in 2013 July. They had a history of tourist trip to Thailand where they had spent 15 days two weeks ago. There was a delay of mosquito bite during the trip. They had a sudden onset of high grade fever (39.3/38.0°C), headache that lasted 5 days, along with a fever rash that spread to their trunk, legs, and ankles, followed by symmetrical, labrality joint showed knee spaces (>4000 µL) in both of them. They had leukopenia (22000 µL), elevated levels of aspartate transaminase (111 U/L), elevated transaminase (80 U/L), and creatinine (1.4 mg/dL) was detected only in man.

The culture for virus obtained were negative. Test results for Phlebotomus spp., Toxoplasma spp., Isospora spp., malaria, varicella zoster and Chikungunya virus were negative. Blood samples were tested in Halk Sağlığı Kurumu Public Health Agency, Virology Laboratory for Chikungunya virus (CHIKV) antibody. Anti-CHIKV IgM antibodies were positive and IgG antibodies were negative with immunofluorescence assay (IFA). Samples were tested in seventh day of the disease.

Conclusion: Turkey is a non-endemic region for Chikungunya virus infection. Chikungunya disease should be considered in the differential diagnosis in patients with a history of travel to hyperendemic areas particularly Africa, South-East Asia, and who experienced acute unexplained fever with joint pain and rash. Although Aedes aegypti is not the vector of Turkey mosquito fauna, Aedes albopictus being a transient member of fauna, may cause epidemics due to import cases.



Aseptic Meningitis due to Yellow Fever Vaccination

Mücahit Yemişen¹, Ümran Şumeyse Ertürk¹, Dilek Yağcı Çağlayık², Dilek Menemenli
Seven Uludokumacı¹, İlker İnanç Balkan¹, Bilgül Mete¹, Fehmi Tabak¹

¹Istanbul University Cerrahpasa Medical Faculty, Division of Infectious Diseases and Clinical Microbiology

²Public Health Institution of Turkey, Microbiology Reference Laboratories Department, National Arbovirus and Zoonoses Reference and Research Laboratory



IgM, IgG Pozitif vakalar	Aşı öyküsü	Klinik	Durumu
Şubat 2012	Var	Aşı yan etkisi- menenjit (1 ay sonra)	Sağ
Aralık 2012	Var	Ateş, Hepatomegal i, LAP (Etiyopya)	Sağ

DIŞ KALİTE KONTROL



OPEN ACCESS Freely available online

PLOS NEGLECTED TROPICAL DISEASES

First International External Quality Assessment of Molecular Detection of Crimean-Congo Hemorrhagic Fever Virus

Camille Escadafal^{1,2*}, Stephan Ölschläger³, Tatjana Avšič-Županc⁴, Anna Papa⁵,
Jessica Vanhomwegen^{2,6}, Roman Wölfel⁷, Ali Mirazimi⁸, Anette Teichmann¹, Oliver Donoso-Manríquez⁹,
Matthias Niedrig¹

¹Centre for Biosafety, Robert Koch Institute, Berlin, Germany, ²European Public Health Microbiology Training Program (EUPHEM), European Centre for Disease Prevention, Berlin, Germany, ³Robert Koch Institute, Berlin, Germany, ⁴University of Ljubljana, Ljubljana, Slovenia, ⁵University of Medicine and Health Sciences, Hanoi, Vietnam, ⁶University of Cologne, Cologne, Germany, ⁷Robert Koch Institute, Berlin, Germany, ⁸University of Tehran, Tehran, Iran, ⁹University of Chile, Santiago, Chile

- Avrupa 1 (Hoti suşu)
 - Avrupa 2 (AP-92 suşu)
 - Asya 1 (Afg09-2990 suşu)
 - Afrika 3 (Moritanya suşu)
- genotiplerinden oluşan panelin tümü yakalandı.
- ENIVD üyesi 47 referans laboratuvar (EHL,SHL,AVZHU)



Analysis of your laboratory's Qualitative EQA data

Analysis of your laboratory's performance on the core proficiency samples



The core proficiency samples in this EQA programme were:

WNV12-01, ~~WNV12-02~~, WNV12-03, WNV12-04, ~~WNV12-05~~, WNV12-06, WNV12-07, WNV12-08, WNV12-10, WNV12-12

You reported 9/9 (100.0 %) of the core samples correctly.

Of the total datasets reported by all participants in this EQA programme, 72.2% reported correct results for all core proficiency samples.

Analysis of your laboratory's performance on all proficiency samples:

Your laboratory's qualitative results and performance scores

Sample	Sample Content	Sample Status	Qualitative		
			Sample Type	Your qualitative result	Your qualitative score
WNV12-06	West Nile Virus NY99	Frequently detected	Core	positive	0
WNV12-12	West Nile Virus NY99	Frequently detected	Core	positive	0
WNV12-10	West Nile Virus NY99	Frequently detected	Core	positive	0
WNV12-02	West Nile Virus NY99	Detected	Core	positive	0
WNV12-08	West Nile Virus NY99	Detected	Core	positive	0
WNV12-11	West Nile Virus NY99	Detected		positive	0
WNV12-03	West Nile Virus Heja	Detected	Core	positive	0
WNV12-07	West Nile Virus Heja	Detected	Core	positive	0
WNV12-01	West Nile Virus Ug37	Frequently detected	Core	positive	0
WNV12-05	Non-WNV Flaviviruses	Negative		negative	0
WNV12-09	Non-WNV Flaviviruses	Negative		negative	0
WNV12-04	WNV Negative	Negative	Core	negative	0
Sum Qualitative Panel Score					0



Dengue PCR- (2013 QCMD paneli) %100 doğruluk

Three participants did not return results. Two of these withdrew officially due to logistical issues.

Panel composition

Sample code	Sample content	Sample * matrix	Sample conc. Copies/ml	Sample status
EPI_DENV13-03	Dengue Virus Type 1	VTM	1.0x10E6	Frequently detected
EPI_DENV13-11	Dengue Virus Type 1	VTM	1.0x10E5	Frequently detected
EPI_DENV13-07	Dengue Virus Type 1	VTM	1.0x10E5	Frequently detected
EPI_DENV13-06	Dengue Virus Type 1	VTM	1.0x10E4	Detected
EPI_DENV13-08	Dengue Virus Type 1	VTM	1.0x10E3	Detected
EPI_DENV13-09	Dengue Virus Type 2	VTM	1.0x10E5	Frequently detected
EPI_DENV13-04	Dengue Virus Type 2	VTM	1.0x10E4	Detected
EPI_DENV13-10	Dengue Virus Type 3	VTM	2.8x10E5	Detected
EPI_DENV13-01	Dengue Virus Type 3	VTM	2.8x10E4	Detected
EPI_DENV13-05	Dengue Virus Type 4	VTM	1.2x10E5	Detected
EPI_DENV13-12	Non-DENV Flaviviruses ²	VTM	Each 1.0x10E5	Negative
EPI_DENV13-02	DENV Negative VTM	VTM		Negative

RESULTS:

Sample ID	Qualitative Results	Quantitative Results		Comments
		Value	Cycle Threshold	
EPI_DENV13-01	positive		27.57	Dengue 3
EPI_DENV13-02	negative			Negative
EPI_DENV13-03	positive		18.50	Dengue 1
EPI_DENV13-04	positive		29.69	Dengue 2
EPI_DENV13-05	positive		22.53	Dengue 4
EPI_DENV13-06	positive		24.45	Dengue 1
EPI_DENV13-07	positive		21.68	Dengue 1
EPI_DENV13-08	positive		27.89	Dengue 1
EPI_DENV13-09	positive		27.79	Dengue 2
EPI_DENV13-10	positive		24.54	Dengue 3
EPI_DENV13-11	positive		22.25	Dengue 1
EPI_DENV13-12	negative			Negative





T.C. SAĞLIK BAKANLIĞI
TÜRKİYE HALK SAĞLIĞI KURUMU



2014 YILI
FİYAT TARİFELERİ
VE
ANALİZ BİLGİLERİ
REHBERİ

a) Ulusal Viroloji Referans Merkez Laboratuvarları Analizleri:

İsim Bilgileri:

Türkiye Halk Sağlığı Kurumu
Mikrobiyoloji Referans Laboratuvarları Daire Başkanlığı
Ulusal Viroloji Referans Merkez Laboratuvarı

Sağlık Bakanlığı Adnan Saygun Cad. No:55 F Blok 1-2. Kat
06100 Sıhhiye-ANKARA
-e-mail: viroloji@thk.gov.tr

İl Bölgeleri:

Ulusal AIDS Doğrulama Merkezi ve Viral Hepatitler Ünitesi
Tel No: (0312) 565 55 53

Ulusal Difteri ve Hastalıklar
Tel No: (0312) 565 55 52

Ulusal İnfluenza Merkezi ve S
Tel No: (0312) 565 55 82

Ulusal Arbovirus ve Viral Zec
Tel No: (0312) 565 55 47

Ulusal Viral Kaynaklı Gastro
Tel No: (0312) 565 55 40

VİROLOJİK KRATİSTİK ANALİZLERİ					
İHTİ / TIBBİ KODU	ANALİZLER	YÖNTEM	MİKTAR	ANALİZ SÜRESİ	ANALİZ ÜCRETİ ₺
120.344	West Nile Virus (WNV) (WNV Antijen) (EIA) Analizi	EIA	4 ml, 3 test		50
120.346	West Nile Virus (WNV) (WNV Antijen) (EIA) Analizi	EIA, EIA, EIA	500		50
908.728	West Nile Virus (WNV) (WNV Antijen) (EIA) Analizi (PCR)	Real time PCR, IFT	500, 4 ml, 10TCA/3 test		***
908.771	West Nile Virus (WNV) (WNV Antijen) (EIA) Analizi (RT-PCR)	Genetik, Transkripsiyon-PCR	500, 4 ml, 10TCA/3 test		***
120.372	Yellow Fever Virus (YFV) (YFV Antijen) (EIA) Analizi	EIA	4 ml, 3 test		60
120.374	Yellow Fever Virus (YFV) (YFV Antijen) (EIA) Analizi	EIA	4 ml, 3 test		60

VİRAL KONTAMİNASYON, VALİDASYON VE KALİTE KONTROLÜ KONTROLLERİ					
İHTİ / TIBBİ KODU	ANALİZLER	YÖNTEM	MİKTAR	ANALİZ SÜRESİ	ANALİZ ÜCRETİ ₺
120.376	Final (EIA) ve (PCR) ve (EIA) ve (PCR) Kontrolü		100 (EIA) ve 5 (PCR)		1400
120.378	Plazma (EIA) ve (PCR) ve (EIA) ve (PCR) Kontrolü		100 (EIA) ve 5 (PCR)		800

** - Yeterince güçlü grip vakanın poliyomavirüs, çökümlü veya bakteriyel poliyomavirüs ile ilgili yetersiz bilgi yeterince değerlendirilmelidir.

*** - 2014 Mali Yılı Sağlık Uygulama ve Denetim Sistemi (SUS) çerçevesinde 2013 yılı fiyat bilgileriyle ilgili detaylar için bakınız.

VİROLOJİK LABORATUVAR ÖRNEK ALMA - GÖNDERME KURALLARI VE ANALİZLERE SIZ ACIKLAMALAR

WNV:
West Nile Virus (WNV) (WNV Antijen) (EIA) Analizi: Analiz için örnekler 4-6 ml (EIA) ve 10 ml (PCR) olarak alınmalıdır. Örnekler 4-6 ml (EIA) ve 10 ml (PCR) olarak alınmalıdır. Örnekler 4-6 ml (EIA) ve 10 ml (PCR) olarak alınmalıdır. Örnekler 4-6 ml (EIA) ve 10 ml (PCR) olarak alınmalıdır.

YFV:
Yellow Fever Virus (YFV) (YFV Antijen) (EIA) Analizi: Analiz için örnekler 4-6 ml (EIA) ve 10 ml (PCR) olarak alınmalıdır. Örnekler 4-6 ml (EIA) ve 10 ml (PCR) olarak alınmalıdır. Örnekler 4-6 ml (EIA) ve 10 ml (PCR) olarak alınmalıdır. Örnekler 4-6 ml (EIA) ve 10 ml (PCR) olarak alınmalıdır.

Plazma:
Plazma (EIA) ve (PCR) ve (EIA) ve (PCR) Kontrolü: Analiz için örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır. Örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır. Örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır. Örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır.

Final (EIA) ve (PCR) ve (EIA) ve (PCR) Kontrolü:
Analiz için örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır. Örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır. Örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır. Örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır.

Plazma (EIA) ve (PCR) ve (EIA) ve (PCR) Kontrolü:
Analiz için örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır. Örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır. Örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır. Örnekler 100 (EIA) ve 5 (PCR) olarak alınmalıdır.



Arbovirus ve Viral Zoonotik Hastalıklar Ünitesi Ekibi

- Uzm. Dr. Dilek Yağcı Çağlayık
- Uzm. Dr. Dilek Menemenlioğlu
- Bio. Ahmet Aydemir
- Bio. Gökhan Kavuncuoğlu
- Bio. Nilgün Gökalp
- Bio. İhsan Durmaz
- Lab. Tekn. Songül Özen

Viroloji Merkez Laboratuvarı,
MRL Daire Başkanlığı,
Kurum Başkanlığına destekleri için teşekkür ederiz.

Sabrınız için teşekkür ederim.





Viral Kanamalı Ateşler Tedavide yeni ne var?

Dr. Güven ÇELEBİ

Bülent Ecevit Üniversitesi Tıp Fakültesi

Enfeksiyon Hastalıkları ve klinik Mikrobiyoloji AD

guvencelebi@yahoo.com

22 Mayıs 2014, Antalya

KKKA - Tedavi

- Destek tedavisi
 - Taze donmuş plazma
 - Trombosit süspansiyonu
 - Kan transfüzyonu
-
- Ribavirin
 - İmmünglobülin
 - İnterferon
 - Steroid

The Efficacy of Oral Ribavirin in the Treatment of Crimean-Congo Hemorrhagic Fever in Iran

M. Mardani,¹ M. Keshtkar Jahromi,¹ K. Holakouie Naieni,²
and M. Zeinali³

Table 2. Absolute and relative frequency of outcome for 187 Iranian patients with suspected Crimean-Congo hemorrhagic fever, 1999–2001.

Patient group	No. (%) of patients	
	Who died	Total
Treated with ribavirin	42 (30.2)	139 (74.7)
Historical control	22 (45.8)	48 (25.7)
Total	64 (34.2)	187 (100)

NOTE. Relative risk, 0.66; 95% CI, 0.45–0.98; χ^2 , 3.87 ($P < .05$).

Characteristics of Patients with Crimean-Congo Hemorrhagic Fever in a Recent Outbreak in Turkey and Impact of Oral Ribavirin Therapy

Önder Ergönül, Aysel Çelikbaş, Başak Dokuzoğuz, Şebnem Eren, Nurcan Baykam, and Harika Esener

Infectious Diseases and Clinical Microbiology Department, Ankara Numune Education and Research Hospital, Ankara, Turkey

We describe the epidemiological, clinical findings and the role of ribavirin therapy in patients who received a diagnosis of Crimean-Congo hemorrhagic fever (CCHF). All patients had immunoglobulin M antibodies and/or PCR results positive for CCHF virus in blood or tissue specimens. Eighty-six percent of the patients were considered to have severe cases of CCHF. The overall case-fatality rate was 2.8%. **Eight patients were given ribavirin, and all 8 survived.** We suggest using ribavirin to treat patients with CCHF, particularly those with severe cases.

Kontrol	Ribavirin
Ribavirin almayan ve ciddi seyirli 22 hastada fatalite % 4,5	Ribavirin alan 8 hastada fatalite yok

Crimean-Congo hemorrhagic fever in Southeast of Iran

Roya Alavi-Naini^{a,*}, Ali Moghtaderi^b, Hamid-Reza Koohpayeh^a,
Batool Sharifi-Mood^a, Mohammad Naderi^a, Malyhe Metanat^a,
Morteza Izadi^a

Table 2 Absolute and relative frequency of outcome for 255 Iranian CCHF patients

Patient group	Number of patients (%)		Total
	Recovery	Death	
Treated with ribavirin	199 (84.3%)	37 (15.7%)	236 (92.5%)
No treatment	7 (36.8%)	12 (63.2%)	19 (7.5%)
Total	206 (80.8%)	49 (19.2%)	255 (100%)

RR=0.25; 95% CI (0.16-0.39); $\chi^2=25.54$; $P=0.000$.

ARTICLE

The role of ribavirin in the therapy of Crimean-Congo hemorrhagic fever: early use is promising

N. Tasdelen Fisgin • O. Ergonul • L. Doganci • N. Tulek

	Erken Ribavirin tedavisi	Geç Ribavirin tedavisi	Kontrol Grup Ribavirin almıyor
	Semptom başladıktan sonraki ilk 4 gün içinde	Semptom başladıktan 5 gün ve sonrasında	
FATALİTE	1/21 (%5)	2/20 (%10)	3/11 (%27)
	P: 0,052 		
	P: 0,067 		

Crimean-Congo hemorrhagic fever in Southeast of Iran

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	P: 0,052 		
	P: 0,067 		



Crimean-Congo hemorrhagic fever in Eastern Turkey: clinical features, risk factors and efficacy of ribavirin therapy

Zulal Ozkurt^{a,*}, İlhami Kiki^b, Serpil Erol^a, Fuat Erdem^b, Neziha Yılmaz^c, Mehmet Parlak^a, Mehmet Gundogdu^b, Mehmet A. Tasyaran^a

Results: In multivariate analysis, farming (OR, 11.4), living in a rural area (OR, 10.05) and being bitten by tick (OR, 6.75) were determined as risk factors for CCHF. The rates of fever during hospitalization, confusion, neck stiffness, bleeding from multiple sites, and presence of petechia/ecchymosis were higher in the patients who died than in surviving ones. Additionally, the mean values of ALT, AST, LHD, CK, PTT, INR and urea were also higher, and the mean PLT count was lower in the patients who died. Mean recovery time was shorter in the cases treated with ribavirin than those of control. But, the need for blood and blood product, mean hospitalization duration, fatality rates, and hospital expenditure values were not significantly different between the group of patients treated with ribavirin and control groups.



Crimean-Congo hemorrhagic fever in Eastern Turkey: clinical features, risk factors and efficacy of ribavirin therapy

**Ribavirin alan grupta iyileşme süresi daha kısa.
Ancak mortalite açısından fark yok**

Res (0.05) and being bitten by tick (OR, 6.75) were determined as risk factors for CHF. The rates of fever during hospitalization, confusion, neck stiffness, bleeding from multiple sites, and presence of petechia/ecchymosis were higher in the patients who died than in surviving ones. Additionally, the mean values of ALT, AST, LHD, CK, PTT, INR and urea were also higher, and the mean PLT count was lower in the patients who died. Mean recovery time was shorter in the cases treated with ribavirin than those of control. But, the need for blood and blood product, mean hospitalization duration, fatality rates, and hospital expenditure values were not significantly different between the group of patients treated with ribavirin and control groups.

Efficacy of oral ribavirin treatment in Crimean-Congo haemorrhagic fever: A quasi-experimental study from Turkey

Journal of Infection (2009) 58, 238–244

Nazif Elaldi ^a, Hurrem Bodur ^b, Sibel Ascioğlu ^{c,d,*}, Aysel Celikbas ^b, Zulal Ozkurt ^e, Haluk Vahaboglu ^f, Hakan Leblebicioglu ^g, Neziha Yilmaz ^h, Aynur Engin ^a, Mehmet Sencan ⁱ, Kemalettin Aydin ^j, Ilyas Dokmetas ^a, Mustafa Aydin Cevik ^b, Basak Dokuzoguz ^b, Mehmet Ali Tasyaran ^e, Recep Ozturk ^k, Mehmet Bakir ^a, Ramazan Uzun ^h

- **2003 yılında Ribavirin almayan 92 hasta,**
- **2004 yılında Ribavirin alan 126 hasta ile kıyaslanıyor**

Efficacy of oral ribavirin treatment in Crimean-Congo haemorrhagic fever: A quasi-experimental study from Turkey

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Nazif Elaldi ^a, Hurrem Bodur ^b, Sibel Ascioğlu ^{c,d,*}, Aysel Celikbas ^b, Zulal Ozkurt ^e, Haluk Vahaboglu ^f, Hakan Leblebicioglu ^g, Neziha Yilmaz ^h, Aynur Engin ^a, Mehmet Sencan ⁱ, Kemalettin Aydin ^j, Ilyas Dokmetas ^a, Mustafa Aydin Cevik ^b, Basak Dokuzoguz ^b, Mehmet Ali Tasyaran ^e, Recep Ozturk ^k, Mehmet Bakir ^a, Ramazan Uzun ^h

- 2003 yılında Ribavirin almayan 92 hasta,
- 2004 yılında Ribavirin alan 126 hasta ile kıyaslanıyor

	Ribavirin Grubu N: 126	Kontrol Grubu N: 92	p
Hastanede yatış süresi	8 gün	9 gün	0,148
Fatalite	9 (%7,1)	11 (%11,9)	0,243

LETTERS TO THE EDITOR

DEBATE (see Elaldi N et al, Efficacy of oral ribavirin treatment in Crimean-Congo haemorrhagic fever: A quasi-experimental study from Turkey. *Journal of Infection* 2009; 58: 238–244)

Biases and misinterpretation in the assessment of the efficacy of oral ribavirin in the treatment of Crimean-Congo hemorrhagic fever

Dear editor,

In March 2009 issue of the journal a study by Elaldi et al. was published on the efficacy of oral ribavirin.¹ The study reported ribavirin as not an efficacious, and even harmful drug in the treatment of Crimean-Congo hemorrhagic fever (CCHF), and stressed the necessity of a randomized controlled trial (RCT) to assess the efficacy of the ribavirin in CCHF infected patients. Such a conclusion was different from the previous reports on the efficacy of ribavirin in CCHF. In previous reports, almost all the authors concluded their reports by stressing that the ribavirin was found to be beneficial, although statistically no significance was reached. Therefore, it was interesting to read such a report in the journal, which has different conclusions. However, there were inconsistencies between the conclusions and the results of the report. The comments of the authors at the discussion were not tied to their results or they were contradictory.

Sir

I read the article by Elaldi N *et al.* appeared on *Journal of Infection* 2009 Mar;58(3):238–44 which is titled "*Efficacy of oral ribavirin treatment in Crimean-Congo haemorrhagic fever: a quasi-experimental study from Turkey*"¹ with a great concern. First of all, ethically, I assume there should be a declaration of "conflict of interest" at the end of the article according to the Committee on Publishing Ethics (COPE) and World Association of Medical Editors

Response to Ergonul: Scientific Evidence Versus Personal Beliefs In Crimean-Congo Haemorrhagic Fever Treatment

Dear editor,

We read Dr Ergonul's detailed letter with great interest, however, it is impossible to agree with any of his comments due to the reasons below.

P-value and sample size are completely different things than the author thinks

We would like to refer Dr. Ergonul to some basic statistics textbooks because he has a misunderstanding of the logic of calculation of sample size and power.¹⁻³ Using a statistical software does not replace theoretical knowledge, nowadays, any statistical software can easily crunch some numbers as a sample size or power whenever you provide the necessary information. However, a wrong interpretation can lead to misleading conclusions like in this letter. First, sample size is not the necessary number that would "make" a finding statistically significant as used in this letter.^{1,2} In our study, effect size that is the difference between the mortality rates of ribavirin users and non-users was 4.8% (7.1% among ribavirin users vs. 11.9% among non-users, p -value = 0.24). This p -value shows that the difference between the mortality rates is more likely to be due to chance rather than due to the effect of ribavirin.³ Of course, increasing the sample size can make this or any other result statistically significant! What if the results were 11.9% vs. 12.0%, then it is possible to calculate 32,350 patients as the necessary sample size to reach statistical significance and therefore the author can still claim that ribavirin was beneficial and sample size was insufficient. According to his logic, for each and every study in the literature with a non-significant p -value, a sample size that would lead to a significant p -value can be calculated and the study can be labeled as having "insufficient" sample size. Obviously, this is the wrong way of thinking and using sample size calculations, it is neither the p -value, nor the sample size but the effect size that matters most in clinical

- **Randomize, kontrollü çalışma!**



The efficacy of ribavirin in the treatment of Crimean-Congo hemorrhagic fever in Eastern Black Sea region in Turkey

Iftihar Koksak*, Gurdal Yilmaz, Firdevs Aksoy, Hava Aydin, Ilknur Yavuz, Serap Iskender, Korhan Akcay, Sukru Erensoy, Rahmet Caylan, Kemalettin Aydin

Karadeniz Technical University School of Medicine, Department of Infectious Disease and Clinical Microbiology, 61080 Trabzon, Turkey

Study design: In this prospective randomized cohort study 136 cases were included between June 2004 and August 2007. The diagnosis was confirmed in the CCHF reference laboratory of Refik Saydam National Hygiene Central Institute of the Turkish Ministry of Health. Patients either received ribavirin plus supportive treatment (Group A) (n=64) or no specific treatment (Group B) (n=72). For the evaluation of efficacy

Prospektif, Randomize

Results: There were no statistical differences in incubation time; hospitalization time; patients requiring platelet replacement therapy; the time taken for platelet levels to return to normal levels and mortality. In Group B, the rate of tick contact was higher ($p=0.03$). In Group A, leukocyte levels took longer to return to the normal levels ($p=0.02$).

Conclusion: In our study, there was no positive effect determined on clinical or laboratory parameters in CCHF patients treated with ribavirin, also it was observed that leukocyte levels took longer to return



	Ribavirin Grubu N: 64	Kontrol Grubu N: 72
İnkübasyon süresi	5,5 gün	4,9 gün
Hastaneye başvuru süresi	4,5 gün	3,9 gün
Fatalite	4 (%6,3)	4 (%5,6)

Study design: In this prospective randomized cohort study 136 cases were included between June 2004 and August 2007. The diagnosis was confirmed in the CCHF reference laboratory of Refik Saydam National Hygiene Central Institute of the Turkish Ministry of Health. Patients either received ribavirin plus supportive treatment (Group A) (n=64) or supportive treatment (Group B) (n=72). For the evaluation of efficacy of ribavirin, patients were divided into two groups: Group A (n=64) and Group B (n=72).

Results: There were no statistical differences in incubation time; hospitalization time; patients requiring platelet replacement therapy; the time taken for platelet levels to return to normal levels and mortality. In Group B, the rate of tick contact was higher ($p=0.03$). In Group A, leukocyte levels took longer to return to the normal levels ($p=0.02$).

Conclusion: In our study, there was no positive effect determined on clinical or laboratory parameters in CCHF patients treated with ribavirin, also it was observed that leukocyte levels took longer to return

Prospektif, Randomize

RESEARCH ARTICLE

Open Access

Ribavirin for Crimean-Congo hemorrhagic fever: systematic review and meta-analysis

Karla Soares-Weiser*¹, Sherine Thomas², Gail Thomson G³ and Paul Garner⁴

Results: 21 unique studies, including one randomised controlled trial of ribavirin, were included. Quality of the evidence was very low, with a Down and Black median score of 4 (maximum possible 33). Ribavirin treatment was not shown to be superior to no ribavirin treatment for mortality rate in a single RCT (RR: 1.13, 95%CI: 0.1 to 4.32, 136 participants, GRADE=low quality evidence) but ribavirin was associated with reduced mortality when compared to no ribavirin treatment (RR: 0.5, 95%CI: 0.1 to 2.1, 136 participants; GRADE=very low quality evidence). No adverse events were reported.

**21 çalışma inceleniyor.
Ribavirin tedavisi daha üstün değil.**

Conclusions: No clear message of benefit is available from the current data on ribavirin as observational data are heavily confounded, and the one trial carried out has limited power. However, ribavirin could potentially have benefits in this condition and these results clearly indicate a pragmatic, randomised controlled trial in the context of good quality supportive care, is urgently needed and ethically justified.

Ribavirin for patients with Crimean–Congo haemorrhagic fever: a systematic review and meta-analysis

Sibel Ascioğlu^{1,2*}, Hakan Leblebicioğlu³, Haluk Vahaboglu⁴ and K. Arnold Chan²

¹Department of Infection Diseases, ²Department of Clinical Microbiology and Infectious Diseases, ³Department of Clinical Microbiology and Infectious Diseases, ⁴Department of Clinical Microbiology and Infectious Diseases, Ankara, Turkey; ²Department of Clinical Microbiology and Infectious Diseases, ³Department of Clinical Microbiology and Infectious Diseases, ⁴Department of Clinical Microbiology and Infectious Diseases, Ankara, Turkey

**8 çalışma inceleniyor.
Ribavirin tedavisi daha üstün değil.**

Results: One randomized controlled trial and seven observational studies met our inclusion criteria. Most observational studies suffered from different types of bias due to inappropriate selection of controls. Compilation of data from all included studies showed that ribavirin did not improve survival in CCHF (relative risk 1.06, 95% confidence interval 0.97–1.16). Analysis of secondary endpoints did not suggest a clinically significant beneficial effect either.

Conclusions: Our systematic review and meta-analysis revealed that the available data in the literature are inadequate to support a claim of efficacy of ribavirin in CCHF. We believe a real uncertainty exists over the benefit of ribavirin in the treatment of CCHF, which necessitates the urgent conduct of a randomized placebo-controlled trial.



ELSEVIER

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journal

Table 1

CCHF-related mortality rate and ribavirin use in Turkey, 2004–2007

Year	Total number of CCHF patients	Number of patients using ribavirin	Mortality rate
2004	249	169 (67.9%)	5.2%
2005	266	58 (21.8%)	4.9%
2006	438	71 (16.2%)	6.2%
2007	717	85 (11.8%)	4.6%

Perspective

Ribavirin is not effective against Crimean–Congo hemorrhagic fever: observations from the Turkish experience[☆]



Bahadır Ceylan, Aylin Calıca, Oznur Ak, Yasemin Akkoyunlu^{*}, Vedat Turhan

Faculty of Medicine, Department of Infectious Diseases and Clinical Microbiology, Bezmialem Vakıf University, Vatan Cad., Fatih 34093, Istanbul, Turkey

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SUMMARY

Crimean–Congo hemorrhagic fever (CCHF) is a viral infection associated with a high mortality rate. Ribavirin is the only drug used in the treatment of this disease. Studies investigating the effectiveness of ribavirin in CCHF have been retrospective and to date have included only a small number of cases. In recent years, due to climate changes, the number of cases of CCHF in Turkey has increased, and experience in the treatment of CCHF has improved. Several studies have evaluated the efficacy of ribavirin in Turkey, including one randomized controlled trial and two studies with a large number of cases. **In these studies, ribavirin therapy was not shown to decrease mortality rates; the mortality rate was 2–9% in patients treated with ribavirin and 5.6–11% in those who were not treated with this drug.** These findings suggest that patients with CCHF should be followed with supportive care only until randomized controlled trials with larger groups have been conducted.

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Short Communication

Prompt Administration of Crimean-Congo Hemorrhagic Fever (CCHF) Virus Hyperimmunoglobulin in Patients Diagnosed with CCHF and Viral Load Monitorization by Reverse Transcriptase-PCR

Ayhan Kubar*, Mustafa Haciomeroglu¹, Aykut Ozkul², Umit Bagriacik³,
Esragul Akinci⁴, Kenan Sener, and Hurrem Bodur⁴

*Gulhane Military School of Medicine, Ankara; ¹Refik Saydam Hygiene Center, Ankara;
²Ankara University Ankara; ³Gazi University Ankara; and*

SUMMARY: Crimean-Congo hemorrhagic fever virus (CCHFV), a member of the genus *Nairovirus* of the family *Bunyaviridae*, causes a severe disease in humans with high mortality rates. In Turkey, the number of patients with CCHF has increased since 2002. Here, we aimed to treat CCHF patients with CCHFV hyperimmunoglobulin. We prepared a CCHFV hyperimmunoglobulin product from 22 individuals who survived CCHF infection. A total of 26 CCHF patients were enrolled into this study. For CCHFV hyperimmunoglobulin administration, a Kubar Unit (KU) was defined. As a standard therapeutic approach, 400 KU of hyperimmunoglobulin were given to each patient as a single dose before viral load was detected. We used one-step real-time reverse transcriptase-PCR to monitor the viral load of CCHF patients. According to the one-step real-time PCR results, 15 patients with a viral load of 10^8 copies/mL or more were defined as high risk. In this high-risk group, the survival rate was found to be 86.6% (13/15) and 2 patients died despite CCHFV hyperimmunoglobulin administration. CCHF is a very serious and highly fatal infection, particularly for patients in the defined high-risk group. Prompt administration of CCHFV hyperimmunoglobulin might be a very promising new treatment approach, especially for high-risk individuals.

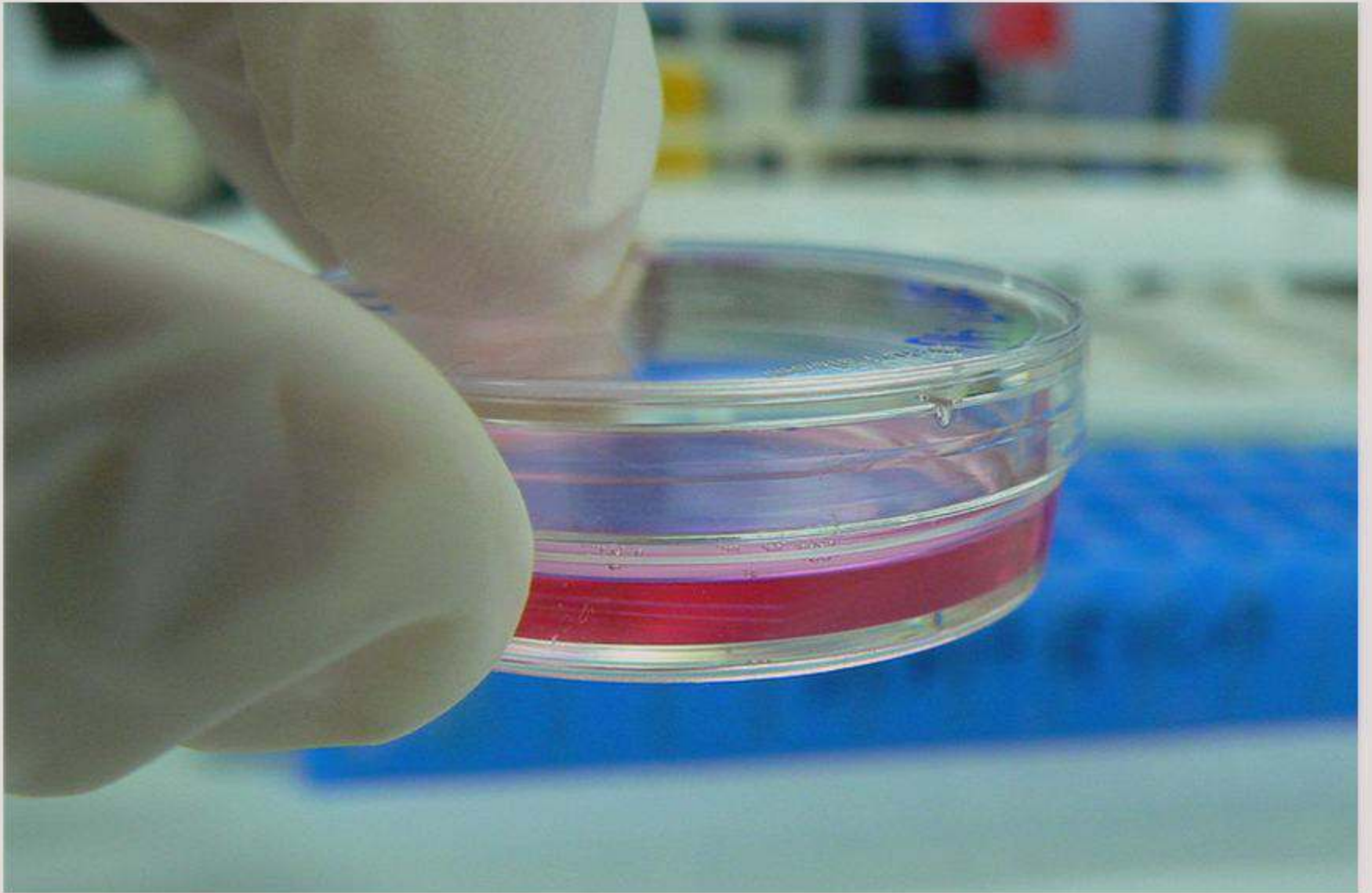
Short Communication

Prompt Administration of Crimean-Congo Hemorrhagic Fever (CCHF) Virus Hyperimmunoglobulin to Patients Diagnosed with CCHF and Viral Load Monitoring

**Yüksek riskli 15 hasta
Spesifik immünglobulin veriliyor.
2 hasta ex.**

Kontrol grubu yok!

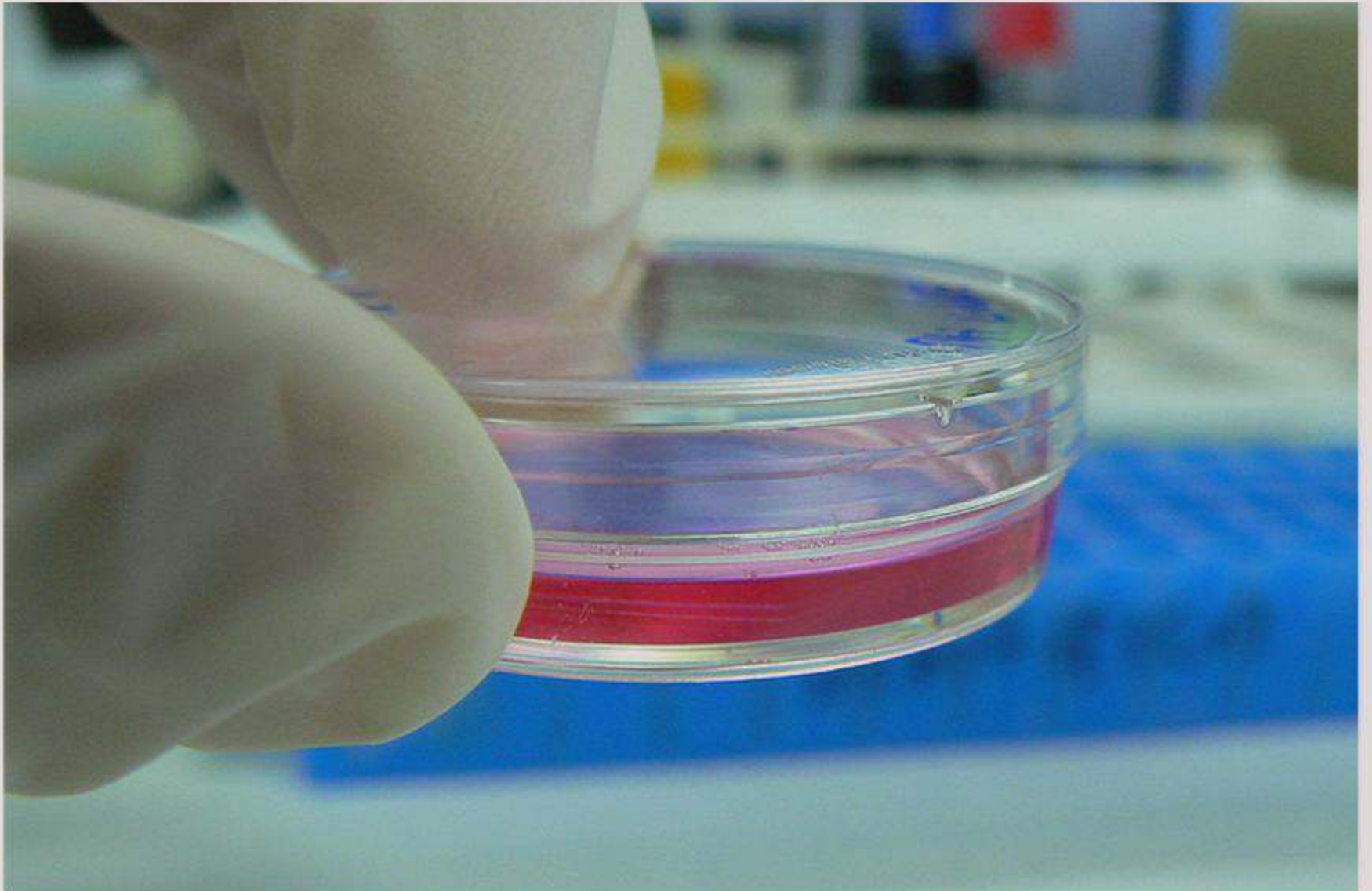
SUMMARY: Crimean-Congo hemorrhagic fever virus (CCHFV) of the family *Bunyviridae*, causes a severe disease in humans with high mortality. In Turkey, the number of patients with CCHF has increased since 2002. Here, we aimed to treat patients with CCHFV hyperimmunoglobulin. We prepared a CCHFV hyperimmunoglobulin product from 22 individuals who survived CCHF infection. A total of 26 CCHF patients were enrolled into this study. For CCHFV hyperimmunoglobulin administration, a Kubar Unit (KU) was defined. As a standard therapeutic approach, 400 KU of hyperimmunoglobulin were given to each patient as a single dose before viral load was detected. We used one-step real-time reverse transcriptase-PCR to monitor the viral load of CCHF patients. According to the one-step real-time PCR results, 15 patients with a viral load of 10^8 copies/mL or more were defined as high risk. In this high-risk group, the survival rate was found to be 86.6% (13/15) and 2 patients died despite CCHFV hyperimmunoglobulin administration. CCHF is a very serious and highly fatal infection, particularly for patients in the defined high-risk group. Prompt administration of CCHFV hyperimmunoglobulin might be a very promising new treatment approach, especially for high-risk individuals.



The Treatment of Crimean-Congo Hemorrhagic Fever With High-dose Methylprednisolone, Intravenous Immunoglobulin, and Fresh Frozen Plasma

Erol Erduran, MD, Aysenur Bahadir, MD,* Nil Palanci, MD,† and Yusuf Gedik, MD†*

- **Pediatric hasta grubu, KTÜ, Trabzon**
- **2010 -2011 yıllarında 12 çocuk hasta**
 - **Yüksek doz metilprednizolon**
 - **IVIG**
 - **Taze donmuş plazma ile tedavi edildi.**
- ***Sonuç başarılı, ancak çalışmada kontrol grubu yok.***



Evaluation of Antiviral Efficacy of Ribavirin, Arbidol, and T-705 (Favipiravir) in a Mouse Model for Crimean-Congo Hemorrhagic Fever



Lisa Oestereich^{1,2,3}, Toni Rieger^{1,2,3}, Melanie Neumann³, Christian Bernreuther⁴, Maria Lehmann^{1,2}, Susanne Krasemann³, Stephanie Wurr^{1,2}, Petra Emmerich^{1,2}, Xavier de Lamballerie⁵, Stephan Ölschläger^{1,2,3}, Stephan Günther^{1,2,3*}

1 Department of Virology, Bernhard-Nocht-Institute for Tropical Medicine, Hamburg, Germany, 2 German Centre for Infection Research (DZIF), Hamburg, Germany, 3 Mouse Pathology Core Facility, University Medical Center Hamburg-Eppendorf, Hamburg, Germany, 4 Institute of Neuropathology, University Medical Center Hamburg-Eppendorf, Hamburg, Germany, 5 Aix Marseille Université, IRD French Institute of Research for Development, EHESP French School of Public Health, UMR_D 190 "Emergence des Pathologies Virales", Marseille, France

- **Transgenik fare modelinde**

- Ribavirin
- Arbidol
- Favipiravir etkinliğinin araştırılması



Metod

- Fareler intraperitoneal (i.p.) yolla KKKA virüsü ile enfekte edildi.
- İnfekte edilen farelere etken maddeler **8 gün** süreyle uygulandı
- Hastalık belirti ve bulguları yönünden **14 gün** izlendi
 - Ciddi hastalık belirtisi (*kanama, abdominal distansiyon, konvülzyon, ishal, agoni, 2 gün içinde %15 üzerinde kilo kaybı*) görülen fareler ekplore edilerek incelendi.
- Biyokimyasal ve virolojik incelemeler için 1-4 gün arayla periyodik venöz kan örnekleri alındı
- Deney sonunda sağ kalan fareler ekplore edilerek histopatolojik ve virolojik incelemeler yapıldı

KKKA virüsü - Letal doz

İnokulmdaki virüs miktarı FFU	Farelerden ölüm oranı (Ölen/infekte edilen)	İnokulum sonrasında yaşam süresi
0,3	4/5	
1	5/5	
3	4/5	
10	6/8	
100	13/13	3-6 gün
1 000	8/8	3-6 gün
10 000	3/3	2 gün

Ribavirin toksisite testleri

LD50 = 220 mg/kg/gün

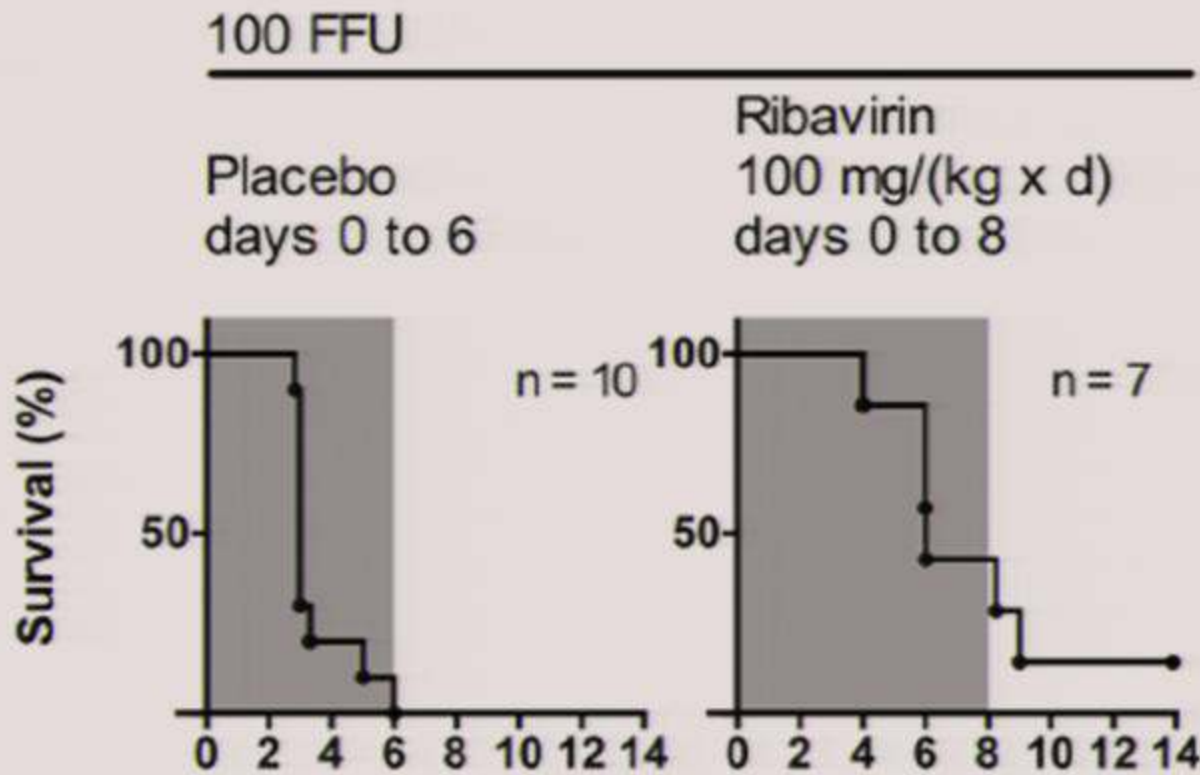
Ribavirin 100 mg/kg/gün

100 FFU virüs inoküle edildikten 1 saat sonra **intraperitoneal** (tek doz) başlandı ve 8 gün süreyle verildi

Plasebo (Serum fizyolojik)

100 FFU virüs inoküle edildikten 1 saat sonra **intraperitoneal** (tek doz) başlandı ve 8 gün süreyle verildi

	Ribavirin	Plasebo	P
Sağ kalım oranı (yaşayan/enfekte edilen)	1/7	0/10	P = 0,4
Ortalama yaşam süresi	6 gün	3 gün	p = 0.0007



p = 0.001

p = 0.006

p = 0.0007

p = 0.002

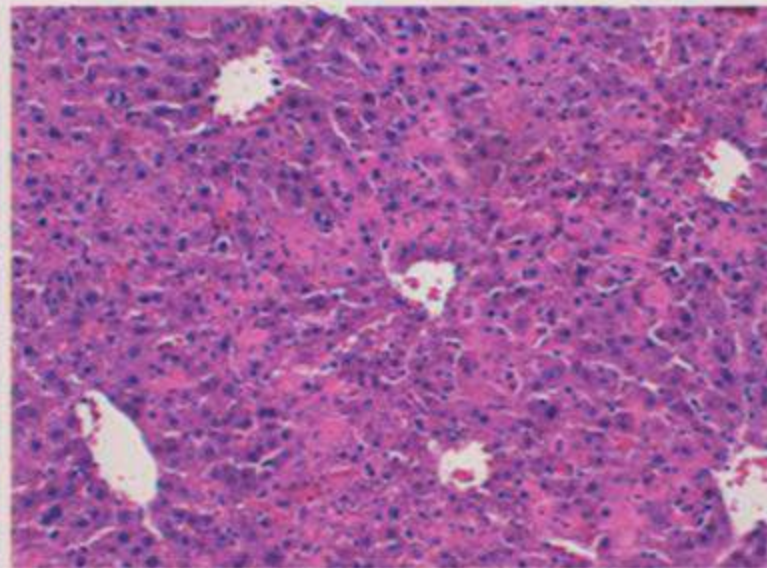
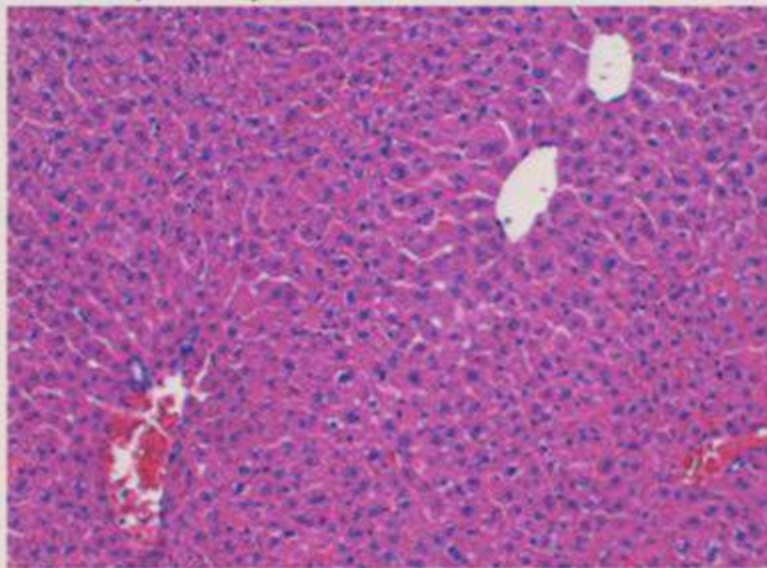
p > 0.001

	Ribavirin	Plasebo	P
Sağ kalım oranı (yaşayan/enfekte edilen)	1/7	0/10	P = 0,4
Ortalama yaşam süresi	6 gün	3 gün	p = 0.0007
AST seviyesinde azalma (2. gün)			p = 0.001
ALT seviyesinde azalma (2. gün)			p = 0.006
Serum viral yükü azalma (2. gün)			p = 0.0007
Ağırlık (3. gün)			p = 0.002
Karaciğer ve dalakta viral yük			p > 0.001

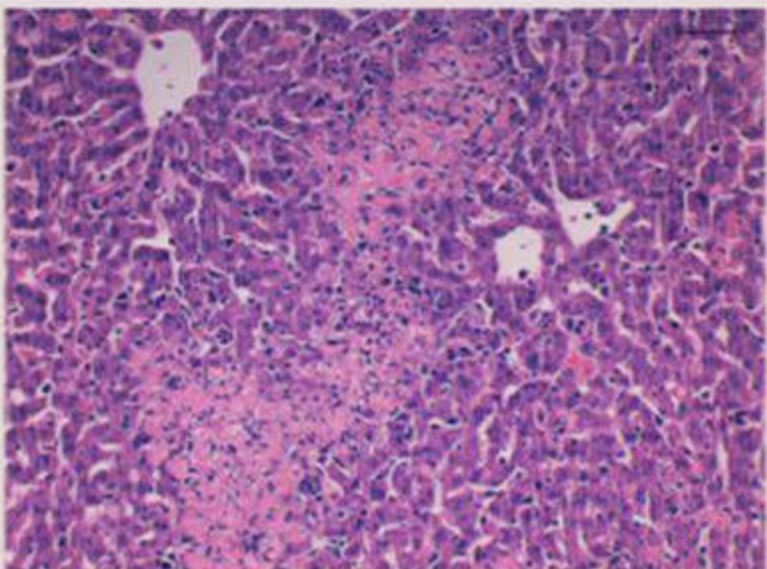
Naïve

Infected

Liver (H&E)



Infected + ribavirin



Histopatolojik inceleme

Plasebo – Ribavirin

	Ribavirin tedavisi alan fareler
3. Gün ölen fareler	Yaygın ancak küçük nekroz odakları
4- 9 gün içinde ölen fareler	Geniş köprüleşmiş hepatoselüler nekroz alanları

KKKA - Transgenik fare modelinde Ribavirin

- Virüs yükünü azalttı
- Hastalığın progresyonunu geciktirdi
- Monosit – makrofaj aktivasyonunu önleyemedi
- Terminal karaciğer nekrozunu önleyemedi
- Ölümü geciktirdi ancak önleyemedi.

KKKA - Transgenik Fare Modelinde Arbidol

Arbidol toksisite testleri

0, 25, 75, 150, 300, 600 mg/kg/gün 8 gün süreyle.
Hiçbir dozda toksik belirti yok

Arbidol 75 mg/gün

1000 FFU virüs inoküle edilmeden 1 gün önce başlandı (*tek doz*).
8 gün süreyle devam edildi (oral yoldan, mide probuyla)

Arbidol 100 mg/gün

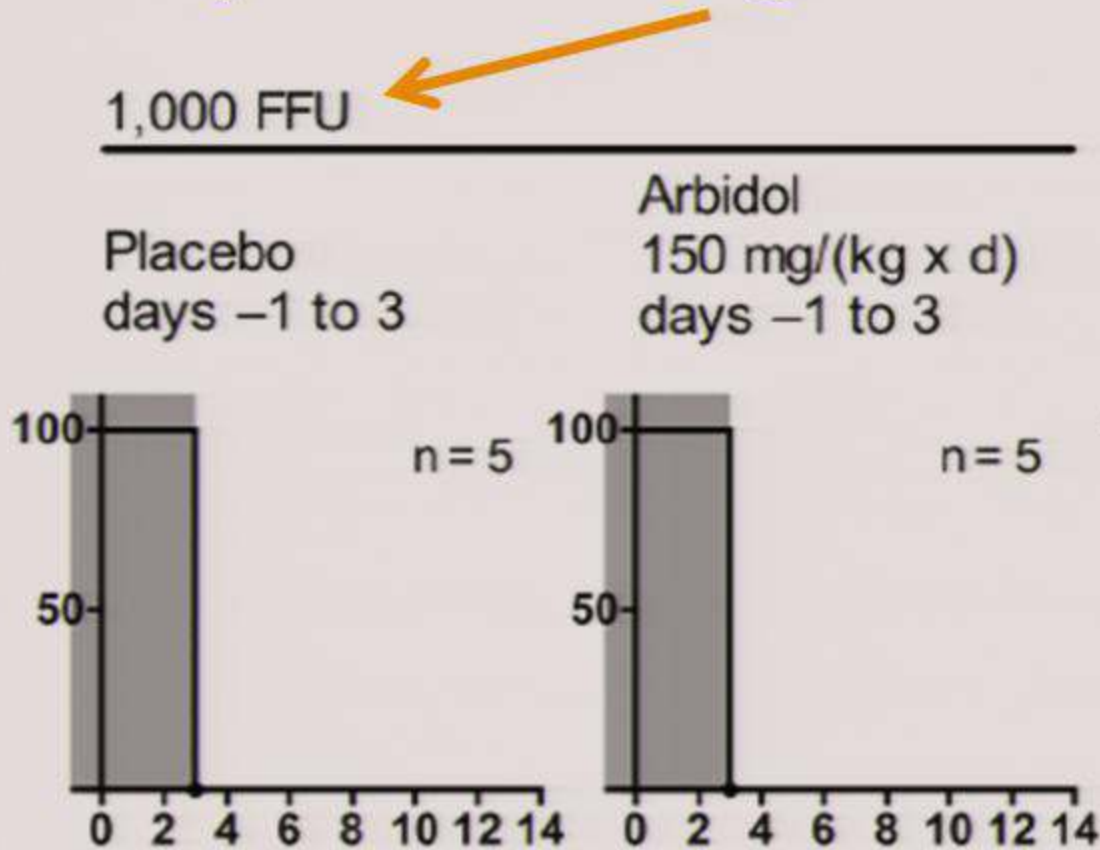
1000 FFU virüs inoküle edilmeden 1 gün önce başlandı (*tek doz*).
8 gün süreyle devam edildi (oral yoldan, mide probuyla)

Plasebo (metil selüloz)

1000 FFU virüs inoküle edilmeden 1 gün önce başlandı (*tek doz*).
8 gün süreyle devam edildi (oral yoldan, mide probuyla)

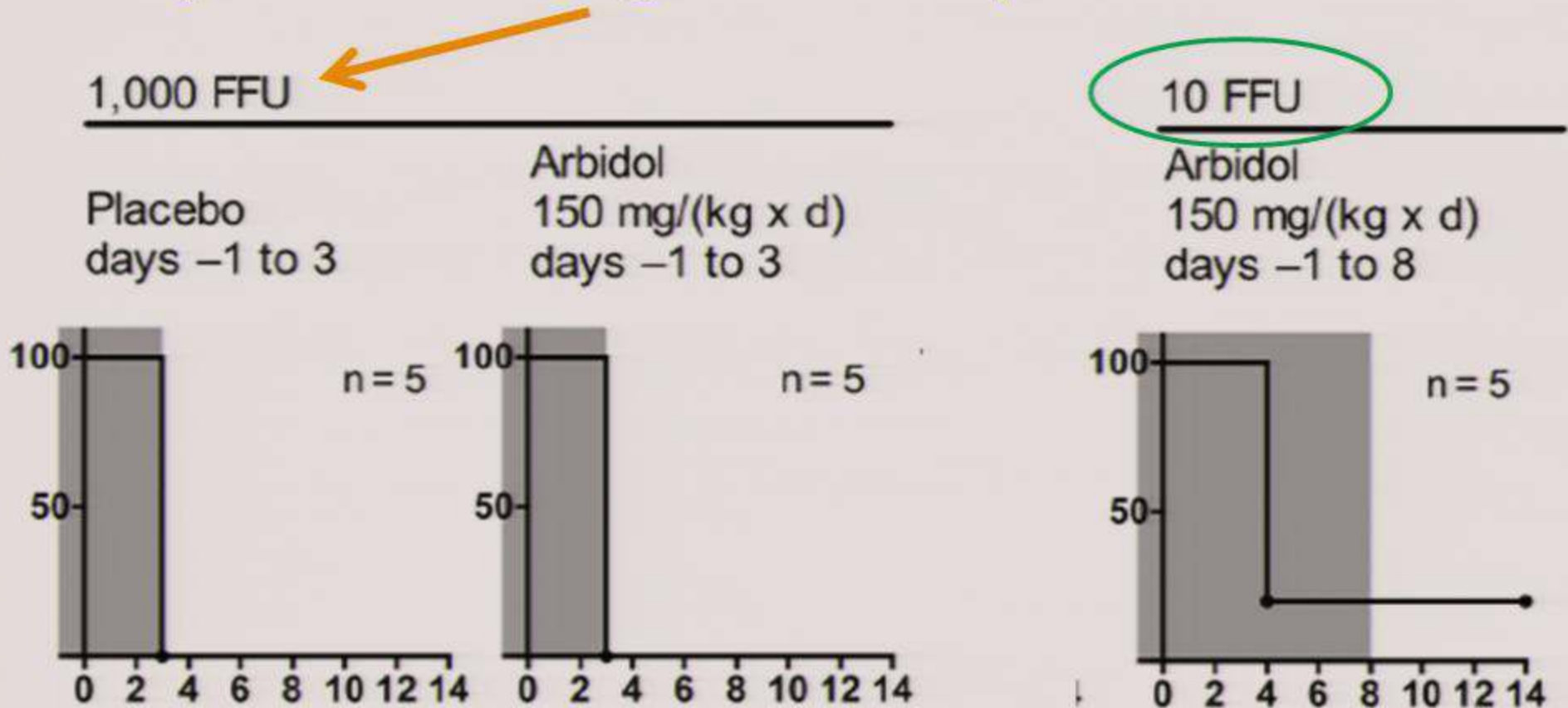
KKKA - Transgenik Fare Modelinde Arbidol

- Yaşam süresi ve Sağ kalım oranı plasebo ile benzer



KKKA - Transgenik Fare Modelinde Arbidol

- Yaşam süresi ve Sağ kalım oranı plasebo ile benzer



KKKA - Transgenik Fare Modelinde Arbidol

- Yaşam süresi ve Sağ kalım oranı plasebo ile benzer

1,000 FFU



Arbidol

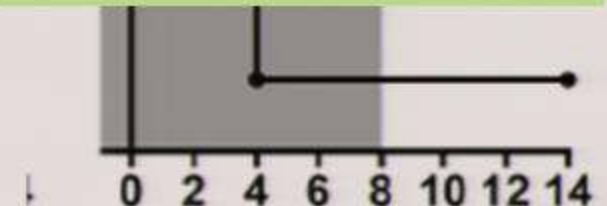
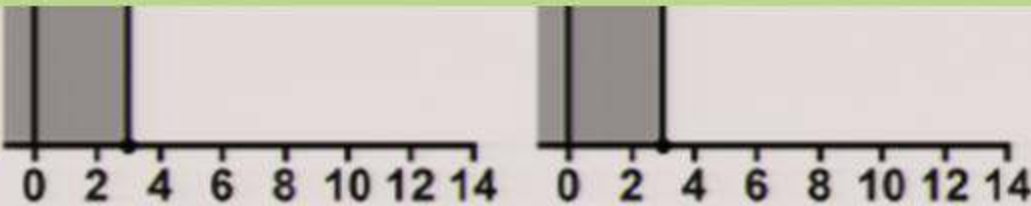
10 FFU

Arbidol

Transgenik fare modelinde

ARBİDOL

KKKA virüsüne etkili değildir

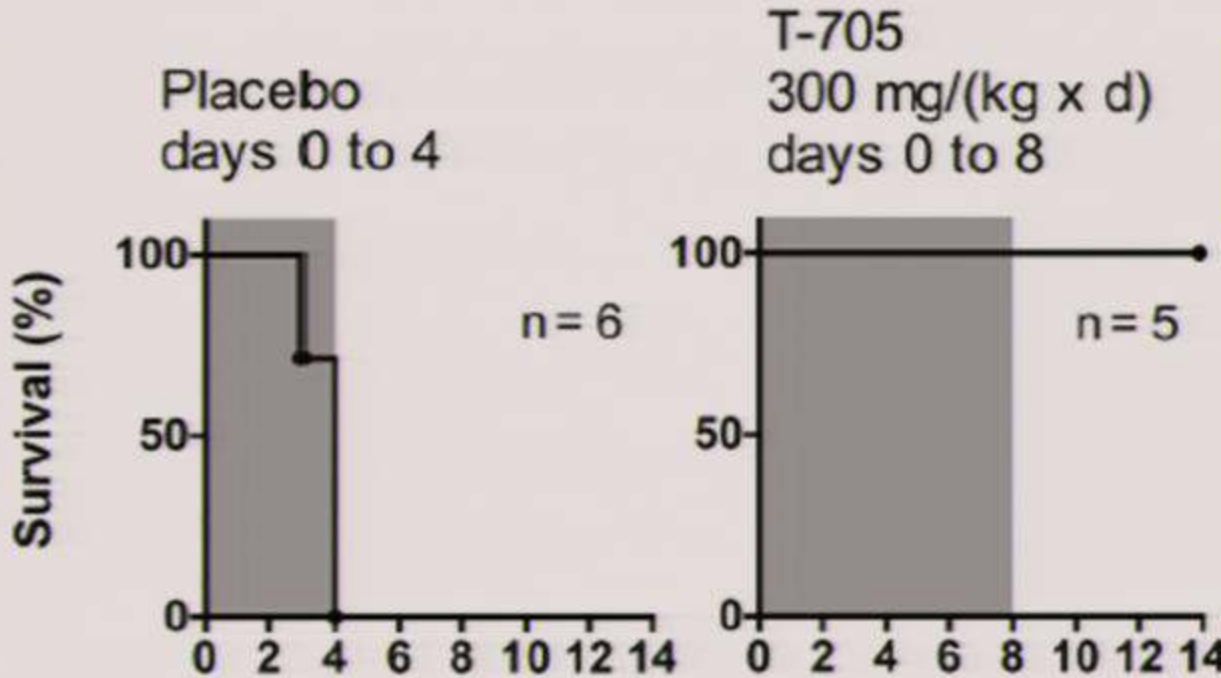


KKKA - Transgenik Fare Modelinde Favipiravir (T-705)

Favipiravir (T-705)	Favipiravir uygulama yolu
300 mg/kg/gün	300 FFU virüs inoküle edildi. Favipiravir aynı gün başlandı (iki doz). 8 gün süreyle oral yoldan (mide probuyla) verildi
Plasebo (metil selüloz)	300 FFU virüs inoküle edildi. Favipiravir aynı gün başlandı (iki doz). 8 gün süreyle oral yoldan (mide probuyla) verildi

KKKA - Transgenik Fare Modelinde Favipiravir (T-705)

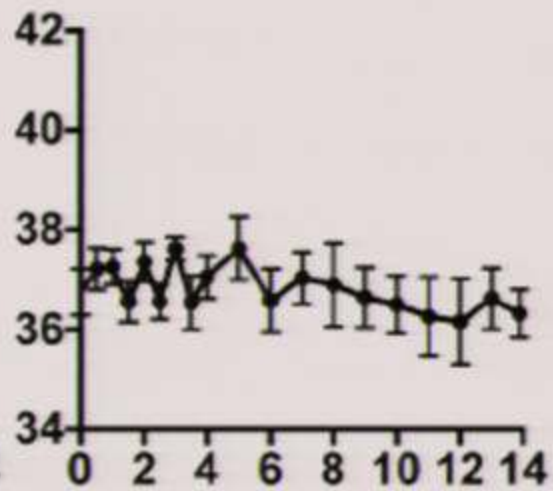
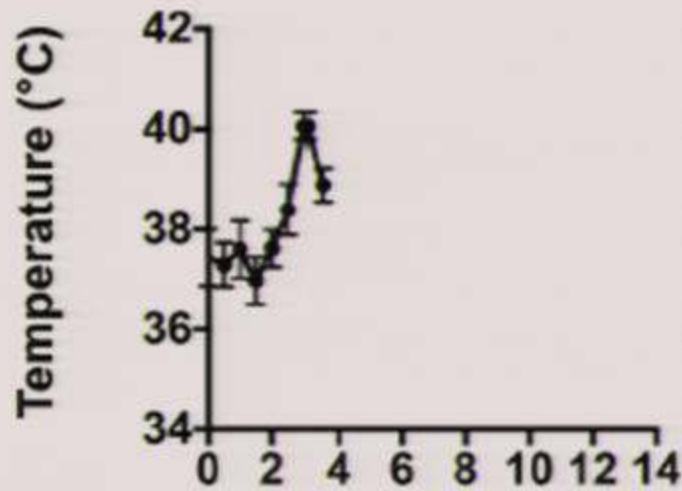
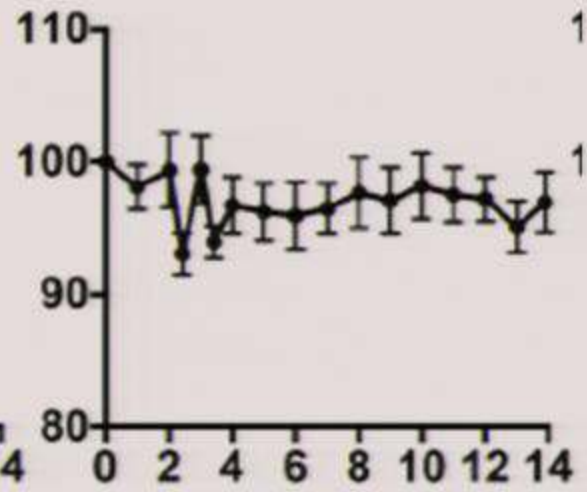
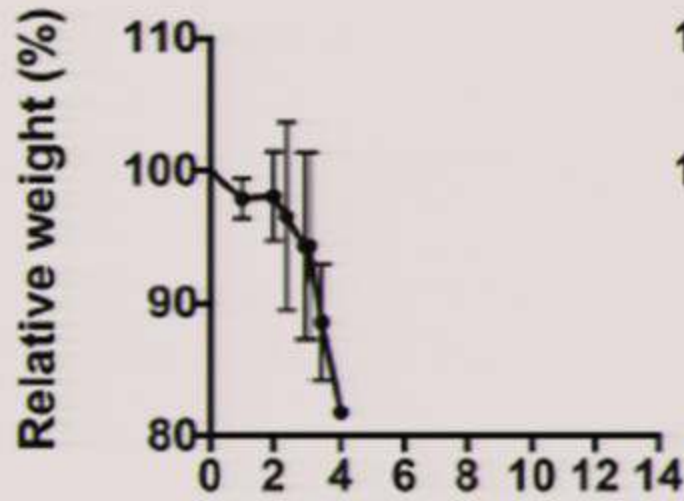
Favipiravir (T-705)	Favipiravir uygulama yolu
300 mg/kg/gün	300 FFU virüs inoküle edildi. Favipiravir aynı gün başlandı (iki doz). 8 gün süreyle oral yoldan (mide probuyla) verildi
Plasebo (metil selüloz)	300 FFU virüs inoküle edildi. Favipiravir aynı gün başlandı (iki doz). 8 gün süreyle oral yoldan (mide probuyla) verildi



Favipiravir alan farelerde
ölüm görülmedi

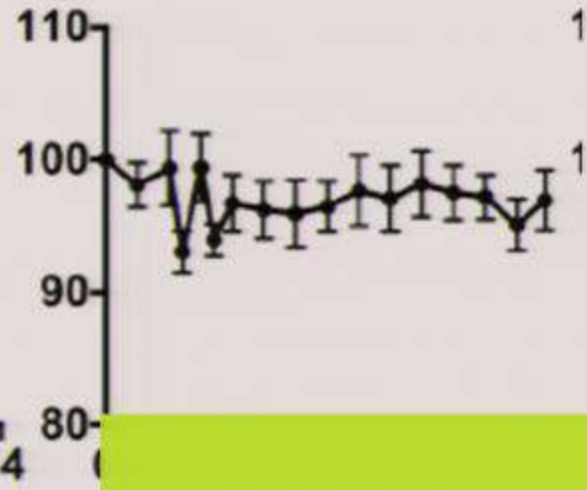
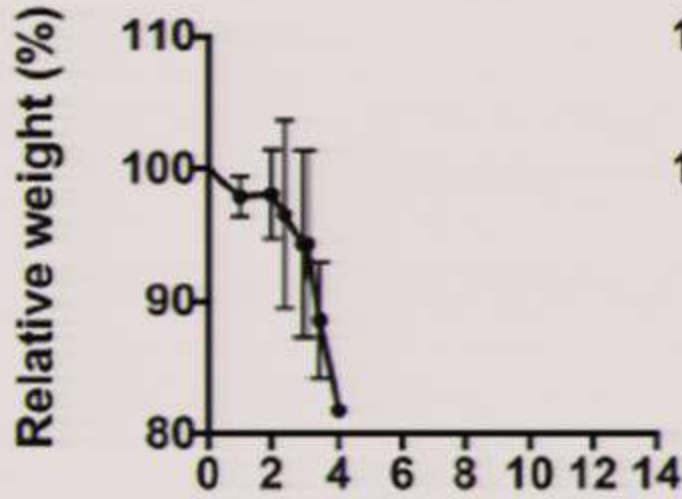
Plasebo

Favipiravir 300mg/kg/gün

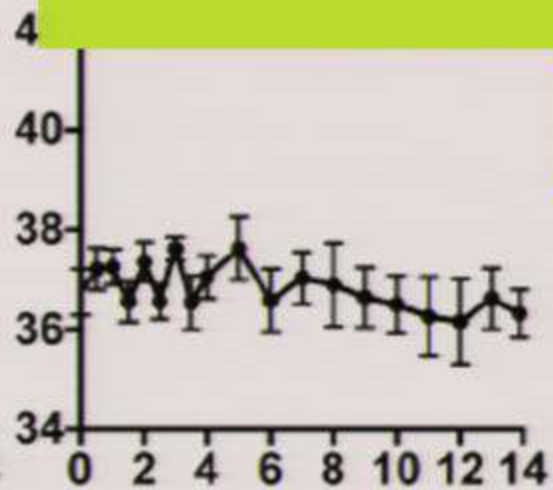
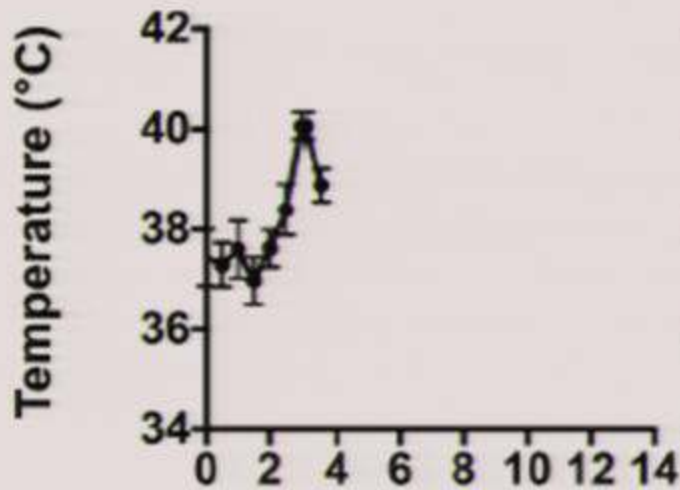


Plasebo

Favipiravir 300mg/kg/gün

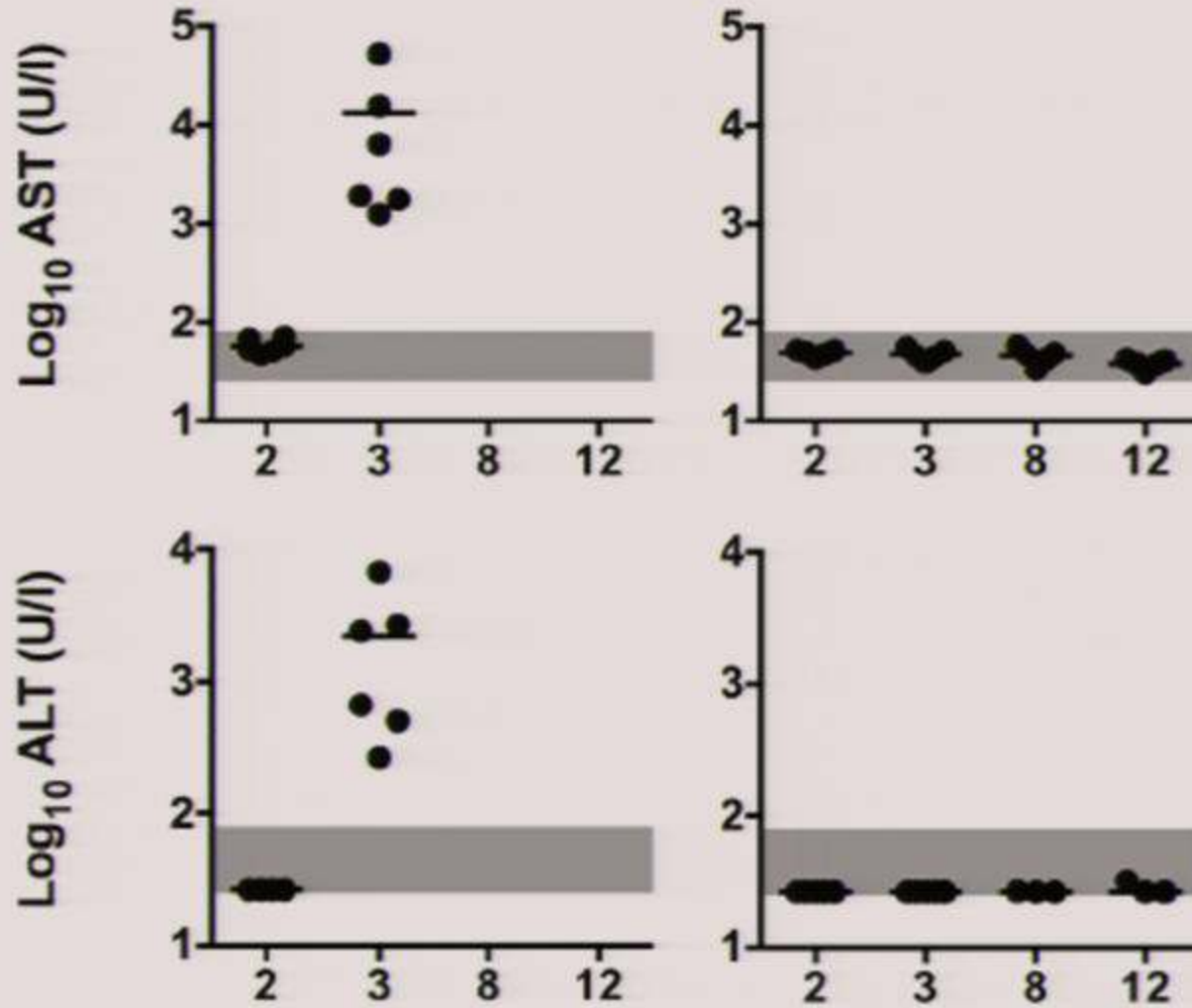


Favipiravir alan farelerde
semptom görülmedi



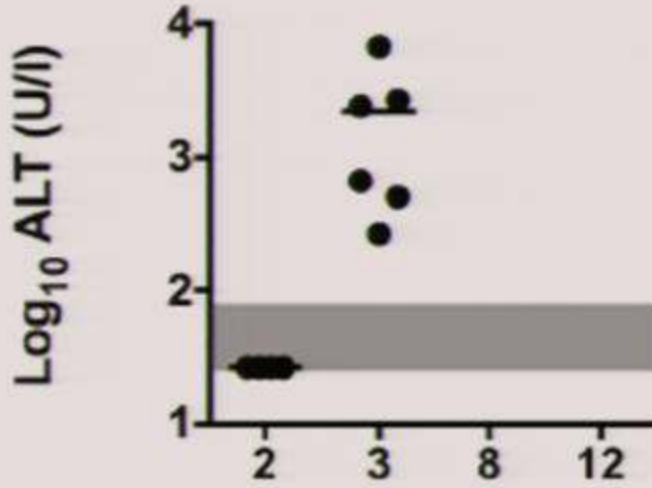
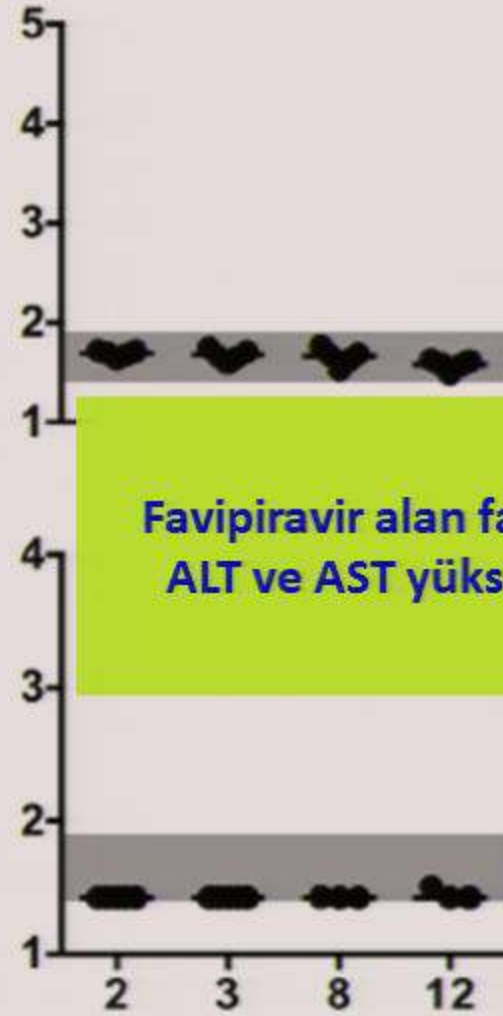
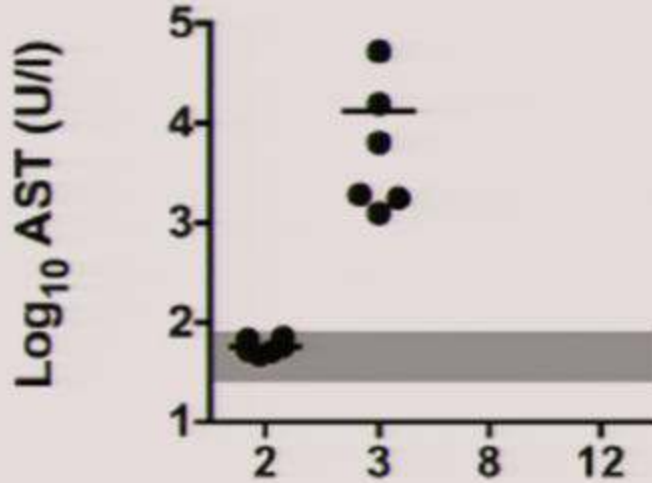
Plasebo

Favipiravir 300mg/kg/gün



Plasebo

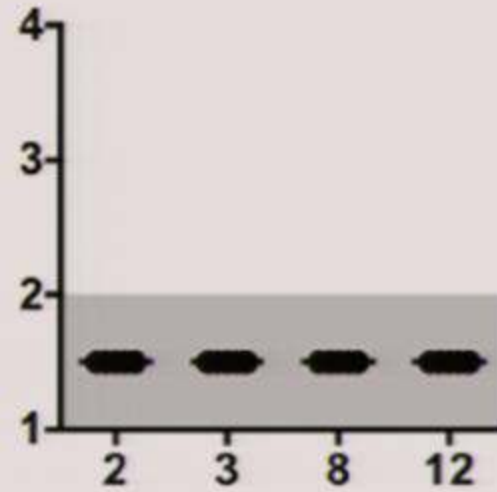
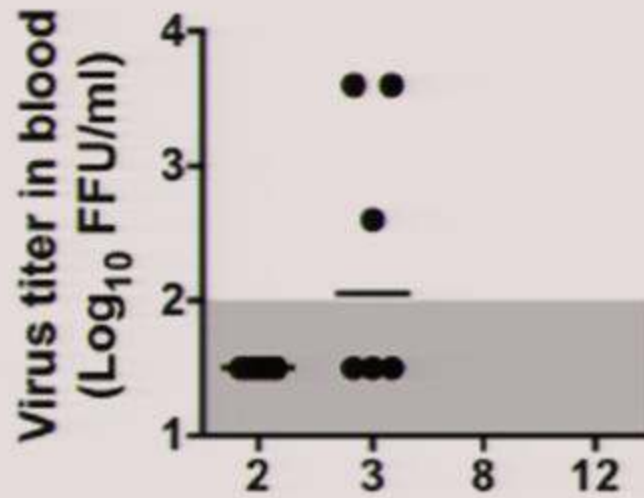
Favipiravir 300mg/kg/gün



Favipiravir alan farelerde
ALT ve AST yükselmedi

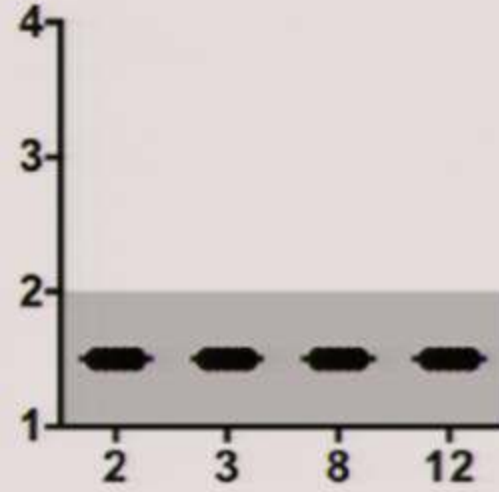
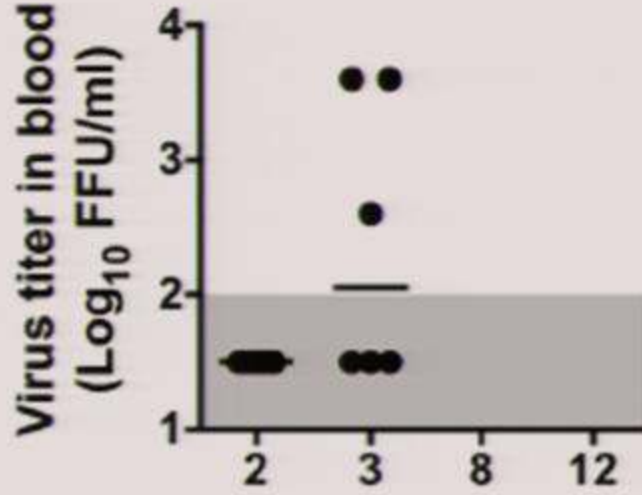
Plasebo

Favipiravir 300mg/kg/gün



Plasebo

Favipiravir 300mg/kg/gün

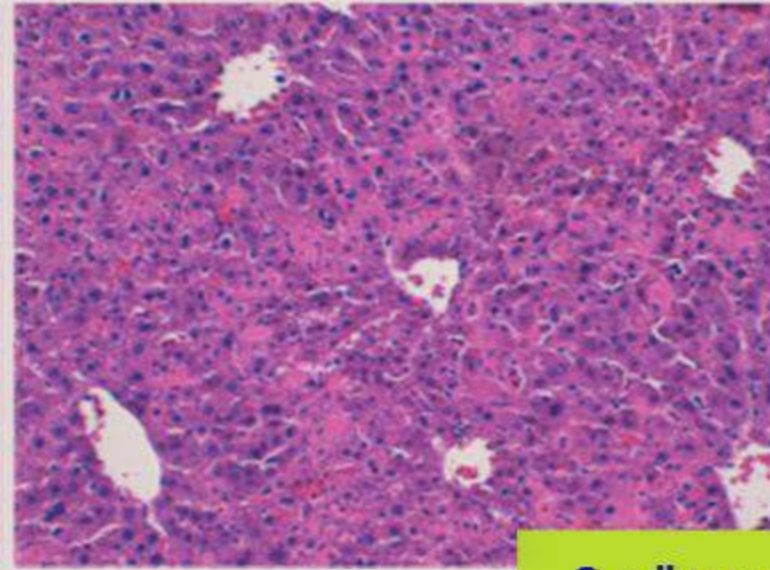
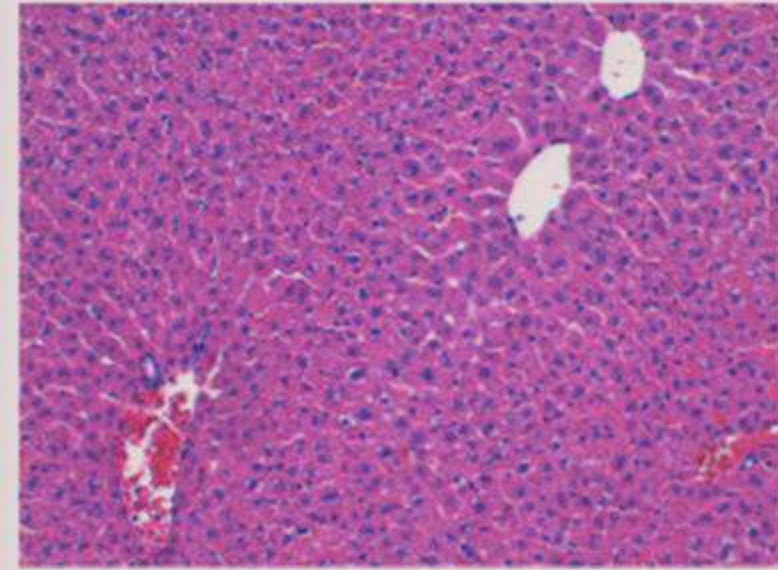


Favipiravir alan farelerin
serumunda ve organlarında
virüs saptanmadı

Naïve

Infected

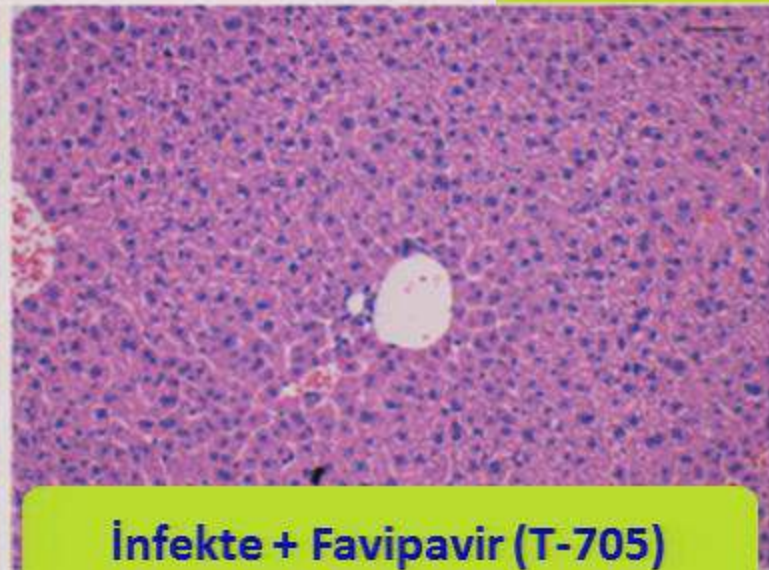
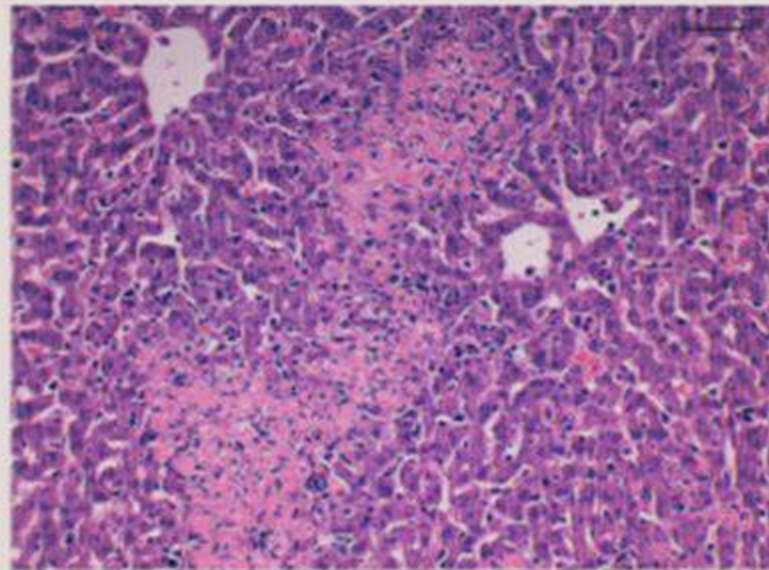
Liver (H&E)



Infected + ribavirin

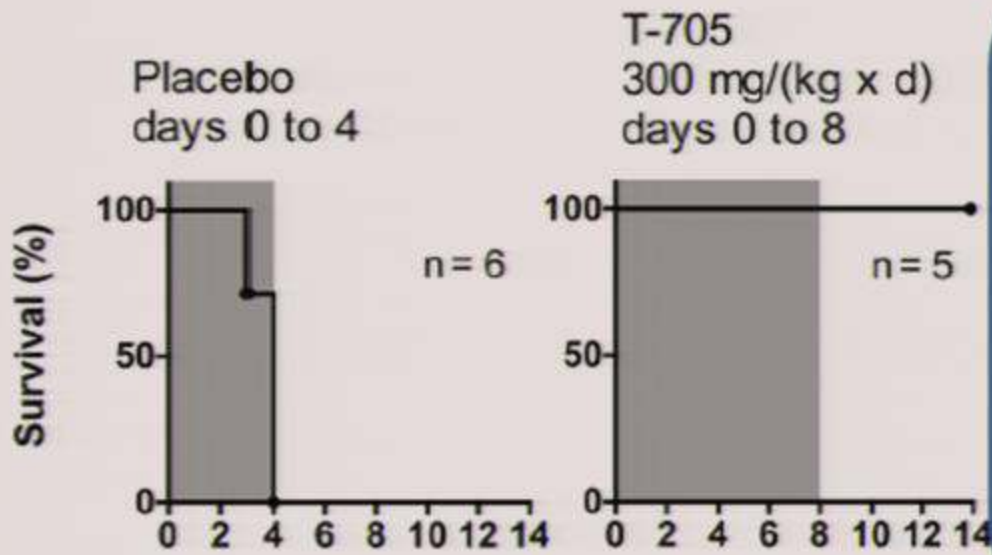
Infected + T-705

3. gün yapılan histopatolojik
incelemede hemen hemen
normal karaciğer bulguları



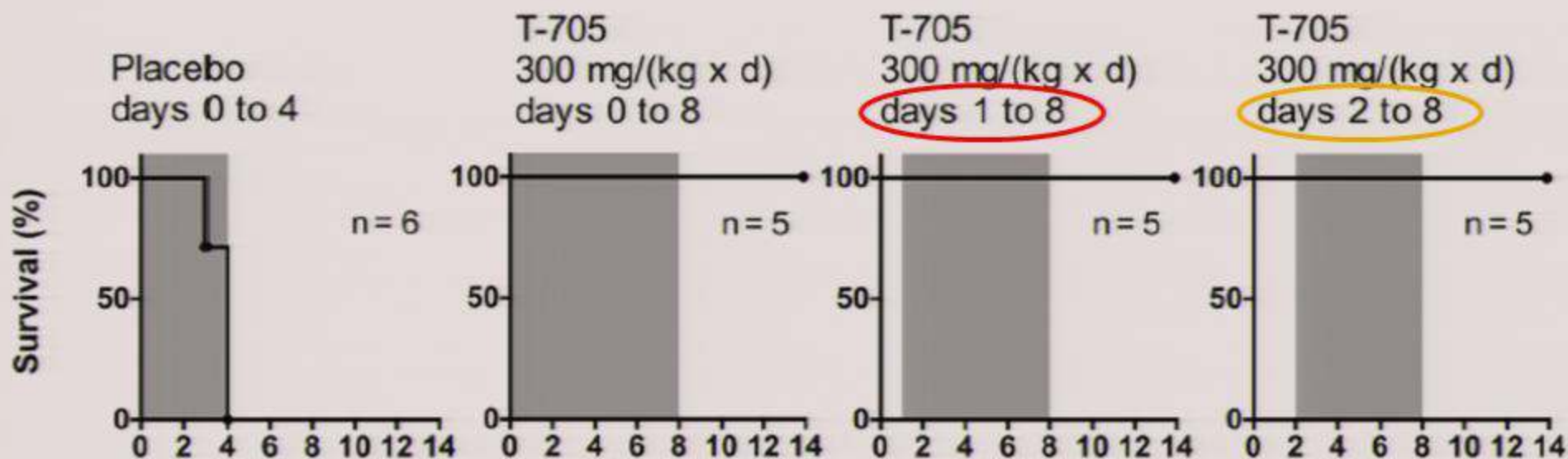
infekte + Favipavir (T-705)

Favipiravir (T-705) “Geç” uygulama



Favipiravir (T-705)

“Geç” uygulama



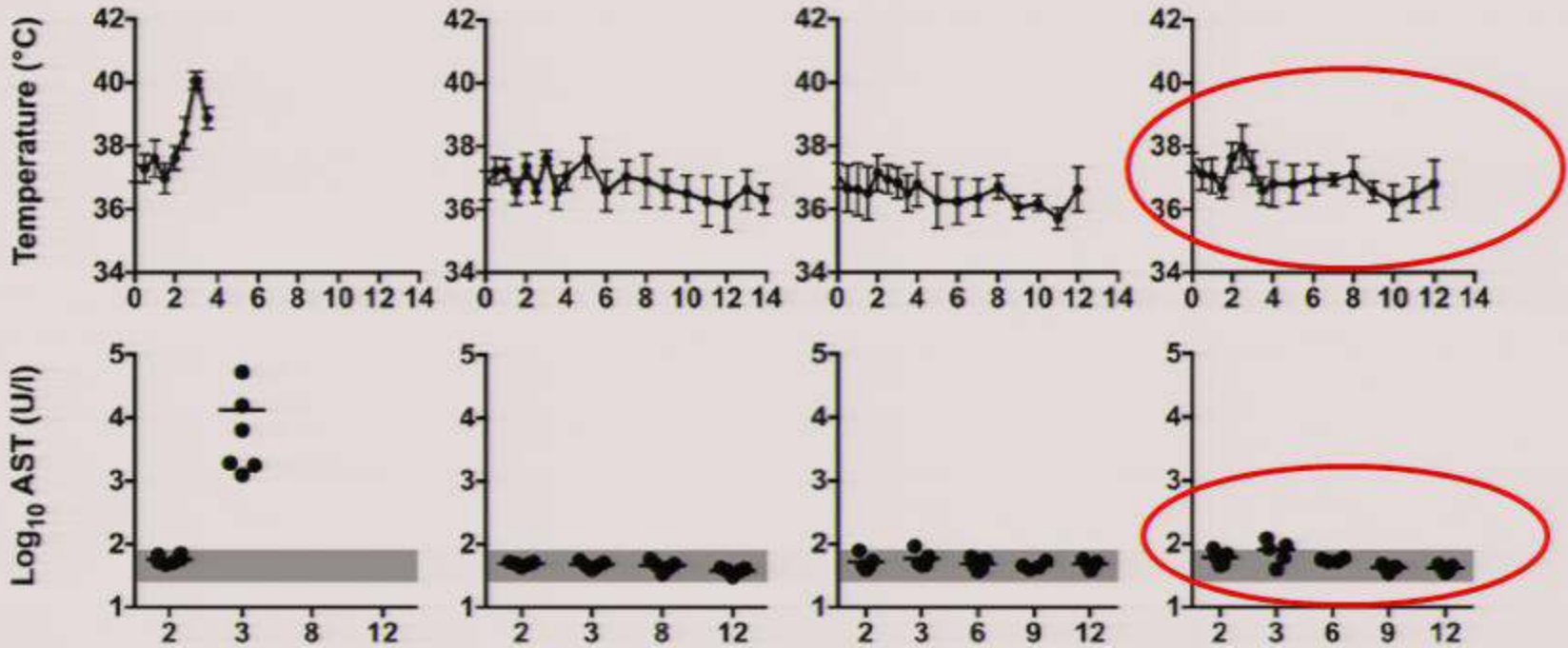
Favipiravir (T-705)

“Geç” uygulama

İnokülasyondan sonra Favipiravir başlama zamanı	21. günde serumda KKKA virüsüne karşı antikor pozitifliği
0.gün	1/10 (%10)
1. gün 2. gün	10/10 (%100)

Favipiravir (T-705) “Geç” uygulama

Plasebo	Favipiravir başlangıcı 0. gün	Favipiravir başlangıcı 1. gün	Favipiravir başlangıcı 2. gün
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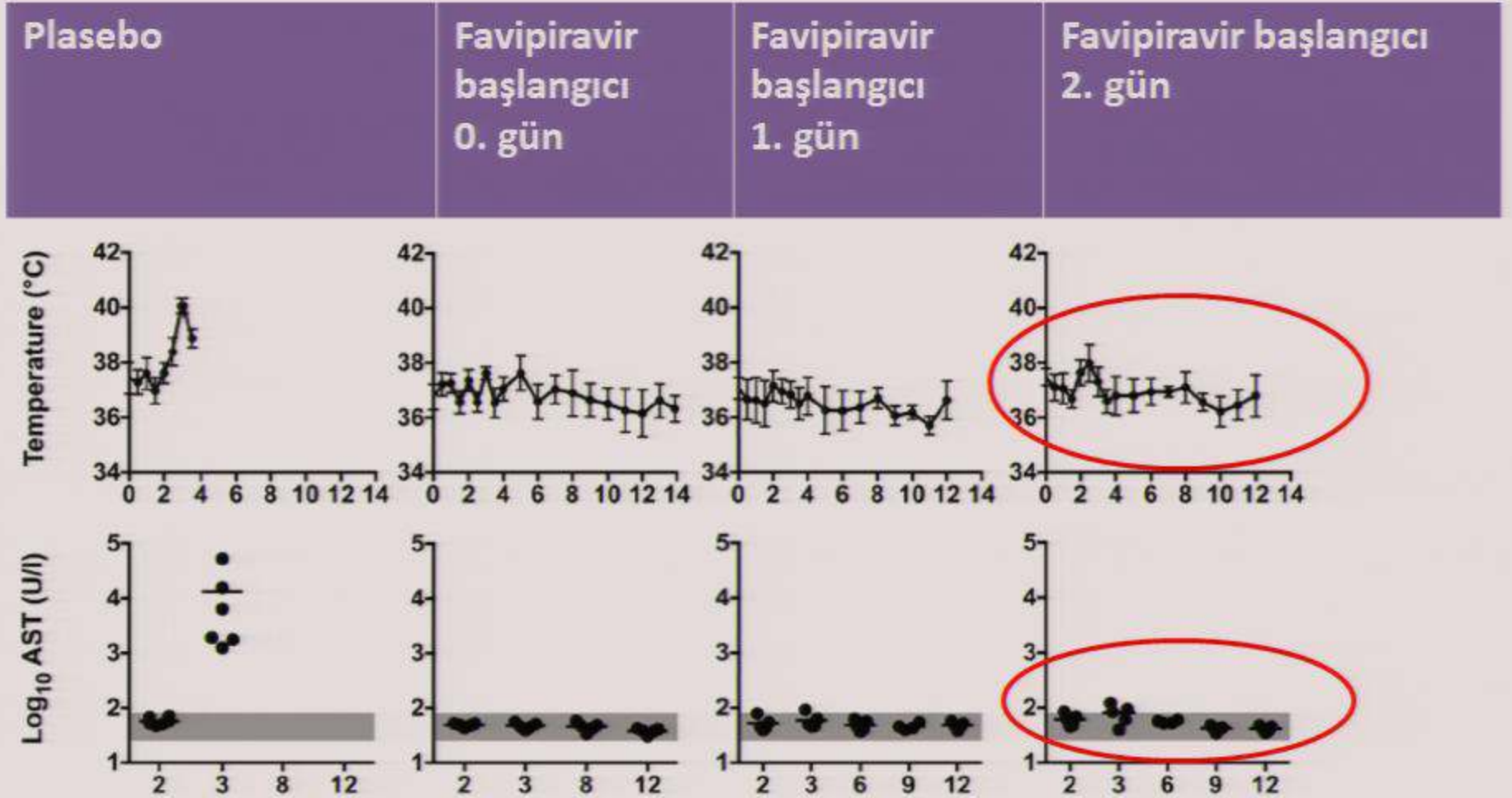


Favipiravir (T-705)

“Geç” uygulama

İnokülasyondan sonra Favipiravir başlama zamanı	21. günde serumda KKKA virüsüne karşı antikor pozitifliği
0.gün	1/10 (%10)
1. gün 2. gün	10/10 (%100)

Favipiravir (T-705) “Geç” uygulama



Favipiravir (T-705)

“Geç” uygulama

İnokülasyondan sonra Favipiravir başlama zamanı	21. günde serumda KKKA virüsüne karşı antikor pozitifliği
0.gün	1/10 (%10)
1. gün 2. gün	10/10 (%100)

Favipiravir (T-705) - Optimal Doz

Favipiravir (T-705)

Plasebo (metil selüloz)

30 mg/kg/gün

15 mg/kg/gün

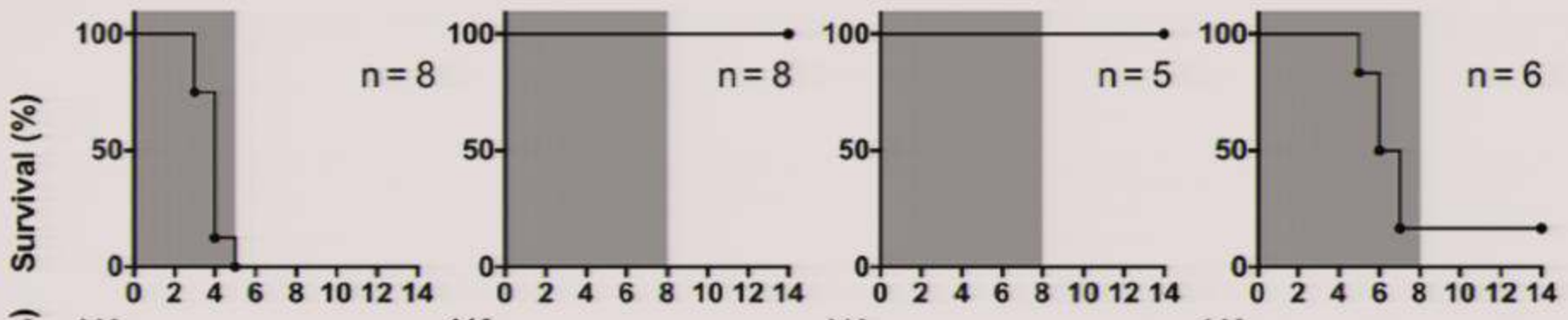
7,5 mg/kg/gün

Placebo
days 0 to 5

T-705
30 mg/(kg x d)
days 0 to 8

T-705
15 mg/(kg x d)
days 0 to 8

T-705
7.5 mg/(kg x d)
days 0 to 8



KKKA - Transgenik fare modelinde Favipiravir (T-705)

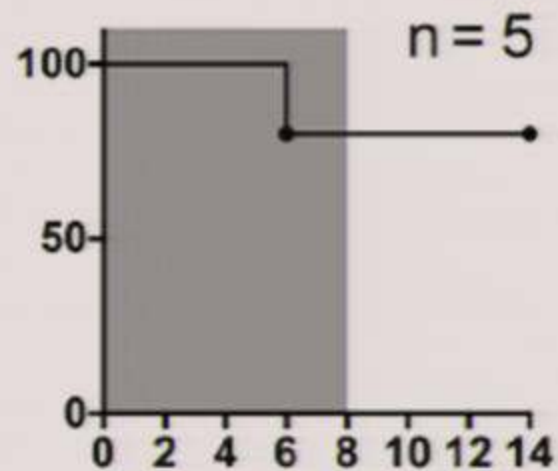
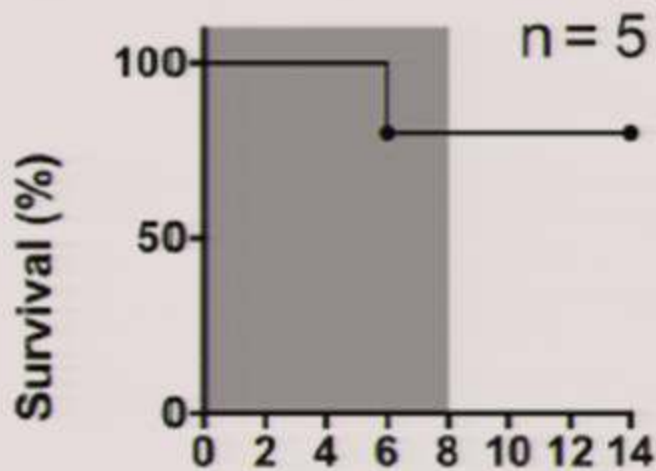
- İnokülasyon ile aynı gün başlanılan 300 mg Favipiravir
 - Enfeksiyon gelişimini önledi.
 - Hastalık belirti ve bulguları oluşmadı.
- Geç başlanılan (1. veya 2. gün) Favipiravir
 - Enfeksiyon gelişti (*serolojik yanıt*)
 - Hastalık belirti ve bulgularını büyük ölçüde görülmedi
 - Fatal seyir görülmedi.

Ribavirin + Favipiravir kombinasyonu

Ribavirin 100 mg/(kg x d)

T-705
30 mg/(kg x d)

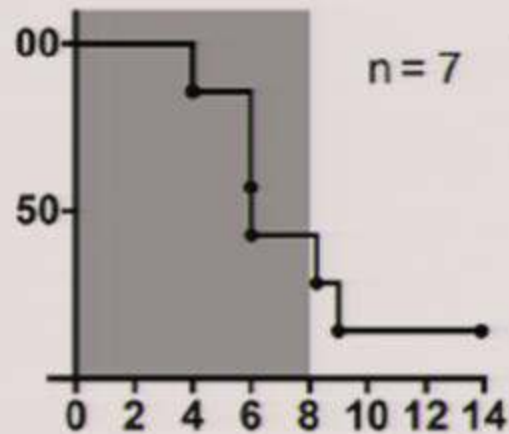
T-705
7.5 mg/(kg x d)



Ribavirin + Favipiravir kombinasyonu

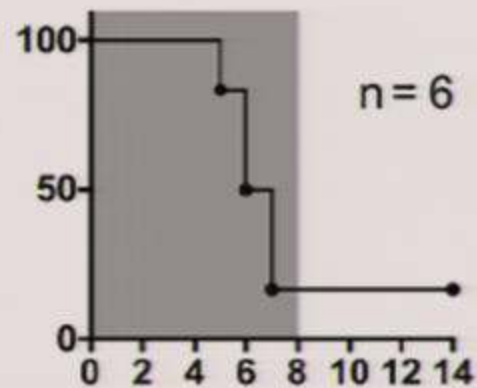
Ribavirin
Monoterapi
100 mg/kg/gün

Ribavirin
100 mg/(kg x d)
days 0 to 8



Favipiravir
Monoterapi
7,5 mg/kg/gün

T-705
7.5 mg/(kg x d)
days 0 to 8



KKKA - Transgenik fare modelinde Favipiravir (T-705)

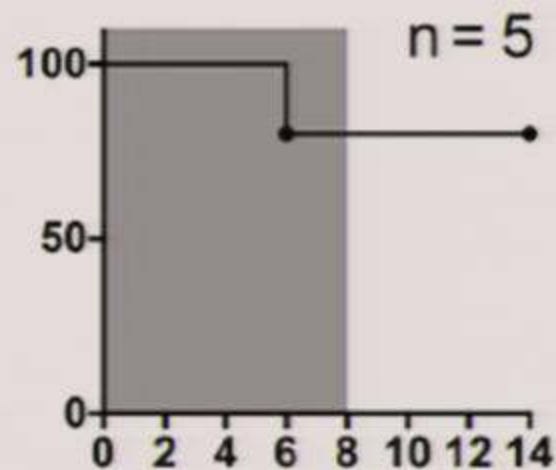
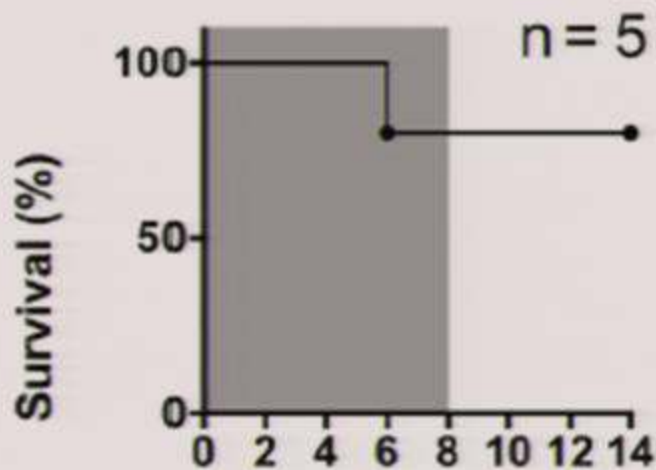
- İnokülasyon ile aynı gün başlanılan 300 mg Favipiravir
 - Enfeksiyon gelişimini önledi.
 - Hastalık belirti ve bulguları oluşmadı.
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 - Enfeksiyon gelişti (*serolojik yanıt*)
 - Hastalık belirti ve bulgularını büyük ölçüde görülmedi
 - Fatal seyir görülmedi.

Ribavirin + Favipiravir kombinasyonu

Ribavirin 100 mg/(kg x d)

T-705
30 mg/(kg x d)

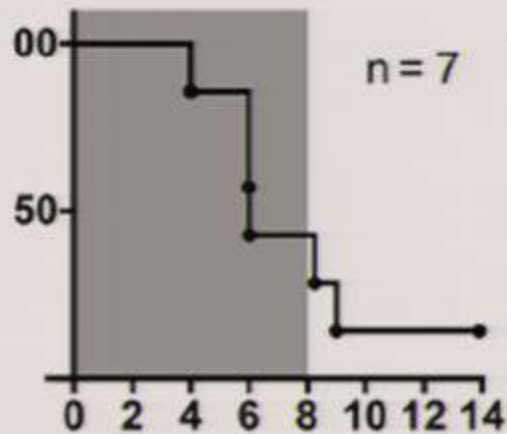
T-705
7.5 mg/(kg x d)



Ribavirin + Favipiravir kombinasyonu

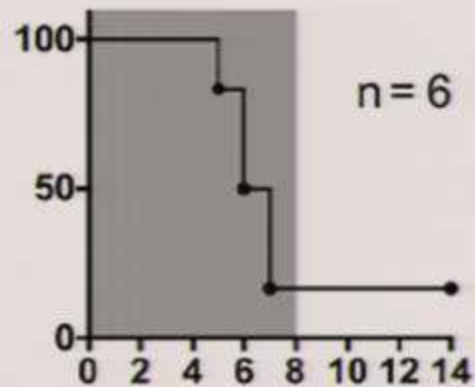
Ribavirin
Monoterapi
100 mg/kg/gün

Ribavirin
100 mg/(kg x d)
days 0 to 8



Favipiravir
Monoterapi
7,5 mg/kg/gün

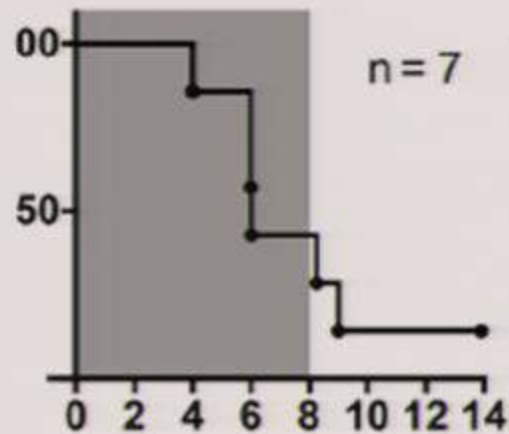
T-705
7.5 mg/(kg x d)
days 0 to 8



Ribavirin + Favipiravir kombinasyonu

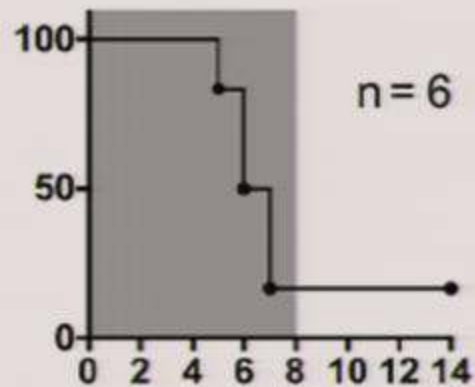
Ribavirin
Monoterapi
100 mg/kg/gün

Ribavirin
100 mg/(kg x d)
days 0 to 8



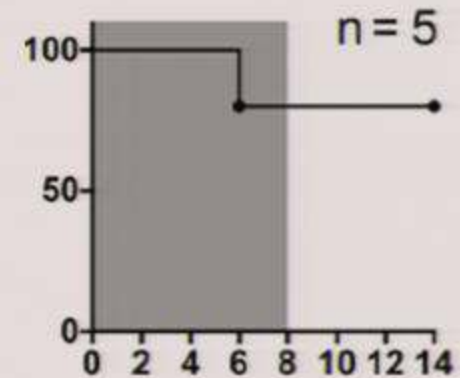
Favipiravir
Monoterapi
7,5 mg/kg/gün

T-705
7.5 mg/(kg x d)
days 0 to 8



Ribavirin + Favipiravir

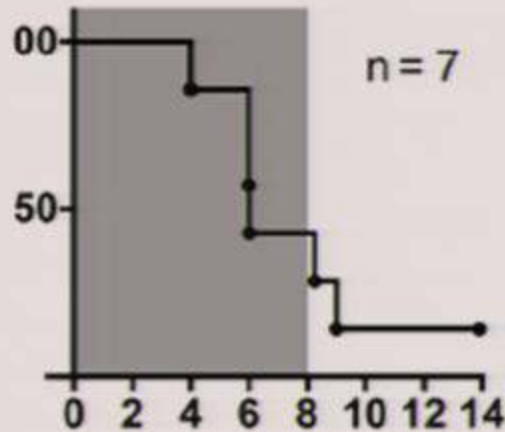
T-705
7.5 mg/(kg x d)



Ribavirin + Favipiravir kombinasyonu

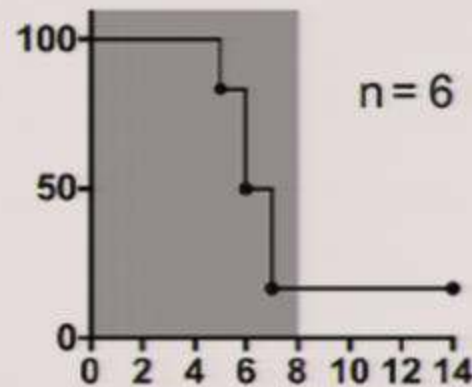
Ribavirin
Monoterapi
100 mg/kg/gün

Ribavirin
100 mg/(kg x d)
days 0 to 8



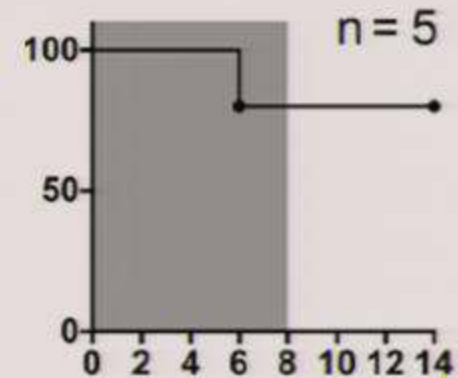
Favipiravir
Monoterapi
7,5 mg/kg/gün

T-705
7.5 mg/(kg x d)
days 0 to 8



Ribavirin + Favipiravir

T-705
7.5 mg/(kg x d)



Ribavirin + Favipiravir
SİNERJİSTİK etki göstermiştir.

A Novel Vaccine against Crimean-Congo Haemorrhagic Fever Protects 100% of Animals against Lethal Challenge in a Mouse Model

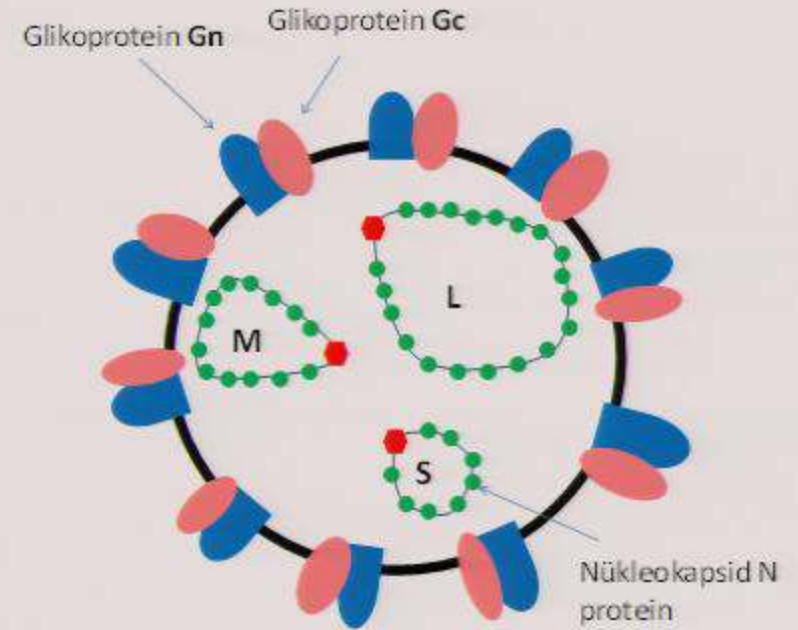
Karen R. Buttigieg, Stuart D. Dowall, Stephen Findlay-Wilson, Aleksandra Miloszezewska, Emma Rayner, Roger Hewson*, Miles W. Carroll

Microbiology Services Research, Public Health England, Porton Down, Wiltshire, [United Kingdom](#)

Abstract

Crimean-Congo Haemorrhagic Fever (CCHF) is a severe tick-borne disease, endemic in many countries in Africa, the Middle East, Eastern Europe and Asia. Between 15–70% of reported cases are fatal. There is no approved vaccine available, and preclinical protection *in vivo* by an experimental vaccine has not been demonstrated previously. In the present study, the attenuated poxvirus vector, Modified Vaccinia virus Ankara, was used to develop a recombinant candidate vaccine expressing the CCHF virus glycoproteins. Cellular and humoral immunogenicity was confirmed in two mouse strains, including type I interferon receptor knockout mice, which are susceptible to CCHF disease. This vaccine protected all recipient animals from lethal disease in a challenge model adapted to represent infection via a tick bite. Histopathology and viral load analysis of protected animals confirmed that they had been exposed to challenge virus, even though they did not exhibit clinical signs. [This is the first demonstration of efficacy of a CCHF vaccine.](#)

- **Modifiye vaksiniya Ankara (*Modified vaccinia Ankara*)** çiçek aşısı suşuna KKA virüsünün M genomu yerleştiriliyor.
- M segmenti yüzey glikoproteinini (Gn ve Gc) sentezini düzenler.
- Gn ve Gc glikoproteinleri virüsün konak hücreye tutunmasında rol alır.
- Konak hücrelerinden Gn ve Gc'ye karşı antikor yanıtı oluşur.



Test edilen aşılar

- Modifiye vaksiniya Ankara-KKKA virüsü glikoproteini içeren suş **(MVA-GP)**
- Modifiye vaksiniya Ankara **(MVA)** suşu
- **Plasebo** (SF)

Denekler



- Transgenik fareler (5-8) haftalık.
- **0. ve 14. gün** iki doz aşı yapıyor.
- Aşılanan fareler **28. gün** (*rapelden 14 gün sonra*) KKKA virüsü ile intraperitoneal yoldan enfekte ediliyor.
- **42. gün** izlem sonlandırılıyor.

Aşı suşu	İçerdiği viral yük	Aşı volümü ve veriliş yolu
MVA-GP	10^7 pfu	100 µl iM yolla
MVA	10^7 pfu	100 µl iM yolla
Plasebo (SF)	-	100 µl iM yolla

Rapel dozdan 7 gün sonra KKKA virüsüne karşı hücresel immün yanıt

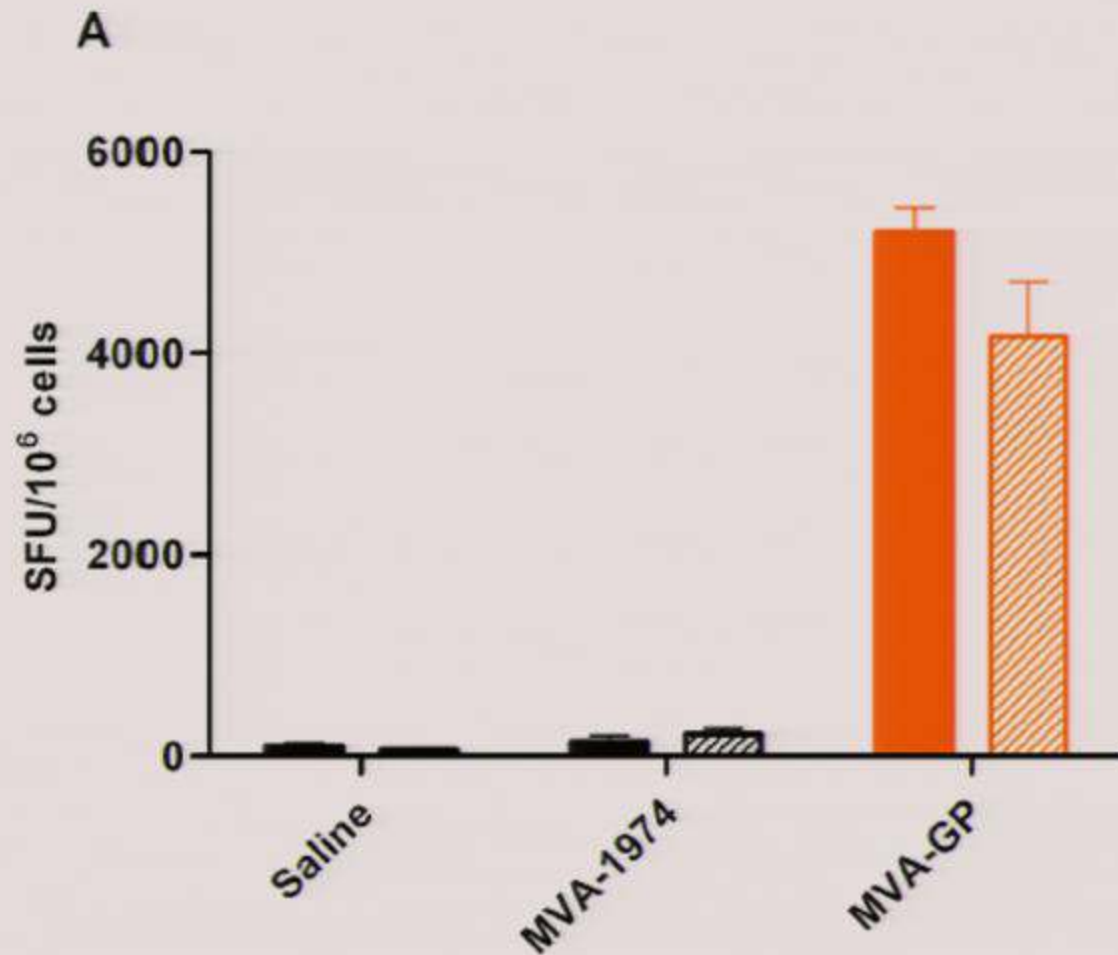


Figure 3. IFN- γ ELISpot responses from vaccinated A129 and 129Sv/Ev mice, 7 days after booster vaccination. Splenocytes

Rapel dozdan 14 gün sonra KKKA virüsüne karşı humoral immün yanıt

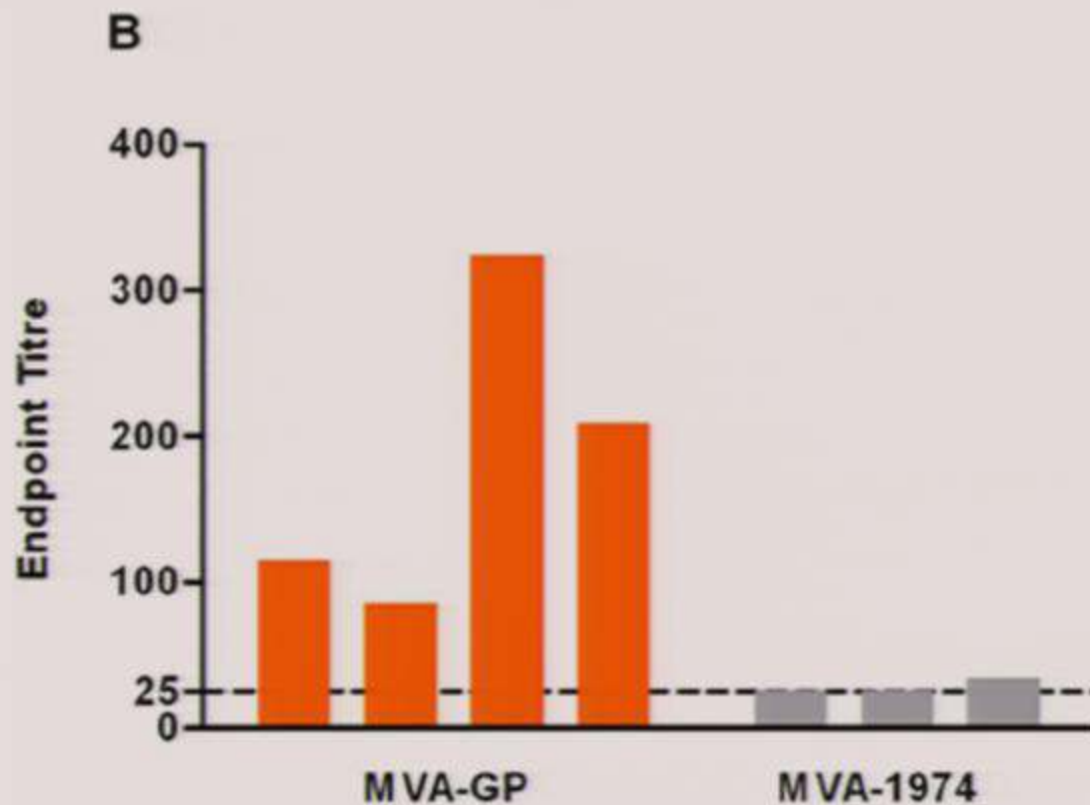
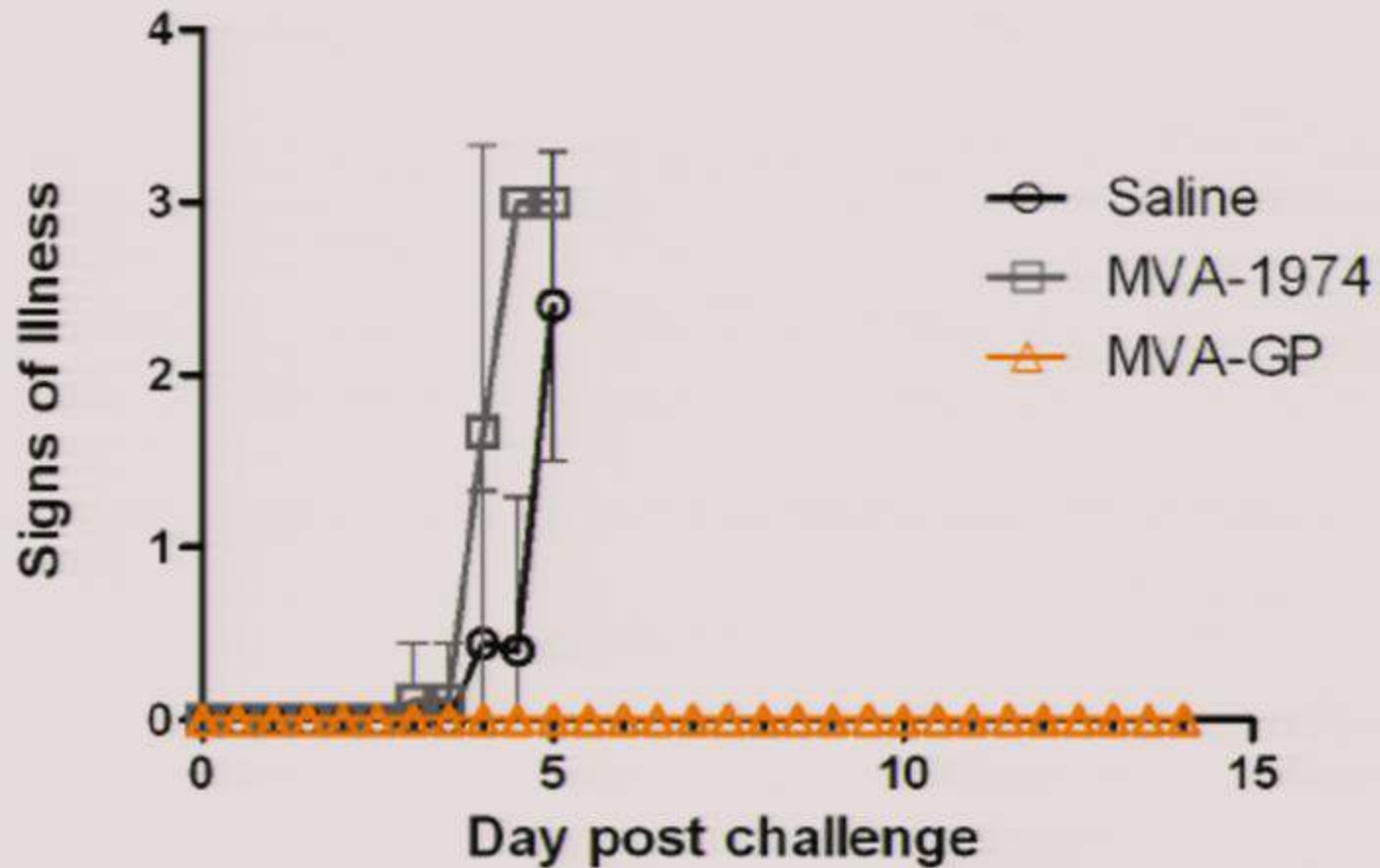
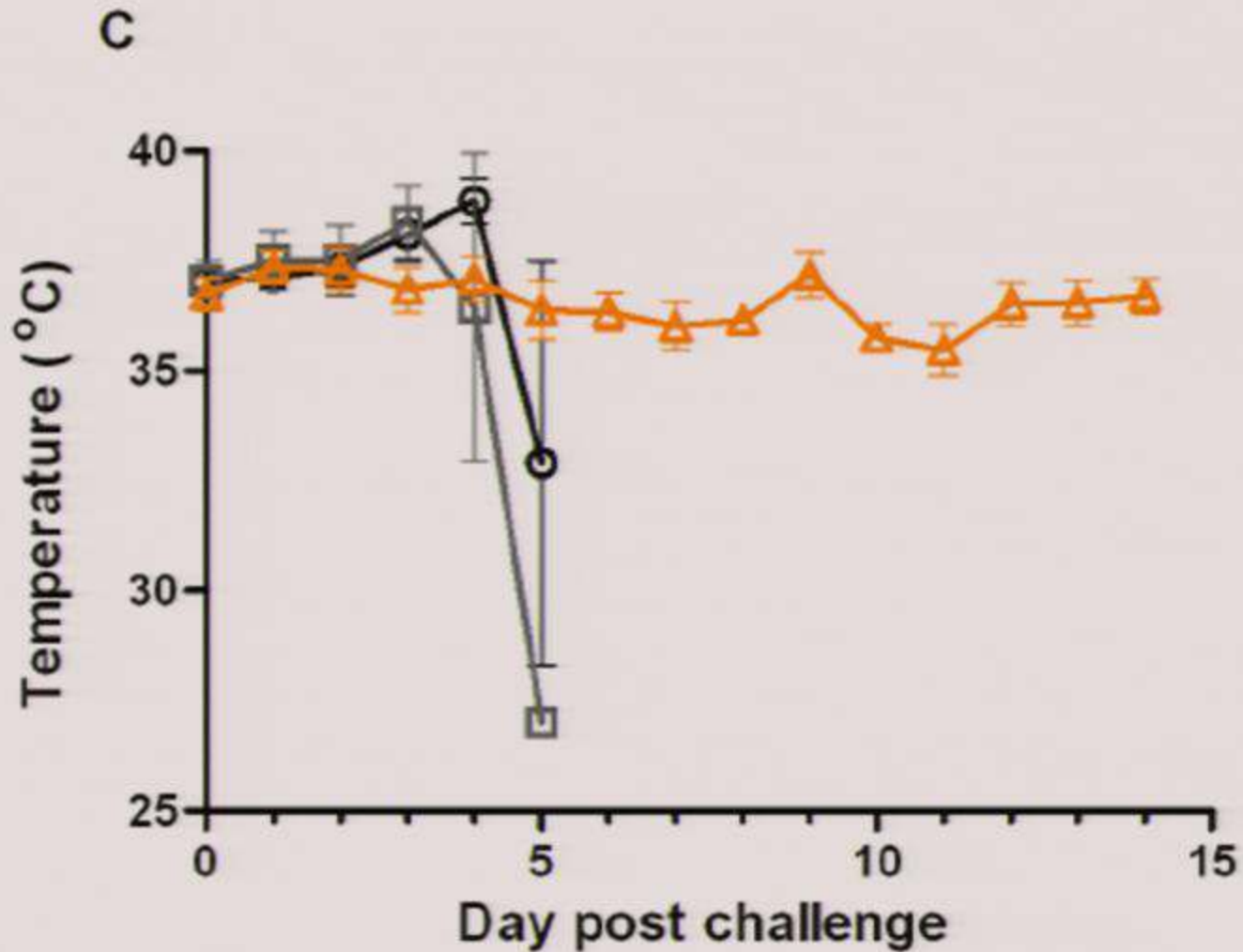


Figure 5. ELISA analysis of antibodies from A129 mice, 14 days after booster vaccination. Pooled sera from A129 mice, 14 days after booster vaccination with MVA-GP (orange) or MVA 1974 (grey), were

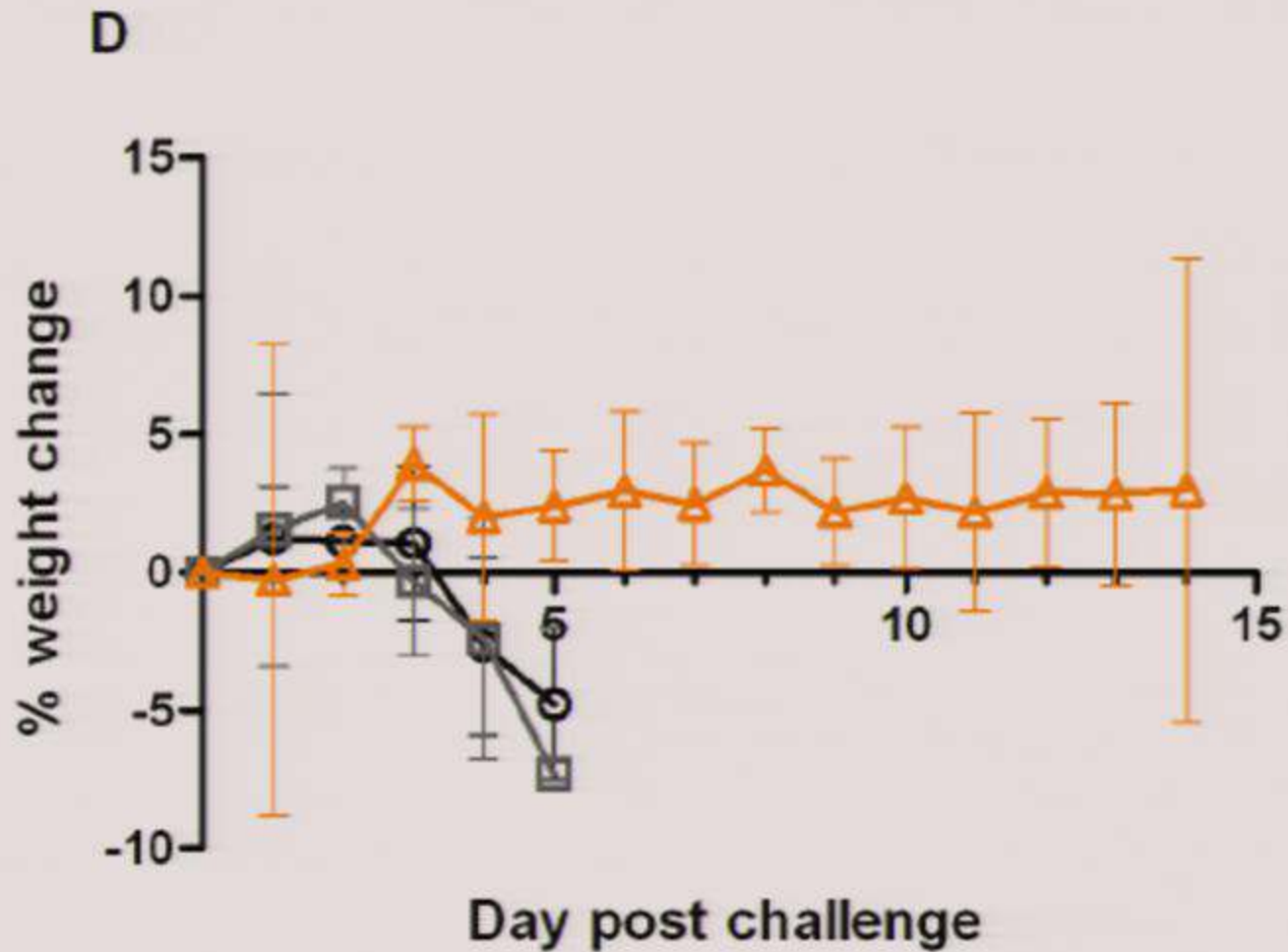
Virüs inokülasyonundan sonra hastalık belirtileri



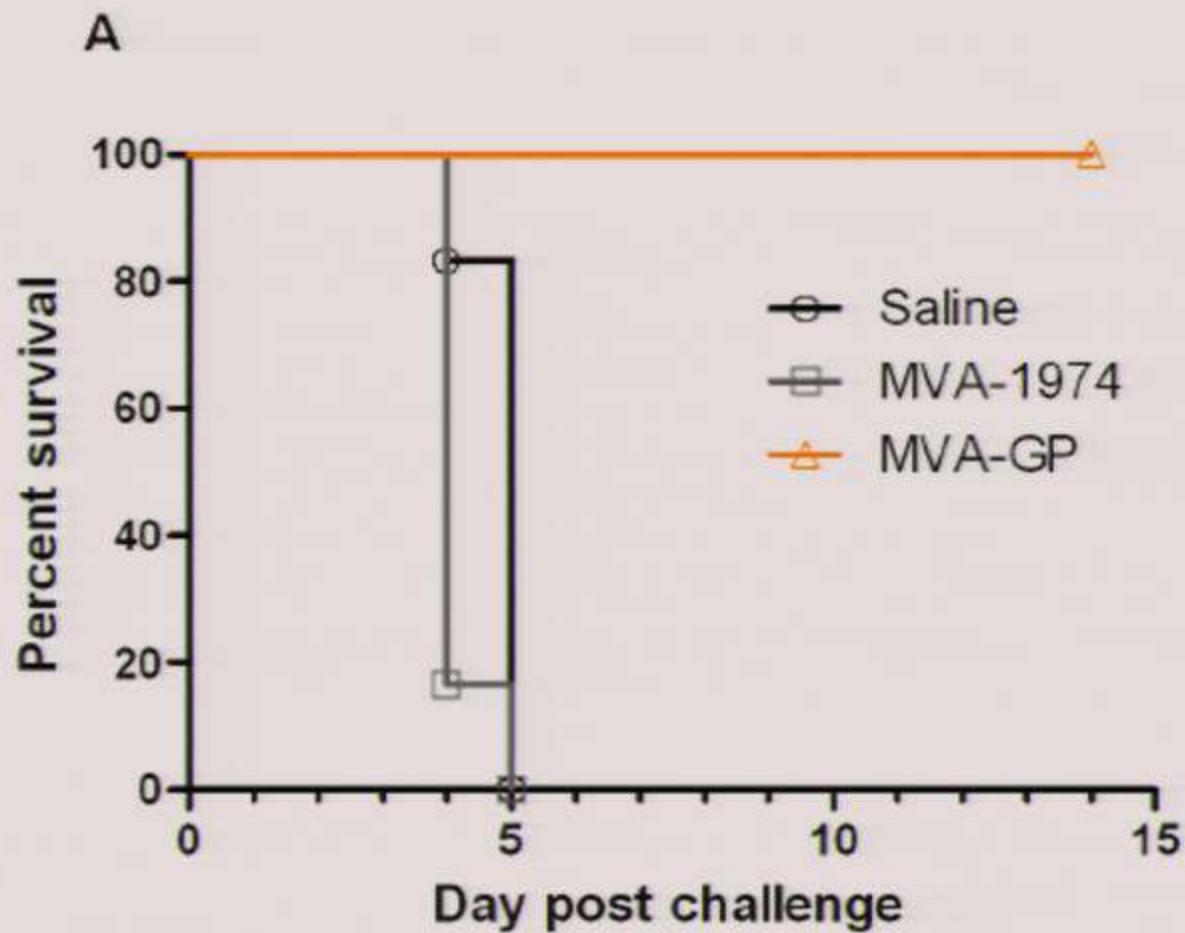
Virüs inokülasyonundan sonra ateş



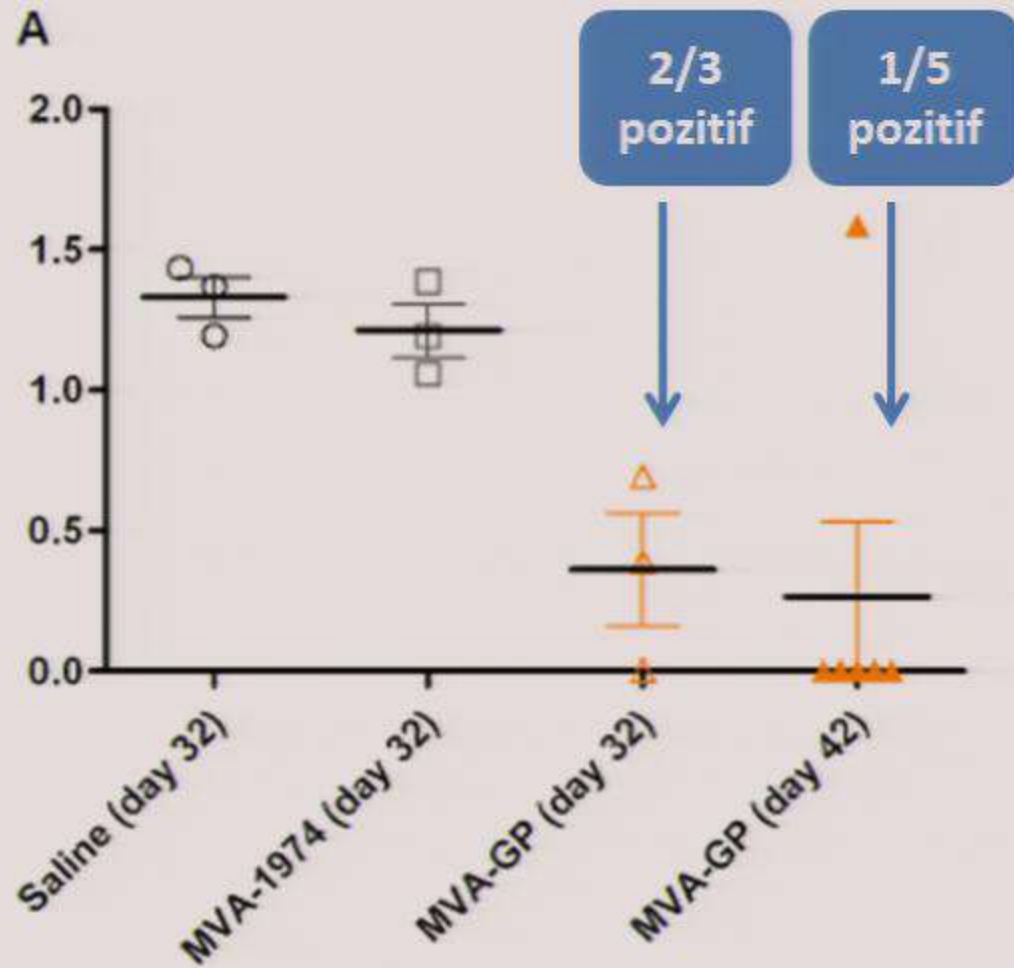
Virüs inokülasyonundan sonra vücut ağırlığı



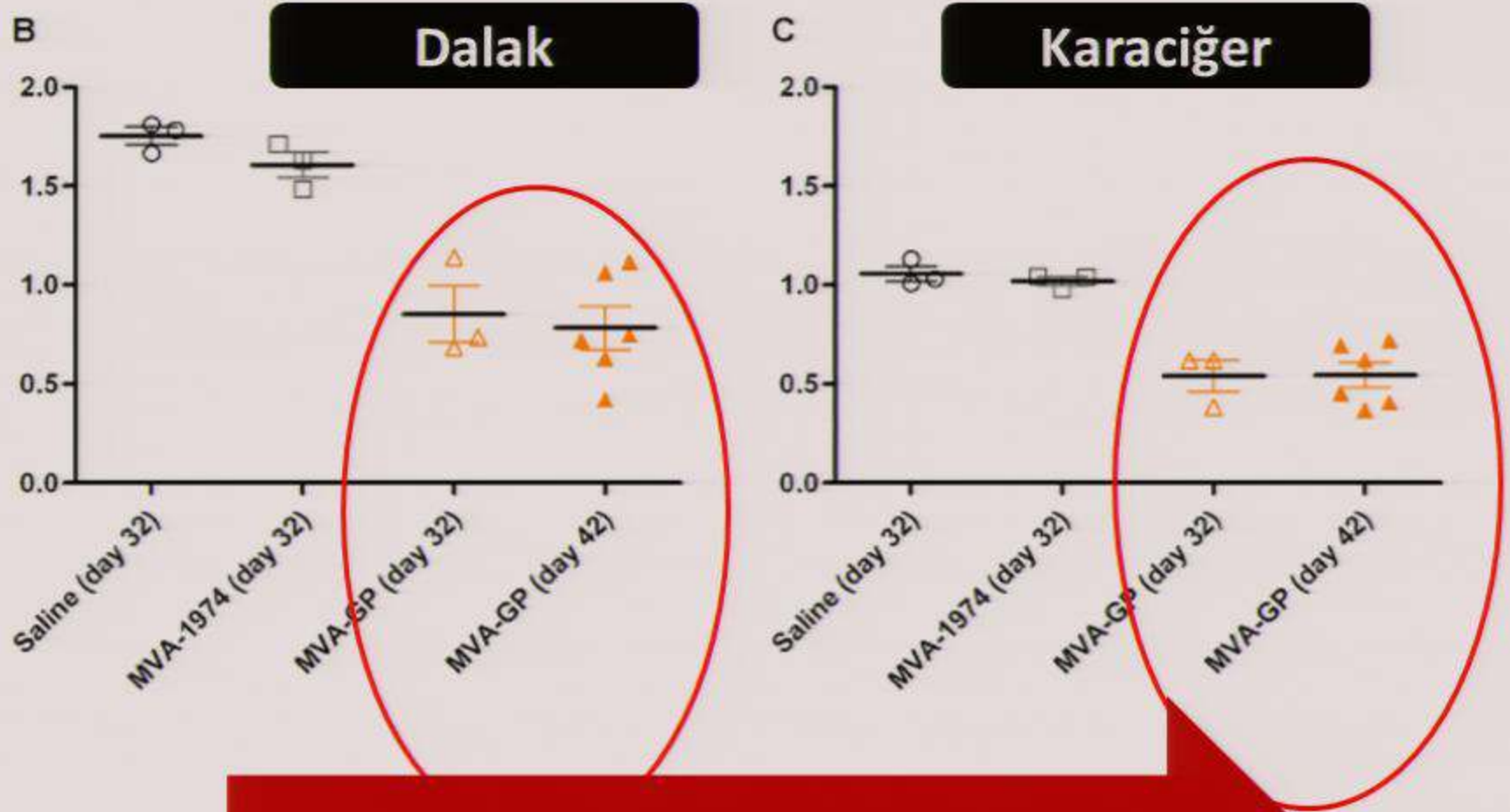
Virüs inokülasyonundan sonra sağ kalım



İnokülasyon sonrası kanda viral RNA



İnokülasyon sonrası dokularda viral RNA



Tüm dokularda viral RNA pozitif

Kan ve dokulardaki viral RNA infeksiyeli mi?

- Dokulardan hazırlanan süspansiyonlar hücre kültürüne ekliyor.
- 48 saat sonunda sitopatik etki yok
- Daha uzun inkübasyon????

KKKA virüs Aşısı

- MVA-GP aşısı denek farelerde hastalık gelişimini önüyor.
- Fatalite görülüyor.
- Ancak aşının bu başarısına rağmen kan ve dokularda viral RNA saptanıyor.

Hantavirüs



Hantaan virüs (HNTV)	İnsanlarda RSKA	<i>Kemirici (Apodemus agrarius – Asya tipi)</i>	Asya
Amur virüs (AMRV)	İnsanlarda RSKA	<i>Kemirici (Apodemus peninsulae)</i>	Asya
Seoul virus (SEOV)	İnsanlarda RSKA	<i>Kemirici (Rattus norvegicus)</i>	Tüm dünya
Puumala virüs (PUUV)	İnsanlarda RSKA	<i>Kemirici (Myodes glareolus)</i>	Batı Avrupa, <u>Türkiye</u>
Dobrova virüs (DOBV)	İnsanlarda RSKA	<i>Kemirici (Apodemus flavicollis)</i>	Doğu Avrupa, <u>Türkiye</u>
Saaremaa virüs (SAAV)	İnsanlarda RSKA	<i>Kemirici (Apodemus agrarius - Avrupa tipi)</i>	Avrupa
Tula virüs (TULV)	İnsanlarda RSKA	<i>Kemirici (Microtus arvalis)</i>	Avrupa
Sin Nombre virüs	İnsanlarda HPS	<i>Kemirici (Peromyscus maniculatus)</i>	Kuzey Amerika
Andes virüs (ANDV)	İnsanlarda HPS	<i>Kemirici (Oligoryzomys longicaudatus)</i>	Güney Amerika

Hantavirüs - Tedavi



- **Destek tedavisi**

- Hipotansiyon ve şoktan korumak için sıvı elektrolit replasmanı
- Kanama kontrolü için destek tedavisi (*taze donmuş plazma, trombosit, kan transfüzyonu*)
- Böbrek yetmezliğinde hemodiyaliz

- **Spesifik Tedavi**

- Antiviral tedavi
- İmmünglobülin tedavisi

Hantavirüs - RİBAVİRİN

- Çin'de 1985-1987 yılları arasında
 - Randomize, çift kör ve plasebo kontrollü bir çalışmada serolojik olarak doğrulanmış 242 RSKA olgusuna
 - **Plasebo veya intravenöz Ribavirin** (*yükleme dozu 33 mg/kg, ilk 4 gün 16 mg/kg her 6 saate bir, sonraki 3 gün 8 mg/kg her 8 saatte bir*).
- Hastalığın ilk 7 günü içerisinde başlanan İV ribavirin tedavisinin mortaliteyi 7 kat azalttığı saptanmıştır



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Experience with intravenous ribavirin in the treatment of hemorrhagic fever with renal syndrome in Korea[☆]

Janice M. Rusnak^{a,*}, William R. Byrne^a, Kyung N. Chung^b, Paul H. Gibbs^c, Theodore T. Kim^b, Ellen F. Boudreau^a, Thomas Cosgriff^a, Philip Pittman^a, Katie Y. Kim^d, Marianne S. Erlichman^e, David F. Rezvani^f, John W. Huggins^g

- **Kore’de yapılan kohort çalışmada 1987-2005 yılları arasında iv Ribavirin alan 33 RSKA olgusu ribavirin tedavisi almayan tarihsel kontroller ile kıyaslanmıştır.**



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- **Kore’de yapılan kohort çalışmada 1987-2005 yılları arasında iv Ribavirin alan 33 RSKA olgusu ribavirin tedavisi almayan tarihsel kontroller ile kıyaslanmıştır.**

	Tarihsel kontrol	Ribavirin alanlar
Oligüri sıklığı	% 39-69	% 3
Hemodiyaliz sıklığı	% 40	% 0

Hantavirüs – Ribavirin

- **Avrupa** ülkelerinde RSKA olgularında ribavirin kullanımını konusunda yayın yok.
 - Asya'da etkenler HNTV ve SEOV
 - Avrupa'da etkenler PUVV ve DOBV

Hantavirüs - Ribavirin

- **ABD**

- 1999-2001
- HPS şüpheli olgulara
- Randomize çift kör **plasebo X iv ribavirin**
- 24. olguda çalışma sonlandırılıyor
- Ribavirin etkili değil !!



In Vitro and *In Vivo* Activity of Ribavirin against Andes Virus Infection

David Safronetz¹, Elaine Haddock¹, Friederike Feldmann², Hideki Ebihara¹, Heinz Feldmann^{1,3*}

¹ Laboratory of Virology, Division of Intramural Research, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Hamilton, Montana, United States of America, ² Office of Operations and Management, Division of Intramural Research, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Hamilton, Montana, United States of America, ³ Department of Medical Microbiology, University of Manitoba, Winnipeg, Manitoba, Canada

- **In vitro model**

- ANDV ile enfekte edilen Vero 6 hücre kültürlerine Ribavirin uygulanıyor
- Amaç üremenin inhibisyonunun değerlendirilmesi

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- **İnvivo (Hamster) modeli**
 - ANDV ile enfekte edilen kobaylara Ribavirin profilaksisi (oral veya intarperitoneal) uygulanıyor
- Amaç;
 - Ribavirine ne zaman başlanmalı?
 - Hangi dozda verilmeli
 - Ne kadar sürdürülmeli?

In Vitro and *In Vivo* Activity of Ribavirin against Andes Virus Infection

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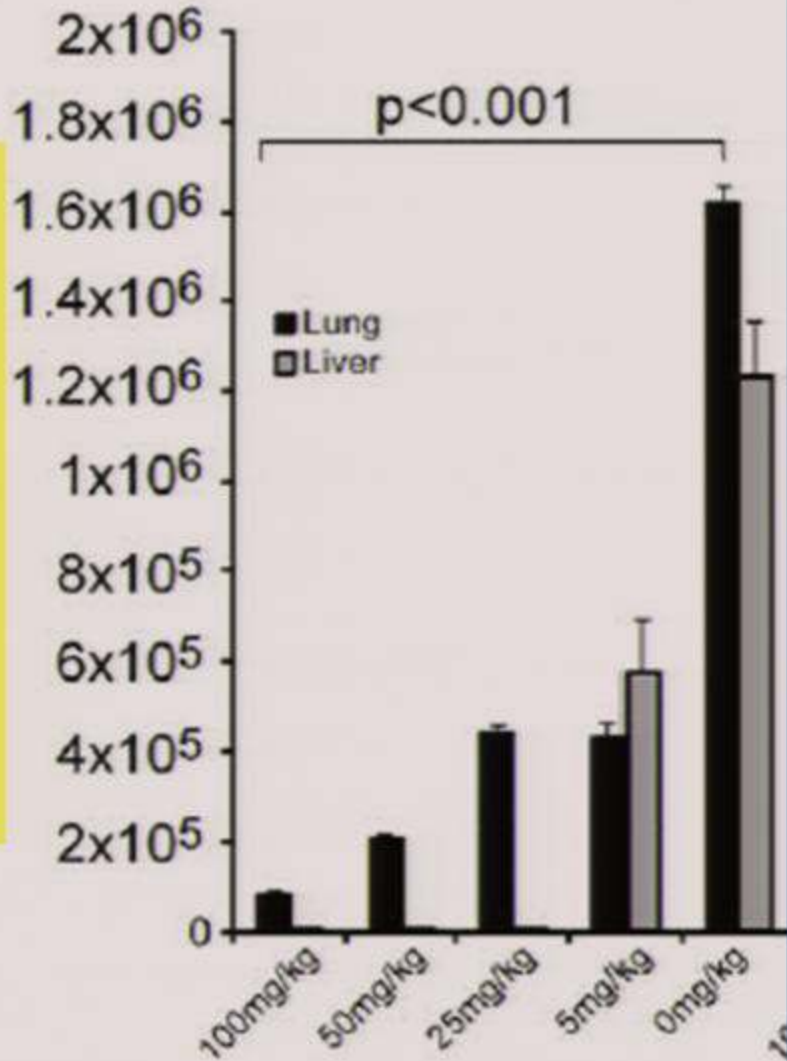
- **İnvivo (Hamster) modeli**

- ANDV ile enfekte edilen kobaylara Ribavirin profilaksisi (oral veya intarperitoneal) uygulanıyor

Temas sonrası durum	Sağ kalım oranı
Ribavirin ilk gün başlandı 10 gün verildi (oral veya intraperitoneal)	% 100 sağ kalım, semptom yok
Ribavirin 3. gün başlandı	% 100 sağ kalım
Ribavirin 5. gün başlandı	% 17 Sağ kalım
Ribavirin 7. gün başlandı	% 0 Sağ kalım

B

Viral
Yük

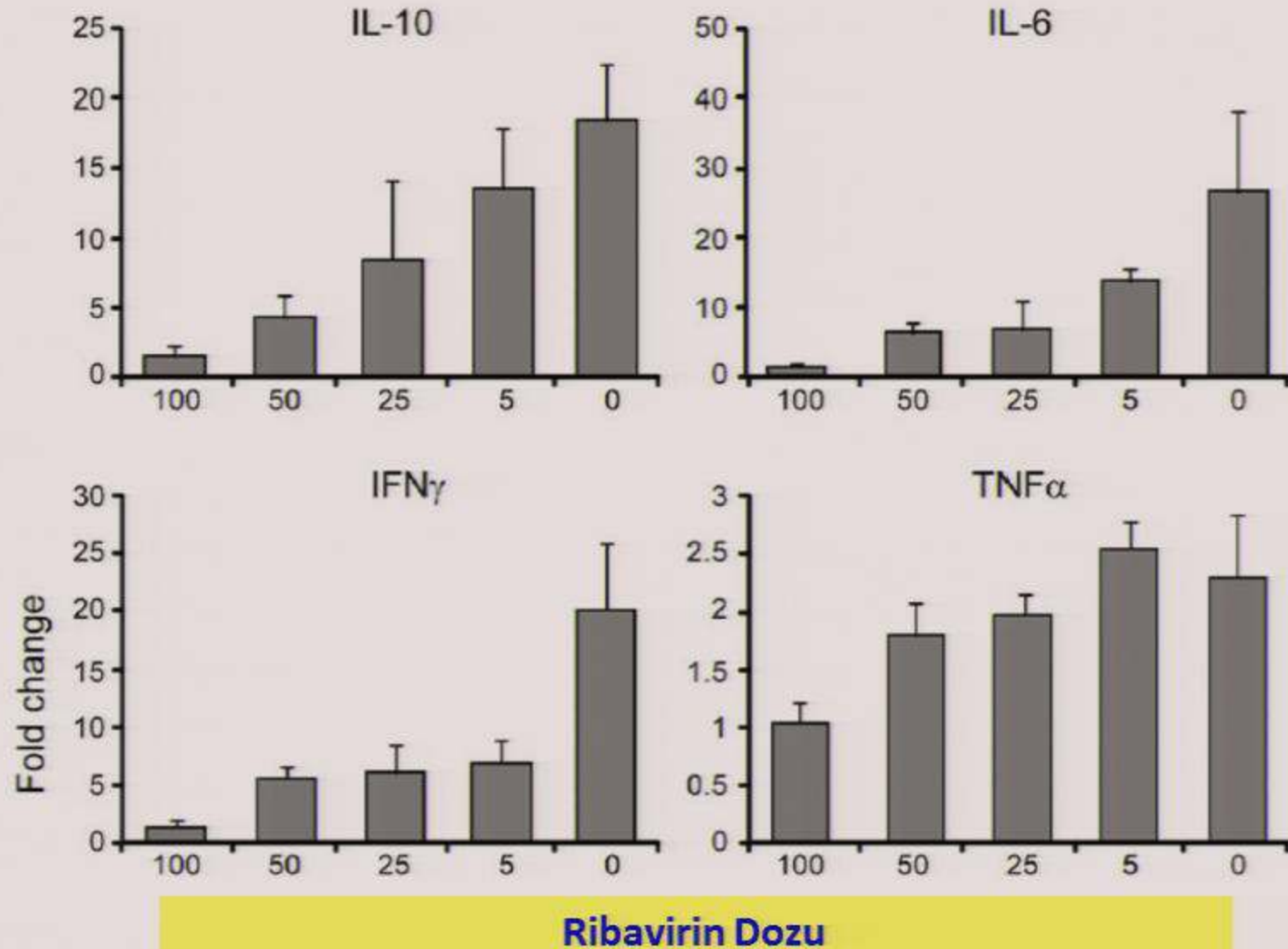


Ribavirin Dozu

Ribavirin etkinliği
doz bağımlı

Ribavirin dozu	Sağ kalım ve Semptom
5 mg/kg	Fatalite yok Hafif enfeksiyon bulguları var
25 mg/kg 50 mg/kg 100 mg/kg	Semptom yok

**Sitokin düzeyleri
temas sonrası 8.gün**



Ribavirin dozu	Sağ kalım ve Semptom
5 mg/kg	Fatalite yok Hafif enfeksiyon bulguları var
25 mg/kg 50 mg/kg 100 mg/kg	Semptom yok

Hantavirüs – Ribavirin

- **Ribavirinin etki mekanizması ??**
 - İnozin monofosfat dehidrogenaz (IMPDH) enzim inhibisyonu.
 - Viral RNA polimeraz inhibisyonu
 - Ölümcül RNA mutasyonuna yol açmak

Vero E6 hücre kültüründe ANDV'e karşı Ribavirin etkinliği

Table 1. Qualitative assessment of antiviral activity in a novel comet assay *.

Concentration	Day 4	Day 5	Day 6	Day 7	Day 8	Day 9	Day 10
ANDV alone	-	+++	+++	+++	+++	+++	+++
10 µg/mL	-	-	-	++	++	++	++
20 µg/mL	-	-	-	+	+	++	++
30 µg/mL	-	-	-	-	-	+	+
40 µg/mL	-	-	-	-	-	-	-
50 µg/mL	-	-	-	-	-	-	-
60 µg/mL	-	-	-	-	-	-	-
70 µg/mL	-	-	-	-	-	-	-

* -, no comets formed; +, few comets formed; ++, comets formed not equal to ANDV; +++, comets equal to ANDV alone.

ANDV ile enfekte edilen Suriye (Syrian) hamsterlerinde Ribavirin etkinliđi

- Hamsterler intranazal yolla ANDV ile enfekte ediliyor.
- Temas sonrası ilk günden itibaren 21 gün süreyle **Ribavirin** veriliyor (*günde iki kez intraperitoneal*)
- Temas sonrası 35 gün izlem



ANDV ile Enfekte Edildi

Günlük Ribavirin dozu

Uygulama süresi

50 mg/kg

21 gün süreyle

100 mg/kg

21 gün süreyle

200 mg/kg

21 gün süreyle

5. Günde toksisite belirtileri

Plasebo verildi

21 gün süreyle

Enfekte EDİLMEYEN Kontrol Grubu

Günlük Ribavirin dozu

Uygulama süresi

50 mg/kg

21 gün süreyle

100 mg/kg

21 gün süreyle

200 mg/kg

21 gün süreyle

5. Günde toksisite belirtileri

Plasebo verildi

21 gün süreyle

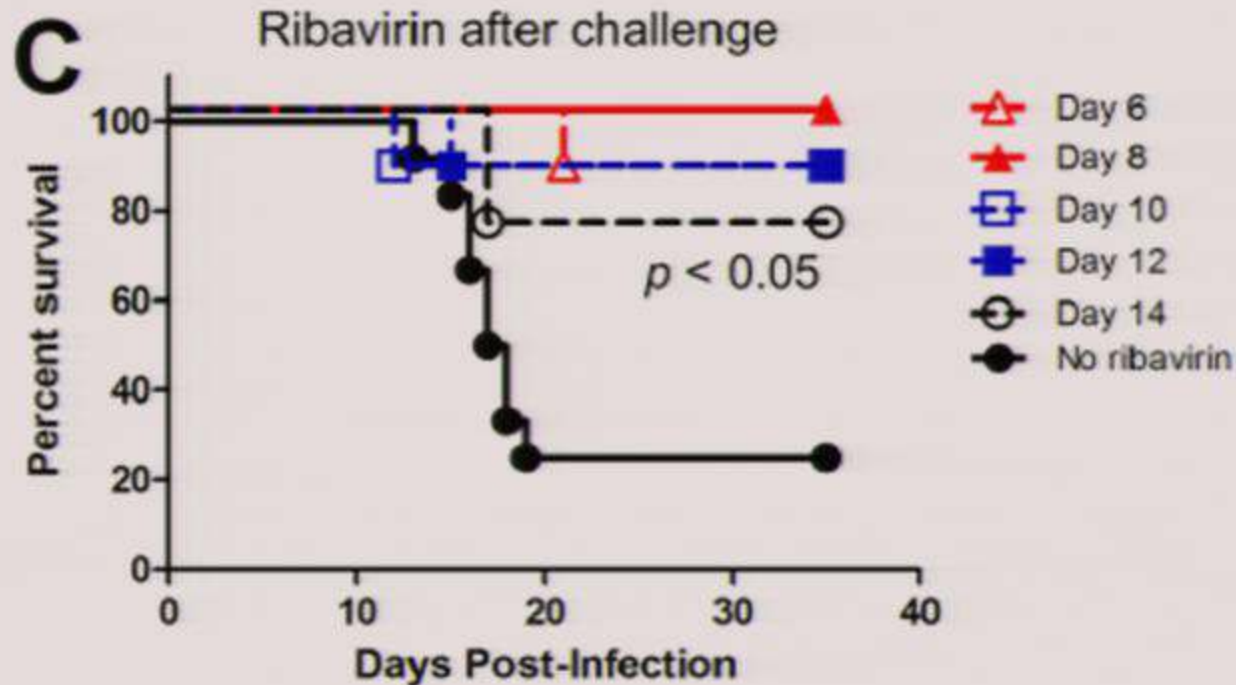
ANDV ile Enfekte Edildi

Günlük Ribavirin dozu	Uygulama süresi		
50 mg/kg	21 gün süreyle	Semptom yok	P=0,02
100 mg/kg	21 gün süreyle	Semptom yok	P=0,03
Plasebo verildi	21 gün süreyle	Letal HPS gelişti (12-24 gün içerisinde)	

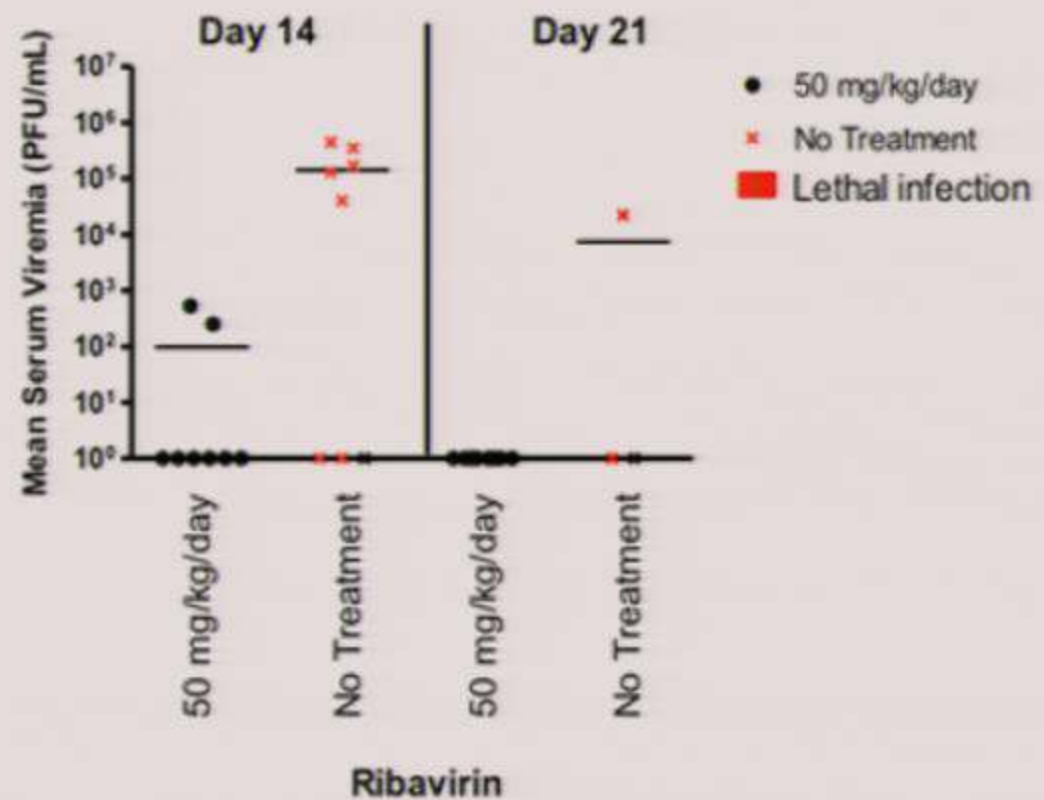
Enfekte EDİLMEYEN Kontrol Grubu (RİBAVİRİN YAN ETKİ ?)

Günlük Ribavirin dozu	Uygulama süresi		
50 mg/kg	21 gün süreyle	Semptom yok	
100 mg/kg	21 gün süreyle	Semptom yok	
Plasebo verildi	21 gün süreyle	Semptom yok	

- Diğer bir grupta **Ribavirin 50 mg/kg/gün** dozunda temas sonrası **6, 8, 10, 12** ve **14.** günlerde başlanıyor.
- Ribavirin alan grupta fatalite daha az

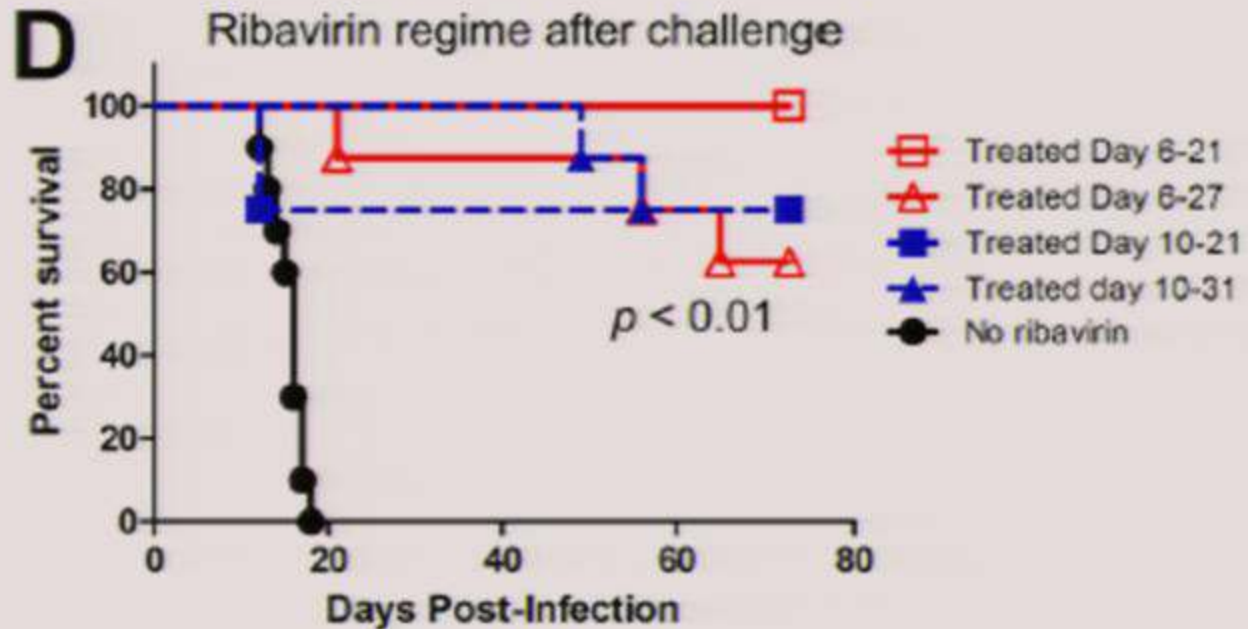


- Temas sonrası 14. ve 21. günlerde hamsterlerden serum alınıyor ve viral yük inceleniyor.
 - 100 mg/kg/gün Ribavirin alanlarda virüs negatif
 - 50 mg/kg/gün Ribavirin alanlarda



“Erken” veya “Geç” Ribavirin tedavisi

- Temas sonrası **6. veya 10. günde** Ribavirin başlanıyor.
- Kısa (14 gün) veya uzun (21 gün) veriliyor.
- Ribavirin alan grupta sağ kalım daha fazla.



Çalışmanın sonucu

- Temas sonrası 35. günde tüm hamsterlerin serumunda ANDV'e karşı antikor pozitifliği. *Tüm hamsterlerde ANDV enfeksiyonu gelişti*
- ANDV ile temastan sonraki **ilk 10 gün** içerisinde başlanan Ribavirin (50mg/kg/gün) HPS gelişimini engelledi
- Hamsterlerde ilk semptomlar yaklaşık **15-16 günde** görüldü
- Ölüm ortalama **18. günde** gelişti
- *Ribavirin temas sonrasında en geç 10 gün içerisinde başlanmalı ve en az 11 gün sürdürülmelidir.*

Antiviral Efficacy of Favipiravir against Two Prominent Etiological Agents of Hantavirus Pulmonary Syndrome

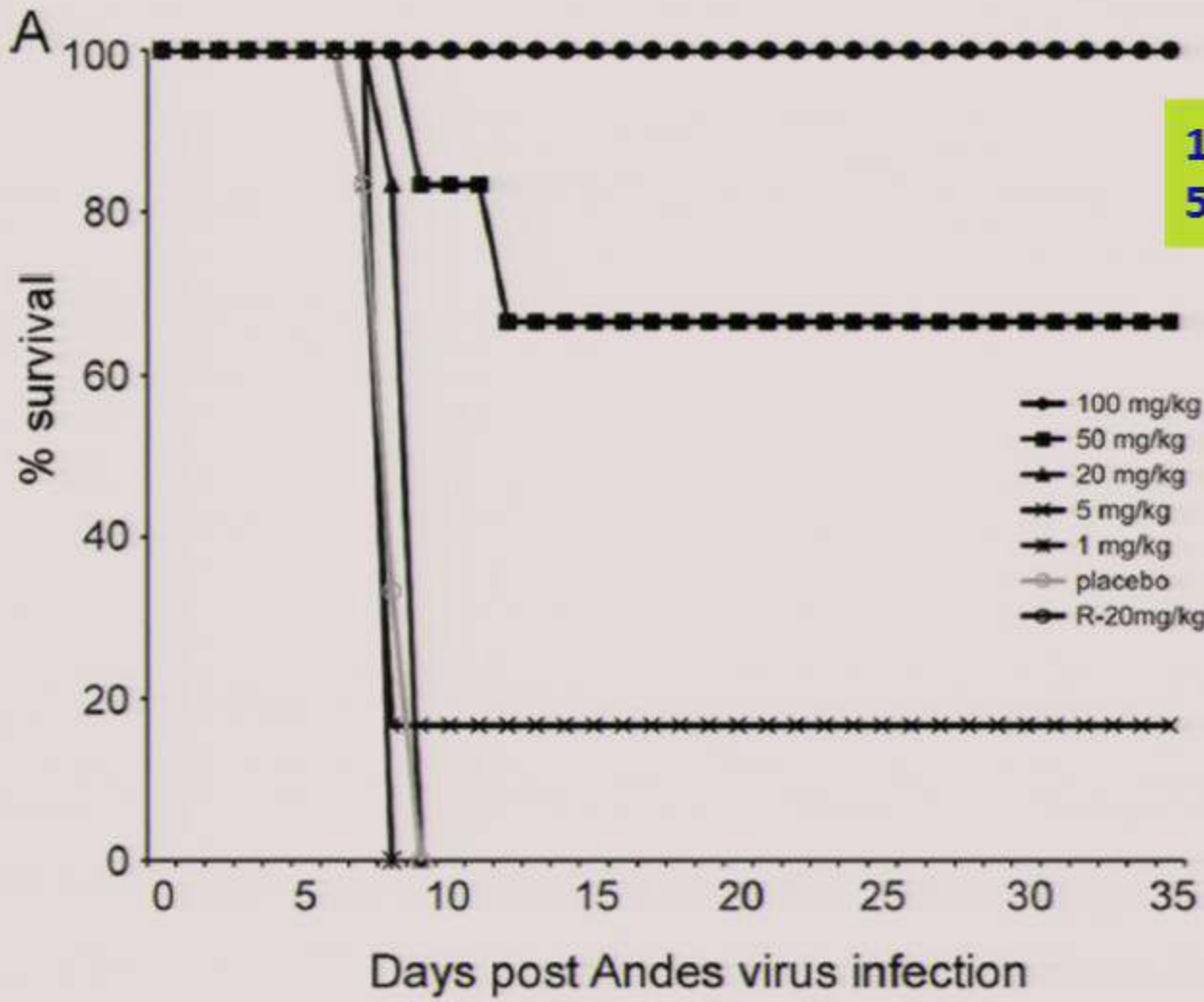
David Safronetz,^a Darryl Falzarano,^a Dana P. Scott,^b Yousuke Furuta,^c Heinz Feldmann,^a Brian B. Gowen^d

Laboratory of Virology^a and Rocky Mountain Veterinary Branch, Division of Intramural Research,^b National Institute of Allergy and Infectious Diseases, National Institutes of Health, Hamilton, Montana, USA; T-705 Project, Toyama Chemical Company, Ltd., Tokyo, Japan^c; Department of Animal, Dairy, and Veterinary Sciences, Utah State University, Logan, Utah, USA^d

- 4-6 haftalık Suriye hamsteri.
- İntra-peritoneal yoldan
 - Andes virus (ANDV) veya
 - SinNombre virüs (SNV) inoküle ediliyor.
- Favipiravir virüs inokülasyonundan **1 gün sonra** oral yoldan (günde iki kez) **14 gün** süreyle veriliyor



ANDV – Favipiravir Sağ kalım

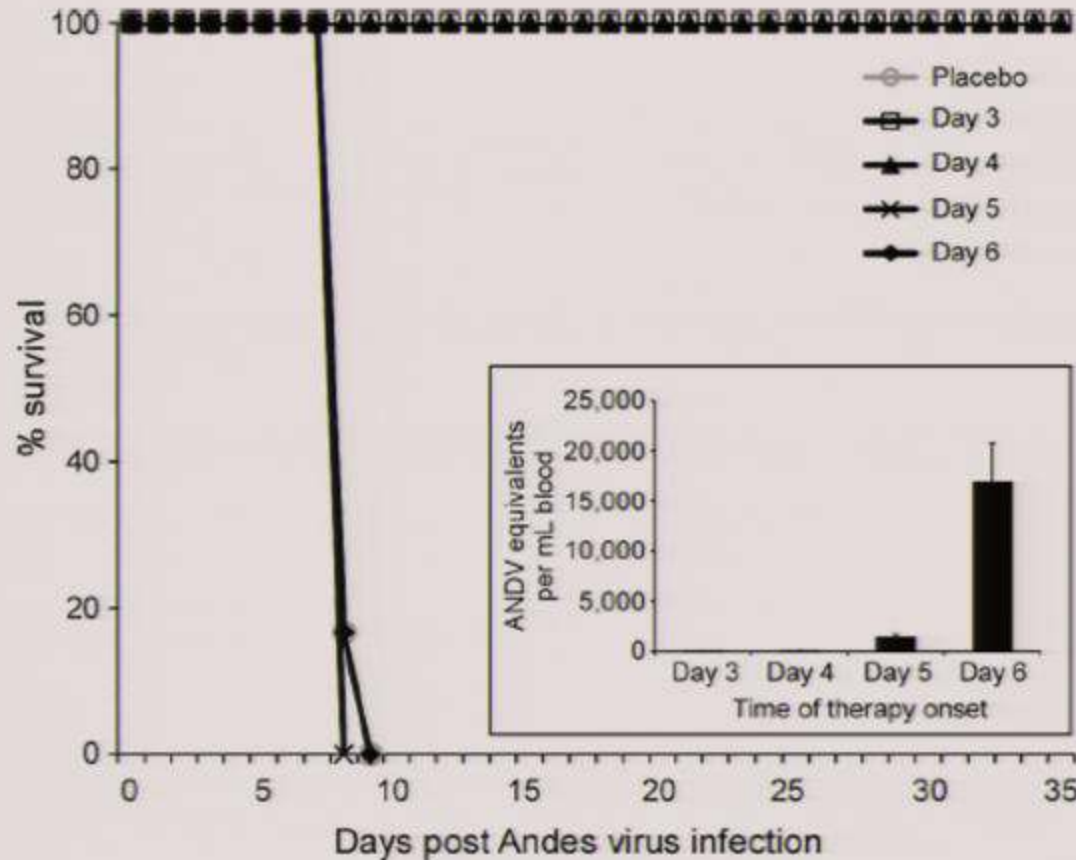


100 mg/kg: %100
50 mg/kg: % 66

İnokülasyondan 8 gün sonra kanda viral yük



“Geç” başlanan Favipiravir - ANDV



İlk 4 gün içerisinde başlanılan Favipiravir fataliteyi önlüyor.

İnokülasyondan 6-7 gün sonra solunum sıkıntısı, 9 gün sonra ölüm

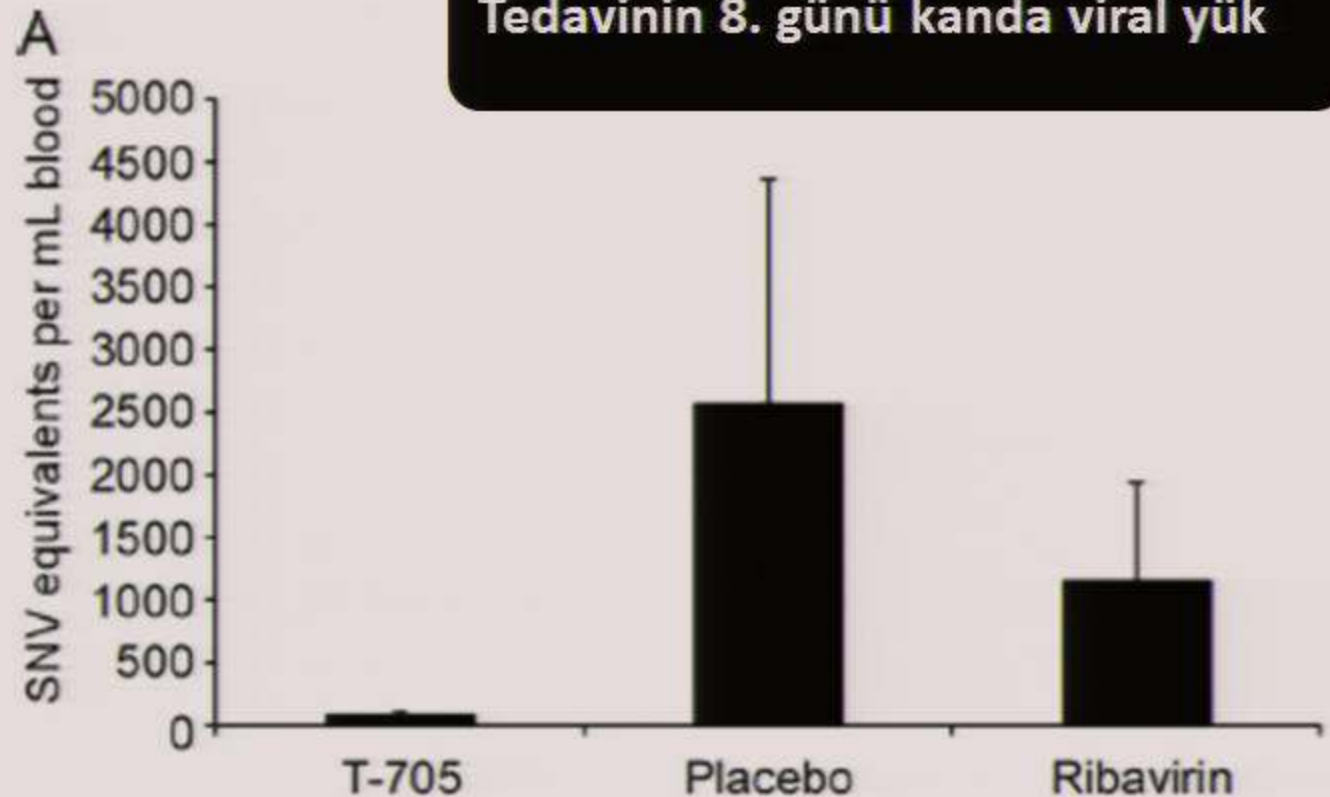
FIG 4 Efficacy of delayed T-705 therapy. Five groups of 6 hamsters were infected with a lethal dose of Andes virus. On each of days 3, 4, 5, and 6 postinfection, T-705 therapy (a 300-mg/kg loading dose, followed by daily treatment with 200 mg/kg) was initiated for a single group of animals. The fifth

SNV

Favipiravir X Ribavirin

- Favipiravir 100 mg/kg/gün
- Ribavirin 20 mg/kg/gün !!!

Tedavinin 8. günü kanda viral yük



Favipiravir (T-705)

- İnfluenza için geliştirilen bir ilaç.
- Kimyasal yapı .
 - *6-fluoro-3-hydroxy-2-pyrazinecarboxamide*
($C_5H_4FN_3O_2$)

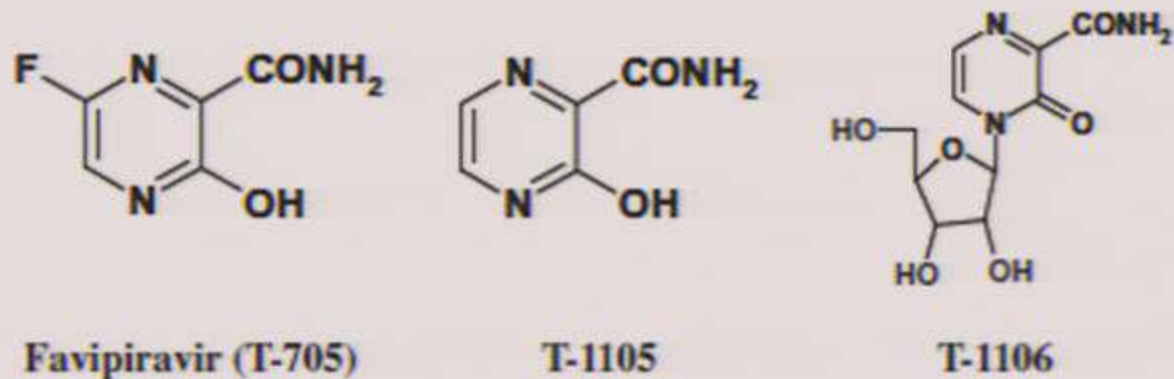


Fig. 1. Chemical structure of favipiravir (T-705), T-1105 and T-1106.

Favipiravir (T-705)

- **Etki mekanizması**
 - Viral RNA polimerazı inhibe eder
 - Konak hücredeki DNA veya RNA polimeraza etkili değil
- İnfluenza için dünya genelinde 1400 kişide denendi. Tolerans iyi.
- Japonya Faz 3 çalışması tamamlandı
- ABD'de Faz 2 çalışmaları sürüyor.

Favipiravir

- **In vitro etkili olduğu virüsler:**
 - *Junin virus,*
 - *Rift Valley Fever virus,*
 - *Yellow fever virus,*
 - *West Nile virus,*
 - *Western equine encephalitis virus,*
 - *Foot-and-mouth disease virus,*
 - *Norovirus*

Broad Spectrum Antiviral Activity of Favipiravir (T-705):
Protection from Highly Lethal Inhalational Rift Valley
Fever

Amy L. Caroline¹, Diana S. Powell¹, Laura M. Bethel¹, Tim D. Oury², Douglas S. Reed^{1,3},
Amy L. Hartman^{1,4*}





Teşekkürler...